

**PROCESS FOR HIGH THROUGHPUT SCREENING OF CpG-BASED
IMMUNO-AGONIST/ANTAGONIST**

Related Applications

This invention claims benefit of U.S. Provisional Application 60/233,035, filed September 15, 2000; U.S. Provisional Application 60/263,657, filed January 23, 2001; U.S. Provisional Application 60/291,726, filed May 17, 2001; and U.S. Provisional Application 60/300,210, filed June 22, 2001.

Field of the Invention

The invention pertains to signal transduction by immunostimulatory nucleic acids.

Background of the Invention

Bacterial DNA is a potent immunomodulatory substance. Yamamoto S et al., *Microbiol Immunol* 36:983-997 (1992). It has been hypothesized to be a pathogen-derived ligand recognized by an unidentified pathogen recognition receptor that initiates a host of innate and adaptive immune responses. Wagner H, *Adv Immunol* 73:329-368 (1999). CpG motif-containing oligodeoxynucleotides (CpG ODN) can mimic the biology of bacterial DNA. Krieg AM et al., *Nature* 374:546-549 (1995). CpG ODN and DNA vectors have recently been shown to be of clinical value due to immunostimulatory, hematopoietic and adjuvant qualities.

The adaptive immune system appeared approximately 450 million years ago when a transposon that carried the forerunners of the recombinase activating genes, RAG-1 and RAG-2, was inserted into the germ line of early jawed vertebrates. Agarwal A. et al., *Nature* 394:744 (1998). The ability to mount an adaptive immune response allowed organisms to remember the pathogens that they had already encountered, and natural selection made the adaptive immune response a virtually universal characteristic of vertebrates. However, this did not lead to discarding the previous form of host defense, the innate immune system. Indeed, this earlier form of host defense has been coopted to serve a second function, stimulating and orienting the primary adaptive immune response by controlling the expression of costimulatory molecules.

It had been surmised for a decade that cells of the innate immune system bear receptors for conserved molecular patterns associated with microbial pathogens. According to this model, when the protein antigens derived from pathogens are processed and presented as peptides that serve as the stimulus for specific T cell receptors, pattern recognition receptors (PRRs) on the antigen-presenting cells also induce the synthesis of costimulatory molecules, cytokines, and chemokines. These activated antigen-presenting cells serve to attract and activate the antigen-specific T cells that are essential to all adaptive immune responses. Janeway CAJ, *Cold Spring Harbor Symp Quant Biol* 54:1 (1989); Fearon DT et al., *Science* 272:50 (1996); and Medzhitov R et al., *Cell* 91:295 (1997). It was known that the substances that can induce costimulation include bacterial lipopolysaccharide (LPS), synthetic double-stranded RNA, glycans, and mannans. Furthermore, experimental evidence indicated that the processed antigen ligand for the T cell had to be on the same cell as the costimulatory molecule. This is obviously of crucial importance for maintaining self-tolerance; bystander presentation of costimulatory molecules would mean that tolerance would be lost whenever an infection occurred.

To validate this model, it was necessary to identify receptors for microbial patterns that, upon binding pathogen ligands, initiate signaling cascades leading to the production of costimulatory molecules and cytokines. Molecules such as mannose binding protein (MBP) do not qualify for this role, because they activate proteolytic cascades or promote phagocytosis but are not known to induce costimulation. The break-through came with the identification of a human homologue of *Drosophila* Toll initially cloned as a cDNA and later named hTLR4 (for human Toll-like receptor). Medzhitov R et al., *Nature* 388:394 (1997); Rock FL et al., *Proc Natl Acad Sci USA* 95:588 (1998); Chaudhary PM et al., *Blood* 91:4020-4027 (1998).

Toll-like receptors (TLRs) are a family of germline-encoded transmembrane proteins that facilitate pathogen recognition and activation of the innate immune system. Hoffmann JA et al., *Science* 284, 1313-1318 (1999); Rock FL et al., *Proc Natl Acad Sci USA* 95:588-593 (1998). TLRs engage conserved pathogen-derived ligands and subsequently activate the TLR/IL-1R signal transduction pathway to induce a variety of effector genes. Medzhitov R et al., *Mol Cell* 2:253-258 (1998); Muzio M et al., *J Exp Med* 187:2097-2101 (1998).

So far, ten different mammalian TLRs have been described. Rock FL et al., *Proc Natl*

Acad Sci USA 95:588-593 (1998); Chaudhary PM et al., *Blood* 91:4020-4027 (1998); Takeuchi O et al., *Gene* 231:59-65 (1999); Aderem A. et al., *Nature* 406:782-7 (2000). So far, genetic data suggest that the TLRs have unique functions and are not redundant. Ligands for and the function of most of these TLRs, aside from TLR2 and TLR4, remain to be elucidated.

It turns out that an LPS-binding and signaling receptor complex is assembled when hTLR4 interacts with LPS bound to CD14, a peripheral membrane protein held to the cell surface by a glycosyl-phosphoinositol tail. The presence of LPS binding protein (LBP) further increases signaling. The hTLR4 protein has a leucine-rich repeat sequence in its extracellular domain that interacts with CD14 complexed with LPS. TLR4 then transduces the LPS signal across the membrane because destructive mutation of this gene lead to an LPS-unresponsive state in mice, which are also deficient in the clearance of Gram-negative bacteria. Poltorak A et al., *Science* 282:2085 (1998); Qureshi ST et al., *J Exp Med* 189:615-625 (1999); Eden CS et al., *J Immunol* 140:180 (1988). It has since become apparent that humans, like flies, have numerous Toll-like receptors (TLRs).

TLR4 and other TLRs have a cytoplasmic Toll/IL-1 receptor (TIR) homology domain. This domain communicates with a similar domain on an adapter protein (MyD88) that interacts with TLR4 by means of a like:like interaction of TIR domains. The next interaction is between the adapter and a kinase, through their respective "death domains." The kinase in turn interacts with tumor necrosis factor (TNF) receptor-associated factor-6 (TRAF6). Medzhitov R et al., *Mol Cell* 2:253 (1998); Kopp EB et al., *Curr Opin Immunol* 11:15 (1999). After TRAF6, two sequential kinase activation steps lead to phosphorylation of the inhibitory protein I κ B and its dissociation from NF- κ B. The first kinase is a mitogen-activated kinase kinase kinase (MAPKKK) known as NIK, for NF- κ B-inducing kinase. The target of this kinase is another kinase made up of two chains, called I κ B kinase α (IKK α) and I κ B kinase β (IKK β), that together form a heterodimer of IKK α :IKK β , which phosphorylates I κ B. NF- κ B translocates to the nucleus to activate genes with κ B binding sites in their promoters and enhancers such as the genes encoding interleukin-1 β (IL-1 β), IL-6, IL-8, the p40 protein of IL-12, and the costimulatory molecules CD80 and CD86.

The types of cells that respond to CpG DNA - B cells, dendritic cells (DCs) and macrophages - are also stimulated by other pathogen-derived pattern-recognition factors, such

as LPS. In general, the PRRs of the innate immune system are situated on the cell surface, where they are probably best able to detect microbes. Although cell-surface proteins that bind DNA are well described, and have been proposed to mediate immune activation by CpG motif (Liang H et al., *J Clin Invest* 98:1119-1129 (1998)), this binding is sequence-independent and does not bring about cell activation. Krieg AM et al., *Nature* 374:546-549 (1995); Yamamoto T et al., *Microbiol Immunol* 38:831-836 (1994); Häcker H et al., *EMBO J* 17:6230-6240 (1998). Because CpG ODNs that have been immobilized to prevent cell uptake are nonstimulatory (Krieg AM et al., *Nature* 374:546-549 (1995); Manzel L et al., *Antisense Nucleic Acid Drug Dev* 9:459-464 (1999)), it appears that CpG ODN probably work by binding to an intracellular receptor. In support of this hypothesis, drugs such as chloroquine, which interfere with the endosomal acidification/processing of ODNs, specifically block the immune stimulatory effects of CpG DNA. Häcker H et al., *EMBO J* 17:6230-6240 (1998); Macfarlane DE et al., *J Immunol* 160:1122-1131 (1998); Yi AK et al., *J Immunol* 160:4755-4761 (1998). It has been proposed that an endosomal step is required for the CpG-induced signal transduction pathways. Häcker H et al., *EMBO J* 17:6230-6240 (1998); Yi AK et al., *J Immunol* 160:4755-4761 (1998). How the information contained in unmethylated CpG-motifs of bacterial DNA trigger changes in gene expression has not previously been discovered.

Since the receptor for bacterial DNA has been unknown, development of screening for optimal CpG motifs through direct binding analysis has been limited. An additional complication appears to be species-specific selectivity for CpG sequence, i.e., an optimal sequence for one species may not be optimal for another.

Summary of the Invention

Nucleic acids encoding three Toll-like receptors, Toll-like receptor 7 (TLR7), TLR8, and TLR9 of the mouse have now been identified, isolated, cloned and sequenced by the inventors. The invention in general provides isolated nucleic acid molecules encoding TLRs and isolated fragments of those nucleic acid molecules; isolated TLR polypeptides and isolated fragments of those polypeptides; expression vectors containing the foregoing nucleic acid molecules; host cells having the foregoing expression vectors; fusion proteins including the TLR polypeptides and fragments thereof; and screening methods useful for identifying,

comparing, and optimizing agents which interact with these TLRs, particularly agents that alter the expression of and signaling associated with these TLR molecules. In preferred embodiments the screening methods are high throughput screening methods.

The invention in some aspects arises from the surprising discovery that TLR9 is involved in immunostimulatory nucleic acid (ISNA)-induced immunostimulation. The invention also stems in part from the surprising discovery that TLR9 transduces immune activating signals in response to ISNA in a manner that is both sequence-specific and species-specific.

In a first aspect the invention provides isolated nucleic acid molecules which encode full-length murine TLR9. According to this aspect of the invention, isolated nucleic acid molecules are provided which are selected from the group consisting of (a) nucleic acid molecules which hybridize under stringent conditions to a nucleic acid molecule having a nucleotide sequence set forth as SEQ ID NO:1, and which code for a murine TLR9 having an amino acid sequence set forth as SEQ ID NO:3; (b) nucleic acid molecules that differ from the nucleic acid molecules of (a) in codon sequence due to degeneracy of the genetic code; and (c) complements of (a) or (b). In a certain embodiment, the isolated nucleic acid molecule codes for SEQ ID NO:3, where SEQ ID NO:3 represents the deduced amino acid sequence of full-length murine TLR9. In some embodiments the isolated nucleic acid molecule comprises the nucleotide sequence of SEQ ID NO:1 or SEQ ID NO:2, where these correspond to full-length cDNA and the open reading frame for murine TLR9, respectively.

The term "stringent conditions" as used herein refers to combined conditions based on parameters including salt, temperature, organic solvents, and optionally other factors with which the practitioner skilled in the art is familiar. Nucleic acid hybridization parameters may be found in references which compile such methods, e.g., *Molecular Cloning: A Laboratory Manual*, J. Sambrook, et al., eds., Second Edition, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York, 1989, or *Current Protocols in Molecular Biology*, F.M. Ausubel, et al., eds., John Wiley & Sons, Inc., New York. More specifically, stringent conditions, as used herein, refers, for example, to hybridization at 65°C in hybridization buffer (3.5 x SSC, 0.02% Ficoll, 0.02% polyvinyl pyrrolidone, 0.02% bovine serum albumin, 2.5mM NaH₂PO₄ (pH7), 0.5% SDS, 2mM EDTA). SSC is 0.15M sodium chloride/0.15M sodium citrate, pH7; SDS is sodium dodecyl sulfate; and EDTA is ethylenediaminetetraacetic

acid. After hybridization, the membrane upon which the DNA is transferred is washed with 2 x SSC at room temperature and then with 0.1 - 0.5 x SSC/0.1 x SDS at temperatures up to 68°C. There are other conditions, reagents, and so forth which can be used, which result in a similar degree of stringency. The skilled artisan will be familiar with such conditions, and thus they are not given here. It will be understood, however, that the skilled artisan will be able to manipulate the conditions in a manner to permit the clear identification of alleles of murine TLR nucleic acids of the invention. The skilled artisan also is familiar with the methodology for screening cells and libraries for expression of such molecules which then are routinely isolated, followed by isolation of the pertinent nucleic acid molecule and sequencing.

The invention in a second aspect provides isolated TLR9 polypeptides or fragments thereof. The isolated TLR9 polypeptides or fragments thereof include at least one amino acid of a murine TLR9 selected from the group consisting of amino acids 2, 3, 4, 6, 7, 18, 19, 22, 38, 44, 55, 58, 61, 62, 63, 65, 67, 71, 80, 84, 87, 88, 91, 101, 106, 109, 117, 122, 123, 134, 136, 140, 143, 146, 147, 157, 160, 161, 167, 168, 171, 185, 186, 188, 189, 191, 199, 213, 217, 220, 227, 231, 236, 245, 266, 269, 270, 271, 272, 273, 274, 278, 281, 285, 297, 298, 301, 305, 308, 311, 322, 323, 325, 326, 328, 332, 335, 346, 348, 353, 355, 358, 361, 362, 365, 367, 370, 372, 380, 381, 382, 386, 389, 392, 394, 397, 409, 412, 413, 415, 416, 419, 430, 432, 434, 435, 438, 439, 443, 444, 446, 447, 448, 450, 451, 452, 454, 455, 459, 460, 463, 465, 466, 468, 469, 470, 472, 473, 474, 475, 478, 488, 489, 494, 495, 498, 503, 508, 510, 523, 531, 539, 540, 543, 547, 549, 561, 563, 565, 576, 577, 579, 580, 587, 590, 591, 594, 595, 597, 599, 601, 603, 610, 611, 613, 616, 619, 632, 633, 640, 643, 645, 648, 650, 657, 658, 660, 667, 670, 672, 675, 679, 689, 697, 700, 703, 705, 706, 711, 715, 716, 718, 720, 723, 724, 726, 729, 731, 735, 737, 743, 749, 750, 751, 752, 754, 755, 759, 760, 772, 774, 780, 781, 786, 787, 788, 800, 814, 821, 829, 831, 832, 835, 844, 857, 858, 859, 862, 864, 865, 866, 879, 893, 894, 898, 902, 910, 917, and 927 of SEQ ID NO:3, wherein the TLR9 polypeptide or fragment thereof has an amino acid sequence which is identical to a human TLR9 polypeptide or fragment thereof except for the at least one amino acid of murine TLR9. The TLR9 polypeptide or fragment thereof in certain embodiments according to this aspect of the invention further includes at least one amino acid of murine TLR9 selected from the group consisting of amino acids 949, 972, 975, 976, 994, 997, 1000, 1003, 1004, 1010,

1011, 1018, 1023, and 1027 of SEQ ID NO:3. Thus specifically excluded from this aspect of the invention are TLR9 fragments restricted to the C-terminal 95 amino acids and fragments thereof.

In certain embodiments the TLR9 polypeptide and fragments thereof according to this aspect of the invention exclude those TLR9 polypeptides and fragments thereof which differ from human TLR9 and fragments thereof only by one or more conservative amino acid substitutions at particular sites noted above. As is well known in the art, a "conservative amino acid substitution" refers to an amino acid substitution which generally does not alter the relative charge or size characteristics of the polypeptide in which the amino acid substitution is made. Conservative substitutions of amino acids typically include substitutions made amongst amino acids within the following groups: methionine (M), isoleucine (I), leucine (L), valine (V); phenylalanine (F), tyrosine (Y), tryptophan (W); lysine (K), arginine (R), histidine (H); alanine (A), glycine (G); serine (S), threonine (T); glutamine (Q), asparagine (N); and glutamic acid (E), aspartic acid (D).

According to this and other aspects of the invention, with reference to TLR "polypeptides and fragments thereof," "fragments thereof" refers to polypeptide fragments having stretches of contiguous amino acid residues that are at least about 8 amino acids long. Generally the fragments are at least about 10 amino acids long; more generally at least 12 amino acids long; often at least about 14 amino acids long; more often at least about 16 amino acids long; typically at least 18 amino acids long; more typically at least 20 amino acids long; usually at least 22 amino acids long; and more usually at least 24 amino acids long. Certain preferred embodiments include larger fragments that are, for example, at least about 30 amino acids long, at least about 40 amino acids long, at least about 50 amino acids long, at least about 100 amino acids long, at least about 200 amino acids long, and so on, up to and including fragments that are a single amino acid shorter than full-length TLR polypeptide.

In certain embodiments, the human TLR9 has an amino acid sequence set forth as SEQ ID NO:6.

In preferred embodiments, the isolated TLR9 polypeptides or fragments thereof include an amino acid sequence selected from the group consisting of SEQ ID NO:3 and fragments of SEQ ID NO:3. In some embodiments according to this aspect of the invention,

the isolated TLR9 polypeptides or fragments thereof include combinations of the foregoing human and murine TLR9 polypeptides.

In certain preferred embodiments the isolated TLR9 polypeptide or fragment thereof is an extracytoplasmic domain (also referred to herein as extracellular domain) of TLR9, or a portion thereof. As described in greater detail further herein, TLR7, TLR8, and TLR9 have certain structural and functional domains. Structural domains of these TLRs include but are not limited to an extracytoplasmic domain, a transmembrane domain, and a cytoplasmic domain. The extracytoplasmic domain extends into the lumen of endosomal/lysosomal vesicles. The cytoplasmic domain includes a Toll/interleukin-1 receptor-like domain (also referred to as Toll/IL-1R domain, TIR homology domain, or TIR domain). In murine TLR9 the extracytoplasmic, transmembrane, and cytoplasmic domains correspond to amino acids 1 to about 819, about 820 to about 837, and about 838 to about 1032, respectively.

As mentioned above, it has been discovered according to the invention that TLR9 is involved in immune activation induced by certain nucleic acid molecules referred to in the art as immunostimulatory nucleic acids (ISNAs), including CpG nucleic acids. It is believed by the inventors that binding of ISNA to TLR9 leads to signal transduction involving the TIR domain of TLR9. Thus in certain embodiments according to this aspect of the invention, the isolated TLR9 polypeptide or fragment thereof selectively binds to an ISNA, including an ISNA that is a CpG nucleic acid.

Also included according to this aspect of the invention are isolated TLR9 polypeptides or fragments thereof which are portions of the extracytoplasmic domain believed by the inventors to interact with immunostimulatory nucleic acids such as CpG nucleic acids. In certain embodiments such portions include an MBD motif set forth as any one of SEQ ID NOs: 126, 127, 210, and 211. In certain embodiments portions of the extracytoplasmic domain believed by the inventors to interact with immunostimulatory nucleic acids include a CXXC motif set forth as any one of SEQ ID NOs: 196, 197, and 198.

According to a third aspect of the invention, isolated nucleic acid molecules are provided which encode the foregoing isolated TLR9 polypeptides or fragments thereof. The isolated nucleic acid molecules according to this aspect of the invention specifically exclude certain expressed sequence tags (ESTs) identified by the following GenBank accession numbers: AA162495, AA197442, AA273731, AA794083, AA915125, AA968074,

AI428529, AI451215, AI463056, AI893951, AV142833, AV326033, AV353853,
AW048117, AW048548, AW215685, AW549817, BB179985, BB215203, BB283380,
BB285606, BB312895, BB497196, BB622397, BF016670, BF150116, BF161011,
BF300296, BF385702, BF539367, BF784415, BG863184, BG922959, BG967012,
5 BG974917, BI105291, BI153921, BI651868, BI653892, and W76964.

In a fourth aspect the invention provides isolated nucleic acid molecules which encode full-length murine TLR7. According to this aspect of the invention, isolated nucleic acid molecules are provided which are selected from the group consisting of (a) nucleic acid molecules which hybridize under stringent conditions to a nucleic acid molecule having a nucleotide sequence set forth as SEQ ID NO:173, and which code for a murine TLR7 having an amino acid sequence set forth as SEQ ID NO:175; (b) nucleic acid molecules that differ from the nucleic acid molecules of (a) in codon sequence due to degeneracy of the genetic code; and (c) complements of (a) or (b). In a certain embodiment, the isolated nucleic acid molecule codes for SEQ ID NO:175, where SEQ ID NO:175 represents the deduced amino acid sequence of full-length murine TLR7. In some embodiments the isolated nucleic acid molecule comprises the nucleotide sequence of SEQ ID NO:173 or SEQ ID NO:174, where these correspond to full-length cDNA and the open reading frame for murine TLR7, respectively.

The invention in a fifth aspect provides isolated TLR7 polypeptides or fragments thereof. The isolated TLR7 polypeptides or fragments thereof include at least one amino acid of a murine TLR7 selected from the group consisting of amino acids 4, 8, 15, 16, 18, 21, 23, 24, 25, 27, 37, 39, 40, 41, 42, 44, 45, 61, 79, 83, 86, 89, 92, 96, 103, 109, 111, 113, 119, 121, 127, 128, 131, 145, 148, 151, 164, 172, 176, 190, 202, 203, 204, 205, 222, 225, 226, 228, 236, 238, 243, 250, 253, 266, 268, 271, 274, 282, 283, 287, 288, 308, 313, 314, 315, 325, 328, 331, 332, 341, 343, 344, 347, 351, 357, 360, 361, 362, 363, 364, 365, 366, 370, 371, 377, 378, 387, 388, 389, 392, 397, 398, 413, 415, 416, 419, 421, 422, 425, 437, 438, 440, 446, 449, 453, 454, 455, 456, 462, 470, 482, 486, 487, 488, 490, 491, 493, 494, 503, 505, 509, 511, 529, 531, 539, 540, 543, 559, 567, 568, 574, 583, 595, 597, 598, 600, 611, 613, 620, 624, 638, 645, 646, 651, 652, 655, 660, 664, 665, 668, 669, 672, 692, 694, 695, 698, 701, 704, 714, 720, 724, 727, 728, 733, 738, 745, 748, 755, 762, 777, 780, 789, 803, 846, 850, 851, 860, 864, 868, 873, 875, 884, 886, 888, 889, 890, 902, 903, 911, 960, 967, 970,

980, 996, 1010, 1018, 1035, and 1045 of SEQ ID NO:175, wherein the TLR7 polypeptide or fragment thereof has an amino acid sequence which is identical to a human TLR7 polypeptide or fragment thereof except for the at least one amino acid of murine TLR7.

In certain embodiments the TLR7 polypeptide and fragments thereof according to this aspect of the invention exclude those TLR7 polypeptides and fragments thereof which differ from human TLR7 and fragments thereof only by one or more conservative amino acid substitutions at particular sites noted above.

In certain embodiments, the human TLR7 has an amino acid sequence set forth as SEQ ID NO:170.

In preferred embodiments, the isolated TLR7 polypeptides or fragments thereof include an amino acid sequence selected from the group consisting of SEQ ID NO:175 and fragments of SEQ ID NO:175. In some embodiments according to this aspect of the invention, the isolated TLR7 polypeptides or fragments thereof include combinations of the foregoing human and murine TLR7 polypeptides.

In certain preferred embodiments the isolated TLR7 polypeptide or fragment thereof is an extracytoplasmic domain of TLR7, or a portion thereof. In certain embodiments according to this aspect of the invention, the isolated TLR7 polypeptide or fragment thereof selectively binds to an ISNA, including an ISNA that is a CpG nucleic acid. Also included according to this aspect of the invention are isolated TLR7 polypeptides or fragments thereof which are portions of the extracytoplasmic domain believed by the inventors to interact with immunostimulatory nucleic acids such as CpG nucleic acids. In certain embodiments such portions include an MBD motif set forth as any one of SEQ ID NOs: 203, 204, 212, and 213.

In certain embodiments portions of the extracytoplasmic domain believed by the inventors to interact with immunostimulatory nucleic acids include a CXXC motif set forth as any one of SEQ ID NOs: 196, 199, and 200.

According to a sixth aspect of the invention, isolated nucleic acid molecules are provided which encode the foregoing isolated TLR7 polypeptides or fragments thereof. The isolated nucleic acid molecules according to this aspect of the invention specifically exclude certain ESTs identified by the following GenBank accession numbers: AA176010, AA210352, AA241310, AA266000, AA266744, AA276879, AA288480, AA871870, AI119722, AI449297, AI466859, AI604175, AV322307, BB033376, BB116163, BB210788,

BB464985, BB466708, BB636153, BF101884, BF124798, BF143871, BG067922, BG080980, BG082140, BG871070, BG964747, BG976560, BI150306, BI411471, and C87987.

In a seventh aspect the invention provides isolated nucleic acid molecules which encode full-length murine TLR8. According to this aspect of the invention, isolated nucleic acid molecules are provided which are selected from the group consisting of (a) nucleic acid molecules which hybridize under stringent conditions to a nucleic acid molecule having a nucleotide sequence set forth as SEQ ID NO:190, and which code for a murine TLR8 having an amino acid sequence set forth as SEQ ID NO:192; (b) nucleic acid molecules that differ from the nucleic acid molecules of (a) in codon sequence due to degeneracy of the genetic code; and (c) complements of (a) or (b). In a certain embodiment, the isolated nucleic acid molecule codes for SEQ ID NO:192, where SEQ ID NO:192 represents the deduced amino acid sequence of full-length murine TLR8. In some embodiments the isolated nucleic acid molecule comprises the nucleotide sequence of SEQ ID NO:190 or SEQ ID NO:191, where these correspond to full-length cDNA and the open reading frame for murine TLR8, respectively.

The invention in an eighth aspect provides isolated TLR8 polypeptides or fragments thereof. The isolated TLR8 polypeptides or fragments thereof include at least one amino acid of a murine TLR8 selected from the group consisting of amino acids 5, 6, 9, 10, 14, 15, 18, 21, 22, 23, 24, 25, 26, 27, 28, 30, 39, 40, 41, 43, 44, 50, 51, 53, 55, 61, 67, 68, 74, 80, 85, 93, 98, 99, 100, 104, 105, 106, 107, 110, 114, 117, 119, 121, 124, 125, 134, 135, 138, 145, 155, 156, 157, 160, 161, 162, 163, 164, 166, 169, 170, 174, 180, 182, 183, 186, 187, 191, 193, 194, 196, 197, 199, 200, 207, 209, 210, 227, 228, 230, 231, 233, 234, 241, 256, 263, 266, 267, 268, 269, 272, 274, 275, 276, 280, 285, 296, 298, 299, 300, 303, 305, 306, 307, 310, 312, 320, 330, 333, 335, 343, 344, 345, 346, 347, 349, 351, 356, 362, 365, 366, 375, 378, 379, 380, 381, 383, 384, 386, 387, 392, 402, 403, 408, 414, 416, 417, 422, 426, 427, 428, 429, 430, 431, 433, 437, 438, 439, 440, 441, 444, 445, 449, 456, 461, 463, 471, 483, 486, 489, 490, 494, 495, 496, 505, 507, 509, 512, 513, 519, 520, 523, 537, 538, 539, 541, 542, 543, 545, 554, 556, 560, 567, 569, 574, 575, 578, 586, 592, 593, 594, 595, 597, 599, 602, 613, 617, 618, 620, 621, 623, 628, 630, 633, 639, 641, 643, 644, 648, 655, 658, 661, 663, 664, 666, 668, 677, 680, 682, 687, 688, 690, 692, 695, 696, 697, 700, 702, 703, 706, 714,

715, 726, 727, 728, 730, 736, 738, 739, 741, 746, 748, 751, 752, 754, 757, 764, 766, 772, 776, 778, 781, 784, 785, 788, 791, 795, 796, 801, 802, 806, 809, 817, 820, 821, 825, 828, 829, 831, 839, 852, 853, 855, 858, 863, 864, 900, 903, 911, 918, 934, 977, 997, 1003, 1008, 1010, 1022, 1023, 1024, 1026, and 1030 of SEQ ID NO:192, wherein the TLR8 polypeptide or fragment thereof has an amino acid sequence which is identical to a human TLR8 polypeptide or fragment thereof except for the at least one amino acid of murine TLR8.

In certain embodiments the TLR8 polypeptide and fragments thereof according to this aspect of the invention exclude those TLR8 polypeptides and fragments thereof which differ from human TLR8 and fragments thereof only by one or more conservative amino acid substitutions at particular sites noted above.

In certain embodiments, the human TLR8 has an amino acid sequence set forth as SEQ ID NO:184.

In preferred embodiments, the isolated TLR8 polypeptides or fragments thereof include an amino acid sequence selected from the group consisting of SEQ ID NO:192 and fragments of SEQ ID NO:192. In some embodiments according to this aspect of the invention, the isolated TLR8 polypeptides or fragments thereof include combinations of the foregoing human and murine TLR8 polypeptides.

In certain preferred embodiments the isolated TLR8 polypeptide or fragment thereof is an extracytoplasmic domain of TLR8, or a portion thereof. In certain embodiments according to this aspect of the invention, the isolated TLR8 polypeptide or fragment thereof selectively binds to an ISNA, including an ISNA that is a CpG nucleic acid. Also included according to this aspect of the invention are isolated TLR8 polypeptides or fragments thereof which are portions of the extracytoplasmic domain believed by the inventors to interact with immunostimulatory nucleic acids such as CpG nucleic acids. In certain embodiments such portions include an MBD motif set forth as any one of SEQ ID NOs: 205, 206, 214, and 215. In certain embodiments portions of the extracytoplasmic domain believed by the inventors to interact with immunostimulatory nucleic acids include a CXXC motif set forth as any one of SEQ ID NOs: 196, 201, and 202.

According to a ninth aspect of the invention, isolated nucleic acid molecules are provided which encode the foregoing isolated TLR8 polypeptides or fragments thereof. The isolated nucleic acid molecules according to this aspect of the invention specifically exclude

certain ESTs identified by the following GenBank accession numbers: AA116795, AA268605, AA920337, AI529457, AI849892, AV097766, AV117427, AV164719, AV169968, AW551677, BB143750, BB214171, BB243478, BB244318, BB254686, BB256660, BB258368, BB278984, BB291470, BB292008, BB364655, BB373674, BB428800, BB439876, BB444812, BB445724, BB465766, BB470182, BB535086, BB573907, BB573981, BB607650, BF135656, BF722808, BG299237, BG918020, BG919592, and W39977.

In a further aspect, the invention provides TLR expression vectors comprising the foregoing isolated nucleic acid molecules operably linked to a promoter. Thus in certain embodiments pertaining to TLR9, the expression vector includes an isolated nucleic acid molecule according to the first aspect or the third aspect of the invention, operably linked to a promoter. In other embodiments, relating to TLR7, the expression vector includes an isolated nucleic acid molecule according to the fourth aspect or the sixth aspect of the invention, operably linked to a promoter. In yet other embodiments, relating to TLR8, the expression vector includes an isolated nucleic acid molecule according to the seventh aspect or the ninth aspect of the invention, operably linked to a promoter.

The expression vectors according to this aspect of the invention are designed and constructed so that when they are introduced into a cell, under proper conditions they direct expression of the gene product encoded by the incorporated isolated nucleic acid molecule. For example, the promoter can be constitutively active or it can be inducible or repressible upon interaction with a suitable inducer or repressor compound.

According to another aspect, host cells are provided that include a TLR expression vector of the invention. While any suitable method can be used, an expression vector typically is introduced into a cell by transfection or transformation. The host cells transformed or transfected with the TLR expression vectors are in some embodiments co-transformed or co-transfected with another expression vector useful for the expression of another polypeptide. Alternatively, a host cell can be transformed or transfected with an expression vector capable of directing expression of a TLR polypeptide or fragment thereof of the invention and (i) at least one additional TLR polypeptide or fragment thereof, or (ii) at least one non-TLR polypeptide or fragment thereof. In certain preferred embodiments, the host cell includes separate expression vectors for any combination of TLR7, TLR8, and

TLR9. In some embodiments, a co-transformed or co-transfected expression vector may be useful for detection or regulation of TLR expression or TLR-related signaling. Specifically, in certain preferred embodiments the host cell includes an expression vector providing a reporter construct capable of interacting with a TIR domain.

5 In another aspect, the invention provides agents which selectively bind the isolated TLR polypeptides and fragments thereof of the invention. In certain embodiments the agent does not bind a human TLR polypeptide or fragment thereof, wherein the human TLR is selected from human TLR7, TLR8, and TLR9. In certain embodiments the agent is a polypeptide, preferably one selected from the group consisting of monoclonal antibodies, polyclonal antibodies, Fab antibody fragments, F(ab')₂ antibody fragments, Fv antibody fragments, antibody fragments including a CDR3 region, and fusion proteins and other polypeptides including any such antibodies or antibody fragments.

10 Also provided are agents which selectively bind the foregoing isolated nucleic acid molecules, preferably antisense nucleic acid molecules which selectively bind to any of the foregoing isolated nucleic acid molecules encoding a TLR polypeptide or fragment thereof. In some embodiments the agent is an isolated nucleic acid molecule which hybridizes under stringent conditions to an isolated nucleic acid molecule provided according to any of the first, third, fourth, fifth, sixth, and eighth aspects of the invention. In certain preferred embodiments the agent is an isolated nucleic acid molecule having a nucleotide sequence which is complementary to an isolated nucleic acid molecule provided according to any of the first, third, fourth, fifth, sixth, and eighth aspects of the invention.

15 In still other aspects of the invention, methods for inhibiting TLR expression and TLR signaling in a cell are provided. The methods include contacting the cell with an amount of an agent effective to inhibit TLR expression and TLR signaling in the cell, wherein the TLR is selected from the group consisting of TLR7, TLR8, and TLR9. In some embodiments the agent brought into contact with the cell is selected from the group consisting of monoclonal antibodies, polyclonal antibodies, Fab antibody fragments, F(ab')₂ antibody fragments, Fv antibody fragments, antibody fragments including a CDR3 region, and fusion proteins and other polypeptides that include any such antibodies or antibody fragments. In some 25 embodiments the cell is contacted with an antisense nucleic acid specific for the TLR, in an amount effective to inhibit TLR expression in the cell. In some embodiments the cell is 30

contacted with an agent such as a cytokine or small molecule, in an amount effective to inhibit TLR expression in the cell.

In yet another aspect the invention provides a method for identifying nucleic acid molecules which interact with a TLR polypeptide or a fragment thereof. The method involves contacting a TLR polypeptide selected from the group consisting of TLR7, TLR8, TLR9, and nucleic acid-binding fragments thereof with a test nucleic acid molecule; and measuring an interaction of the test nucleic acid molecule with the TLR polypeptide or fragment thereof. Nucleic acid-binding fragments of TLRs preferably include the extracytoplasmic domain or subportions thereof, such as those which include at least an MBD motif, a CXXC motif, or both an MBD motif and a CXXC motif.

In this and other aspects of the invention involving methods of use of TLR polypeptides and fragments thereof, in some embodiments the TLR polypeptide or fragment thereof is TLR7. Likewise in this and other aspects of the invention involving methods of use of TLR polypeptides and fragments thereof, in some embodiments the TLR polypeptide or fragment thereof is TLR8. Also in this and other aspects of the invention involving methods of use of TLR polypeptides and fragments thereof, in some embodiments the TLR polypeptide or fragment thereof is TLR9.

In this and other aspects of the invention involving methods of use of TLR polypeptides and fragments thereof, in some embodiments the TLR polypeptide or fragment thereof is expressed in a cell. The cell expressing the TLR polypeptide or fragment thereof may naturally express the TLR polypeptide or fragment thereof, or it may be a host cell as provided by other aspects of the instant invention.

In this and other aspects of the invention involving methods of use of TLR polypeptides and fragments thereof, in some embodiments the TLR polypeptide or fragment thereof is an isolated TLR polypeptide or fragment thereof. In certain preferred embodiments the isolated TLR polypeptide or fragment thereof is immobilized on a solid support, for example a multiwell plate, a slide, a BIAcore chip, a bead, a column, and the like. The immobilization can be accomplished by any chemical or physical method suitable for the purpose of the assay to be performed according to the method of the invention.

In certain embodiments the TLR polypeptide or fragment thereof is fused with an Fc fragment of an antibody. The Fc fragment portion of such a fusion molecule may be useful,

for example, for attaching the TLR polypeptide or fragment thereof to a substrate, or for providing a target for detecting the presence of the TLR polypeptide or fragment thereof. The Fc fragment can be selected from any suitable vertebrate species and will typically, but not necessarily, be derived from an antibody belonging to the IgG class of antibodies. For example, the Fc can be a human or a murine Fc γ . In certain embodiments the TLR polypeptide or fragment thereof is fused with an Fc fragment of an antibody with a specific cleavage site at or near the junction between the TLR polypeptide or fragment thereof and the Fc fragment. In one preferred embodiment the cleavage site is a thrombin protease recognition site. In a preferred embodiment the TLR polypeptide or fragment thereof fused with the Fc fragment includes a TLR extracytoplasmic domain.

In certain embodiments the interaction involving the TLR polypeptide or fragment thereof and the test nucleic acid molecule is binding between the TLR polypeptide or fragment thereof and the test nucleic acid molecule.

In certain embodiments according to this aspect of the invention, the measuring is accomplished by a method selected from the group consisting of enzyme-linked immunosorbent assay (ELISA), biomolecular interaction assay (BIA), electromobility shift assay (EMSA), radioimmunoassay (RIA), polyacrylamide gel electrophoresis (PAGE), and Western blotting.

In certain embodiments the measuring is accomplished by a method comprising measuring a response mediated by a TLR signal transduction pathway. For example, the response mediated by a TLR signal transduction pathway can be selected from the group consisting of induction of a gene under control of NF- κ B promoter and secretion of a cytokine. In certain preferred embodiments the gene under control of NF- κ B promoter is selected from the group consisting of IL-8, IL-12 p40, NF- κ B-luc, IL-12 p40-luc, and TNF-luc. In certain preferred embodiments the secreted cytokine is selected from the group consisting of IL-8, TNF- α , and IL-12 p40.

In another embodiment the method according to this aspect of the invention can be used to determine if the test nucleic acid molecule is an immunostimulatory nucleic acid. The method involves the additional steps of comparing (a) the response mediated by a TLR signal transduction pathway as measured in the presence of the test nucleic acid molecule with (b) a response mediated by a TLR signal transduction pathway as measured in the absence of the

test nucleic acid molecule; and determining the test nucleic acid molecule is an immunostimulatory nucleic acid when (a) exceeds (b).

In yet another embodiment the method according to this aspect of the invention can be used to determine if the response to the test nucleic acid molecule is stronger or weaker than a response to a reference nucleic acid molecule. The method involves the additional steps of comparing the response to a reference response when the TLR polypeptide is independently contacted with a reference nucleic acid molecule; and determining if the response is stronger or weaker than the reference response. In this embodiment the test nucleic acid molecule and the reference nucleic acid molecule are not able to compete or interact. For example, the reference response can be a parallel control or a historical control.

In another embodiment the method involves the additional steps of comparing the response to a reference response when the TLR polypeptide is concurrently contacted with a reference nucleic acid molecule; and determining if the response is stronger or weaker than the reference response. In this embodiment the test nucleic acid molecule and the reference nucleic acid molecule are potentially able to compete or interact since they are both present, for example, in a single reaction.

In another aspect the invention provides a screening method for identifying an immunostimulatory nucleic acid. The method according to this aspect involves contacting a functional TLR selected from the group consisting of TLR7, TLR8, and TLR9 with a test nucleic acid molecule; detecting presence or absence of a response mediated by a TLR signal transduction pathway in the presence of the test nucleic acid molecule arising as a result of an interaction between the functional TLR and the test nucleic acid molecule; and determining the test nucleic acid molecule is an ISNA when the presence of a response mediated by the TLR signal transduction pathway is detected. A functional TLR refers to a TLR polypeptide or fragment thereof that can bind with a ligand and as a consequence of the binding engage at least one step or additional polypeptide in a TLR signal transduction pathway.

In one embodiment the method according to this aspect of the invention includes the further step of comparing (a) the response mediated by the TLR signal transduction pathway arising as a result of an interaction between the functional TLR and the test nucleic acid molecule with (b) a response arising as a result of an interaction between the functional TLR and a reference ISNA. In this and other screening assays of the instant invention, in preferred

embodiments the screening method is performed on a plurality of test nucleic acids. In certain preferred embodiments the response mediated by the TLR signal transduction pathway is measured quantitatively, and the response mediated by the TLR signal transduction pathway associated with each of the plurality of test nucleic acid molecules is compared with a response arising as a result of an interaction between the functional TLR and a reference ISNA.

In certain preferred embodiments a subset of the plurality of test nucleic acid molecules is selected based on the ability of the subset to produce a specific response mediated by the TLR signal transduction pathway. For example, the specific response can be induction of a specific cytokine or panel of cytokines, e.g., Th1 cytokines, or, alternatively, inhibition of a specific cytokine or panel of cytokines, e.g., Th2 cytokines. The specific response can be induction, or, alternatively, inhibition of a specific class or subclass of antibody or panel of classes or subclasses of antibodies, e.g., Th1-associated antibodies or Th2-associated antibodies. The specific response in some embodiments can be activation or inhibition of certain types of immune cells, e.g., B cells, dendritic cells (DCs), and natural killer (NK) cells. In some embodiments the specific response can be induction or inhibition of proliferation of certain types of immune cells, e.g., B cells, T cells, NK cells, dendritic cells, monocytes/macrophages. The subset of the plurality of test nucleic acids is therefore selected on the basis of the common association between the test nucleic acids of the subset and the particular type of response mediated by the TLR signal transduction pathway. The particular type of response mediated by the TLR signal transduction pathway is typically, but not necessarily, an immune cell response.

In certain embodiments the response mediated by a TLR signal transduction pathway is selected from the group consisting of induction of a gene under control of NF- κ B promoter and secretion of a cytokine. In certain preferred embodiments the gene under control of NF- κ B promoter is selected from the group consisting of IL-8, IL-12 p40, NF- κ B-luc, IL-12 p40-luc, and TNF-luc. In certain preferred embodiments the cytokine is selected from the group consisting of IL-8, TNF- α , and IL-12 p40.

In certain preferred embodiments the reference ISNA is a CpG nucleic acid.

In certain preferred embodiments the test nucleic acid molecule is a CpG nucleic acid. According to this and other aspects of the invention involving functional TLR in a

screening assay, in some embodiments the functional TLR is expressed in a cell. In some embodiments the functional TLR is naturally expressed by the cell. In certain preferred embodiments the cell is an isolated mammalian cell that naturally expresses the functional TLR. Whether the cell expresses the TLR naturally or the cell expresses the TLR because an expression vector having an isolated nucleic acid molecule encoding the TLR operatively linked to a promoter has been introduced into the cell, in some embodiments the cell further includes an expression vector comprising an isolated nucleic acid which encodes a reporter construct selected from the group consisting of IL-8, IL-12 p40, NF- κ B-luc, IL-12 p40-luc, and TNF-luc, operatively linked to a promoter.

Also according to this and other aspects of the invention involving functional TLR in a screening assay, in certain embodiments the functional TLR is part of a cell-free system.

Also according to this and other aspects of the invention involving functional TLR in a screening assay, in certain embodiments the functional TLR is part of a complex with another TLR. In certain preferred embodiments the complex is a complex of TLR9 and TLR7. In certain preferred embodiments the complex is a complex of TLR9 and TLR8.

Also according to this and other aspects of the invention involving functional TLR in a screening assay, in certain embodiments the functional TLR is part of a complex with a non-TLR protein selected from the group consisting of MyD88, IRAK, TRAF6, I κ B, NF- κ B, and functional homologues and derivatives thereof.

Further according to this and other aspects of the invention involving functional TLR in a screening assay, in certain embodiments the response mediated by a TLR signal transduction pathway is selected from the group consisting of induction of a gene under control of NF- κ B promoter and secretion of a cytokine.

Also according to this and other aspects of the invention involving functional TLR in a screening assay, in certain embodiments the gene under control of NF- κ B promoter is selected from the group consisting of IL-8, IL-12 p40, NF- κ B-luc, IL-12 p40-luc, and TNF-luc.

Also according to this and other aspects of the invention involving functional TLR in a screening assay, in certain embodiments wherein the cytokine is selected from the group consisting of IL-8, TNF- α , and IL-12 p40.

In a further aspect, the invention provides a screening method for comparing TLR

signaling activity of a test compound with an ISNA. The method entails contacting a functional TLR selected from the group consisting of TLR7, TLR8, and TLR9 with a reference ISNA and detecting a reference response mediated by a TLR signal transduction pathway; contacting a functional TLR selected from the group consisting of TLR7, TLR8, and TLR9 with a test compound and detecting a test response mediated by a TLR signal transduction pathway; and comparing the test response with the reference response to compare the TLR signaling activity of the test compound with the ISNA.

In certain embodiments according to this aspect of the invention, the reference ISNA is a CpG nucleic acid.

In certain embodiments according to this aspect of the invention, the test compound is a polypeptide. In certain embodiments the test compound is part of a combinatorial library of compounds.

In certain embodiments the functional TLR is contacted with the reference ISNA and the test compound independently. Accordingly, in certain embodiments the screening method is a method for identifying an ISNA mimic, and the test compound is determined to be an ISNA mimic when the test response is similar to the reference response obtained with the reference ISNA. A test response is similar to the reference response when the test and reference responses are qualitatively alike, even if not quantitatively alike. Thus, for example, the test and reference responses are considered alike when both responses include induction of a Th1-like immune response. The test response can be quantitatively less than, about the same as, or greater than the reference response.

In certain other embodiments the functional TLR is contacted with the reference ISNA and the test compound concurrently to produce a test-reference response mediated by a TLR signal transduction pathway, wherein the test-reference response may be compared to the reference response. In certain preferred embodiments the screening method is a method for identifying an ISNA agonist, wherein the test compound is an ISNA agonist when the test-reference response is greater than the reference response. In certain preferred embodiments the screening method is a method for identifying an ISNA antagonist, wherein the test compound is an ISNA antagonist when the test-reference response is less than the reference response.

In a further aspect the invention provides a screening method for identifying species

specificity of an ISNA. The method according to this aspect of the invention involves contacting a functional TLR selected from the group consisting of TLR7, TLR8, and TLR9 of a first species with a test ISNA; contacting a functional TLR selected from the group consisting of TLR7, TLR8, and TLR9 of a second species with the test ISNA; measuring a response mediated by a TLR signal transduction pathway associated with the contacting the functional TLR of the first species with the test ISNA; measuring a response mediated by the TLR signal transduction pathway associated with the contacting the functional TLR of the second species with the test ISNA; and comparing (a) the response mediated by a TLR signal transduction pathway associated with the contacting the functional TLR of the first species with the test ISNA with (b) the response mediated by the TLR signal transduction pathway associated with the contacting the functional TLR of the second species with the test ISNA. In preferred embodiments the TLR of the first species corresponds to the TLR of the second species, e.g., the TLR of the first species is human TLR9 and the TLR of the second species is murine TLR9. In certain embodiments the functional TLR may be expressed in a cell, part of cell-free system, or part of a complex with another TLR or with a non-TLR protein, as previously described.

In yet another aspect the invention provides a method for identifying lead compounds for a pharmacological agent useful in the treatment of disease associated with TLR9 signaling activity. The method according to this aspect of the invention involves providing a cell comprising a TLR9 polypeptide or fragment thereof as provided in the second aspect of the invention; contacting the cell with a candidate pharmacological agent under conditions which, in the absence of the candidate pharmacological agent, cause a first amount of TLR9 signaling activity; and determining a second amount of TLR9 signaling activity as a measure of the effect of the pharmacological agent on the TLR9 signaling activity, wherein a second amount of TLR9 signaling activity which is less than the first amount indicates that the candidate pharmacological agent is a lead compound for a pharmacological agent which reduces TLR9 signaling activity and wherein a second amount of TLR9 signaling activity which is greater than the first amount indicates that the candidate pharmacological agent is a lead compound for a pharmacological agent which increases TLR9 signaling activity.

These and other aspects of the invention are described in greater detail below.

Brief Description of the Figures

FIG. 1 is two paired bar graphs showing (A) the induction of NF- κ B and (B) the amount of IL-8 produced by 293 fibroblast cells transfected with human TLR9 in response to exposure to various stimuli, including CpG-ODN, GpC-ODN, LPS, and medium.

5 FIG. 2 is a bar graph showing the induction of NF- κ B produced by 293 fibroblast cells transfected with murine TLR9 in response to exposure to various stimuli, including CpG-ODN, methylated CpG-ODN (Me-CpG-ODN), GpC-ODN, LPS, and medium.

FIG.3 is a series of gel images depicting the results of reverse transcriptase-polymerase chain reaction (RT-PCR) assays for murine TLR9 (mTLR9), human TLR9 (hTLR9), and glyceraldehyde-3-phosphate dehydrogenase (GAPDH) in untransfected control 293 cells, 293 cells transfected with mTLR9 (293-mTLR9), and 293 cells transfected with hTLR9 (293-hTLR9).

FIG. 4 is a graph showing the degree of induction of NF- κ B-luc by various stimuli in stably transfected 293-hTLR9 cells.

FIG. 5 is a graph showing the degree of induction of NF- κ B-luc by various stimuli in stably transfected 293-mTLR9 cells.

FIG. 6 is an image of a Coomassie-stained polyacrylamide gel depicting the presence of soluble hTLR9 in the supernatants of yeast cells transfected with hTLR9, either induced (lane 1) or not induced (lane 2).

20 FIG. 7 is a graph showing proliferation of human B cells in response to various stimuli, including *Escherichia coli* (*E. coli*) DNA, DNase-digested *E. coli* DNA, CpG-ODN, GpC-ODN, and LPS.

FIG. 8 is two paired bar graphs showing induction of (top) IL-8 and (bottom) TNF in plasmacytoid dendritic cells (CD123+ DC) and monocyte-derived dendritic cells (MDDC) in response to various stimuli, including *E. coli* DNA, DNase-digested *E. coli* DNA, CpG-ODN, GpC-ODN, and LPS.

FIG. 9 is a series of images of stained gels showing results of semi-quantitative RT-PCR comparing relative levels of human TLR9, TLR2, and TLR4 mRNA expression in human peripheral blood cells: MDDC (lane 1), purified CD14+ monocytes (lane 2), B cells (lane 3), CD123+ DC (lane 4), CD4+ T cells (lane 5), and CD8+ T cells (lane 6). GAPDH is a control for equalizing amounts of cDNA.

FIG. 10 is a pair of graphs showing amounts of IL-12 induced in (A) human peripheral blood mononuclear cells (PBMC) and (B) murine splenocytes in response to shown concentrations of various ODN, including ODN 2006 (filled circles), 2006-GC (open circles), 1668 (filled triangles), and 1668-GC (open triangles).

FIG. 11 is a quartet of graphs depicting responsiveness of 293 cells transfected with hTLR9 (left panels) or mTLR9 (right panels) upon stimulation with shown concentrations of various ODN, including ODN 2006 (filled circles), 2006-GC (open circles), 1668 (filled triangles), and 1668-GC (open triangles). Responses are shown in terms of induction of NF- κ B-luc (upper panels) and IL-8 (lower panels).

FIG. 12 is a bar graph depicting the dose-response of 293-hTLR9 cells to *E. coli* DNA (black bars) and to DNase-digested *E. coli* DNA (gray bars).

FIG. 13 is a pair of graphs showing the responsiveness of (A) 293-hTLR9 and (B) 293-mTLR9 cells to shown concentrations of phosphodiester versions of ODN 2006 (filled circles), 2006-GC (open circles), 1668 (filled triangles), and 1668-GC (open triangles).

Fig. 14 is a pair of graphs showing the responsiveness of 293-hTLR9 and 293-mTLR9 cells to shown concentrations of ODN 5002 (filled circles) and ODN 5007 (open circles).

FIG. 15 is a bar graph showing the response of 293 cells transfected with mTLR9 to CpG-ODN 1668 is inhibited in a dose-dependent manner by co-exposure to non-CpG-ODN PZ2.

FIG. 16 is a bar graph showing the response of 293-hTLR9 cells to CpG-ODN (black bars) or to TNF (gray bars) in the presence of shown amounts of blocking non-CpG-ODN.

FIG. 17 is a bar graph showing blockade of response of 293-hTLR9 cells to CpG-ODN, but not to IL-1 or TNF, in the presence of Bafilomycin A (gray bars). Control treatment with dimethyl sulfoxide (DMSO) is shown in black bars.

FIG. 18 is a graph showing the effect of varying concentrations of dominant negative human MyD88 on the induction of NF- κ B in 293-hTLR9 cells stimulated with CpG-ODN (open circles), TNF- α (filled circles), or control (filled triangles).

FIG. 19 is a series of three Western blot images showing the response of various polyclonal antibodies to purified hTLR9-FLAG and mTLR9-FLAG: upper panel, anti-human and anti-mouse intracellular; middle, anti-mouse extracellular; and lower, anti-human extracellular. Arrows indicate position of TLR9 in each blot.

FIG. 20 is a bar graph depicting the responsiveness of native form hTLR9 and hTLR9 variant form hTLR9-CXXCm to various stimuli at different concentrations.

FIG. 21 is a bar graph depicting the responsiveness of native form mTLR9 and mTLR9 variant form mTLR9-CXXCm to various stimuli at different concentrations.

FIG. 22 is a bar graph showing the responsiveness of native form mTLR9, mTLR9 variant form mTLR9-Phmut, and mTLR9 variant form mTLR9-MBDmut to various stimuli at different concentrations.

FIG. 23 is a bar graph showing the responsiveness of native form hTLR9, hTLR9 variant form hTLR9-PHmut, and hTLR9 variant form hTLR9-MBDmut to various stimuli at different concentrations.

FIG. 24 is a bar graph showing the responsiveness of native form mTLR9 and mTLR9 variant form mTLR9-TIRh to various stimuli at different concentrations.

FIG. 25 is a bar graph showing the responsiveness of native form hTLR9 and hTLR9 variant form hTLR9-TIRm to various stimuli at different concentrations.

FIG. 26 is a series of linear maps representing various features of human TLR7, TLR8, and TLR9 polypeptides.

FIG. 27 is an image of a silver stained polyacrylamide gel and schematic representation of a fusion protein in which the extracellular domain of human TLR9 (hTLR9) is fused to a human IgG1 Fc domain (hIgG-Fc) with a thrombin protease recognition site interposed. From left to right, the gel was loaded with (1) supernatant of transfectants; (2) lysates of transfectants, treated with thrombin; (3) untreated lysates of transfectants; (4) molecular weight markers; (5) supernatant of mock transfectants; (6) lysates of mock transfectants, treated with thrombin; and (7) untreated lysates of mock transfectants. Soluble hTLR9 and Fc are the products released from intact hTLR9-IgG-Fc following thrombin treatment. Molecular weights are indicated along the right side of the silver stain gel image.

Brief Description of Selected Sequences

SEQ ID NO:1 is the nucleotide sequence encoding a cDNA for murine TLR9.

SEQ ID NO:2 is the nucleotide sequence encoding the coding region of murine TLR9.

SEQ ID NO:3 is the amino acid sequence of a murine TLR9 encoded by SEQ ID NO:1.

SEQ ID NO:173 is the nucleotide sequence encoding a cDNA for murine TLR7.

SEQ ID NO:174 is the nucleotide sequence encoding the coding region of murine TLR7.

SEQ ID NO:175 is the amino acid sequence of a murine TLR7 encoded by SEQ ID NO:173.

SEQ ID NO:190 is the nucleotide sequence encoding a cDNA for murine TLR8.

SEQ ID NO:191 is the nucleotide sequence encoding the coding region of murine TLR8.

SEQ ID NO:192 is the amino acid sequence of a murine TLR8 encoded by SEQ ID NO:190.

Detailed Description of the Invention

The present invention in one aspect involves the identification of cDNAs encoding mouse TLR9, referred to herein as murine TLR9 and, equivalently, mTLR9. The nucleotide sequence of the cDNA for murine TLR9 is presented as SEQ ID NO:1, the coding region of the cDNA for murine TLR9 is presented as SEQ ID NO:2, and the amino acid sequence of the murine TLR9 is presented as SEQ ID NO:3. The closely related human TLR9 (equivalently, hTLR9) was deposited in GenBank under accession numbers AF245704 and NM_017742.

The nucleotide sequence of the cDNA for murine TLR9 presented as SEQ ID NO:1 is 3200 nucleotides long and includes the open reading frame (ORF, bases 40-3135) presented as SEQ ID NO:2 which spans 3096 nucleotides (excluding the stop codon). The amino acid sequence of the murine TLR9 presented as SEQ ID NO:3 is 1032 amino acids (aa) long, and it is believed to include an extracellular domain (aa 1-819), a transmembrane domain (aa 820-837), and an intracellular domain (aa 838-1032).

The amino acid sequence of human TLR9 (SEQ ID NO:6) and the amino acid sequence of the murine TLR9 (SEQ ID NO:3) are thus both 1032 amino acids long. Comparison of the aligned amino acid sequences for the murine and the human TLR9 molecules reveals a single base insertion at aa 435 of the murine TLR9 and a single base deletion at aa 860 of the human TLR9. (See Table 4 below.)

Whereas much of the polypeptide presented herein is identical to human TLR9,

murine TLR9 has several single amino acid differences. These differences in amino acids are specifically amino acids 2, 3, 4, 6, 7, 18, 19, 22, 38, 44, 55, 58, 61, 62, 63, 65, 67, 71, 80, 84, 87, 88, 91, 101, 106, 109, 117, 122, 123, 134, 136, 140, 143, 146, 147, 157, 160, 161, 167, 168, 171, 185, 186, 188, 189, 191, 199, 213, 217, 220, 227, 231, 236, 245, 266, 269, 270, 271, 272, 273, 274, 278, 281, 285, 297, 298, 301, 305, 308, 311, 322, 323, 325, 326, 328, 332, 335, 346, 348, 353, 355, 358, 361, 362, 365, 367, 370, 372, 380, 381, 382, 386, 389, 392, 394, 397, 409, 412, 413, 415, 416, 419, 430, 432, 434, 435, 438, 439, 443, 444, 446, 447, 448, 450, 451, 452, 454, 455, 459, 460, 463, 465, 466, 468, 469, 470, 472, 473, 474, 475, 478, 488, 489, 494, 495, 498, 503, 508, 510, 523, 531, 539, 540, 543, 547, 549, 561, 563, 565, 576, 577, 579, 580, 587, 590, 591, 594, 595, 597, 599, 601, 603, 610, 611, 613, 616, 619, 632, 633, 640, 643, 645, 648, 650, 657, 658, 660, 667, 670, 672, 675, 679, 689, 697, 700, 703, 705, 706, 711, 715, 716, 718, 720, 723, 724, 726, 729, 731, 735, 737, 743, 749, 750, 751, 752, 754, 755, 759, 760, 772, 774, 780, 781, 786, 787, 788, 800, 814, 821, 829, 831, 832, 835, 844, 857, 858, 859, 862, 864, 865, 866, 879, 893, 894, 898, 902, 910, 917, 927, 949, 972, 975, 976, 994, 997, 1000, 1003, 1004, 1010, 1011, 1018, 1023, and 1027 of SEQ ID NO:3

In some forms the mouse protein mTLR9 contains a signal sequence at the N-terminus (amino acids 1-26) which allows transport to the endoplasmic reticulum and subsequently to the cell surface or intracellular compartments. A transmembrane region (amino acids 820-837) anchors the protein to the cell membrane. The cytoplasmic tail contains a Toll/IL-1 receptor (TIR) homology domain which is believed to function in signaling upon ligand binding. Leucine-rich-repeats (LRR) can be found in the extracellular region (a common feature of TLRs) and may be involved in ligand binding or dimerization of the molecule.

Both mouse and human TLR9 have an N-terminal extension of approximately 180 amino acids compared to other TLRs. An insertion also occurs at amino acids 253-268, which is not found in TLRs 1-6 but is present in human TLR7 and human TLR8. (See **Figure 26.**) This insert has two CXXC motifs which participate in forming a CXXC domain. The CXXC domain resembles a zinc finger motif and is found in DNA-binding proteins and in certain specific CpG binding proteins, e.g., methyl-CpG binding protein-1 (MBD-1). Fujita N et al., *Mol Cell Biol* 20:5107-5118 (2000). Both human and mouse TLR9 CXXC domains occur at aa 253-268:

CXXC motif:	GNCXXCXXXXXXXXCXXC	SEQ ID NO:196
Human TLR9:	GNCRRCDHAPNPCMEC	SEQ ID NO:197
Murine TLR9:	GNCRRCDHAPNPCMIC	SEQ ID NO:198

An additional motif involved in CpG binding is the MBD motif, also found in MBD-1, listed below as SEQ ID NO:125. Fujita, N et al., *Mol Cell Biol* 20:5107-18 (2000); Ohki I et al., *EMBO J* 18:6653-6661 (1999). Amino acids 524-554 of hTLR9 and aa 525-555 of mTLR9 correspond to the MBD motif of MBD-1 as shown:

MBD motif:

MBD-1	R-XXXXXXXX-R-X-D-X-Y-XXXXXXXXXX-R-S-XXXXXX-Y	SEQ ID NO:125
hTLR9	Q-XXXXXXXX-K-X-D-X-Y-XXXXXXXXXX-R-L-XXXXXX-Y	SEQ ID NO:126
mTLR9	Q-XXXXXXXX-K-X-D-X-Y-XXXXXXXXXX-Q-L-XXXXXX-Y	SEQ ID NO:127
hTLR9	Q-VLDLSRN-K-L-D-L-Y-HEHSFTELP-R-L-EALDLS-Y	SEQ ID NO:210
mTLR9	Q-VLDLSHN-K-L-D-L-Y-HWKSFSLEP-Q-L-QALDLS-Y	SEQ ID NO:211

Although the signaling functions of MBD-1 and TLR9 are quite different, the core D-X-Y is involved in CpG binding and is conserved. The C-terminal octamer S-XXXXXX-Y of the MBD motif may not be involved in binding and the S is not conserved by TLR9. The other mismatches are highly conserved or moderately conserved; example R to K or R to Q. These changes could explain MBD-1 as a methyl-CpG binder and TLR9 as a non-methyl-CpG binder. The differences between mouse and human TLR9 may explain inter-species differences in CpG-motif sequence selectivity. See **Figure 14** for inter-species sequence selectivity.

As discussed in Example 11 below and shown in **Figures 22 and 23**, the D-X-Y core of this MBD motif occurs as D-L-Y in both mTLR9 (aa 535-537) and hTLR9 (aa 534-536). Substitution of A for D and A for Y in the D-X-Y core, resulting in A-L-A in place of D-L-Y, destroys receptor activity for mTLR9 and hTLR9 alike.

The invention involves in one aspect murine TLR9 nucleic acids and polypeptides, as

well as therapeutics relating thereto. The invention also embraces isolated functionally equivalent variants, useful analogs and fragments of the foregoing nucleic acids and polypeptides; complements of the foregoing nucleic acids; and molecules which selectively bind the foregoing nucleic acids and polypeptides.

5 The murine TLR9 nucleic acids and polypeptides of the invention are isolated. As used herein with respect to nucleic acids, the term "isolated" means: (i) amplified *in vitro* by, for example, polymerase chain reaction (PCR); (ii) recombinantly produced by cloning; (iii) purified, as by cleavage and gel separation; or (iv) synthesized by, for example, chemical synthesis. An isolated nucleic acid is one which is readily manipulable by recombinant DNA techniques well known in the art. Thus, a nucleotide sequence contained in a vector in which 10 5' and 3' restriction sites are known or for which PCR primer sequences have been disclosed is considered isolated, but a nucleic acid sequence existing in its native state in its natural host is not. An isolated nucleic acid may be substantially purified, but need not be. For example, a nucleic acid that is isolated within a cloning or expression vector is not pure in that it may 15 comprise only a tiny percentage of the material in the cell in which it resides. Such a nucleic acid is isolated, however, as the term is used herein because it is readily manipulable by standard techniques known to those of ordinary skill in the art. An isolated nucleic acid as used herein is not a naturally occurring chromosome.

As used herein with respect to polypeptides, "isolated" means separated from its 20 native environment and present in sufficient quantity to permit its identification or use. Isolated, when referring to a protein or polypeptide, means, for example: (i) selectively produced by expression cloning or (ii) purified as by chromatography or electrophoresis. Isolated proteins or polypeptides may be, but need not be, substantially pure. The term "substantially pure" means that the proteins or polypeptides are essentially free of other 25 substances with which they may be found in nature or *in vivo* systems to an extent practical and appropriate for their intended use. Substantially pure polypeptides may be produced by techniques well known in the art. Because an isolated protein may be admixed with a pharmaceutically acceptable carrier in a pharmaceutical preparation, the protein may comprise only a small percentage by weight of the preparation. The protein is nonetheless 30 isolated in that it has been separated from the substances with which it may be associated in living systems, i.e., isolated from other proteins.

As used herein a murine TLR9 nucleic acid refers to an isolated nucleic acid molecule which codes for a murine TLR9 polypeptide. Such nucleic acid molecules code for murine TLR9 polypeptides which include the sequence of SEQ ID NO:3 and fragments thereof. The nucleic acid molecules include the nucleotide sequences of SEQ ID NO:1, SEQ ID NO:2, and nucleotide sequences which differ from the sequences of SEQ ID NO:1 and SEQ ID NO:2 in codon sequence due to the degeneracy of the genetic code. The murine TLR9 nucleic acids of the invention also include alleles of the foregoing nucleic acids, as well as fragments of the foregoing nucleic acids. Such fragments can be used, for example, as probes in hybridization assays and as primers in a polymerase chain reaction. Preferred murine TLR9 nucleic acids include the nucleic acid sequence of SEQ ID NO:1 and SEQ ID NO:2. Complements of the foregoing nucleic acids also are embraced by the invention.

As used herein a murine TLR9 nucleic acid or murine TLR9 polypeptide also embraces homologues and alleles of murine TLR9. In general homologues and alleles typically will share at least 40% nucleotide identity and/or at least 50% amino acid identity to the sequences of specified nucleic acids and polypeptides, respectively. Thus homologues and alleles of murine TLR9 typically will share at least 40% nucleotide identity and/or at least 50% amino acid identity to the sequences of murine TLR9 nucleic acids and TLR9 polypeptides, respectively. In some instances homologues and alleles will share at least 50% nucleotide identity and/or at least 65% amino acid identity and in still other instances will share at least 60% nucleotide identity and/or at least 75% amino acid identity. Preferably the homologues and alleles will share at least 80% nucleotide identity and/or at least 90% amino acid identity, and more preferably will share at least 90% nucleotide identity and/or at least 95% amino acid identity. Most preferably the homologues and alleles will share at least 95% nucleotide identity and/or at least 99% amino acid identity. The homology can be calculated using various publicly available software tools developed by the National Center for Biotechnology Information (NCBI, Bethesda, Maryland) that can be obtained through the internet (<ftp://ncbi.nlm.nih.gov/pub/>). Exemplary tools include the BLAST system available from the NCBI at <http://www.ncbi.nlm.nih.gov>, used with default settings. Pairwise and ClustalW alignments (BLOSUM30 matrix setting) as well as Kyte-Doolittle hydropathic analysis can be obtained, for example, using the MacVector sequence analysis software (Oxford Molecular Group). Watson-Crick complements of the foregoing nucleic acids also

are embraced by the invention.

Alleles of the murine TLR9 nucleic acids of the invention can be identified by conventional techniques. For example, alleles of murine TLR9 can be isolated by hybridizing a probe which includes at least a fragment of SEQ ID NO:1 or SEQ ID NO:2 under stringent conditions with a cDNA library and selecting positive clones. Thus, an aspect of the invention is those nucleic acid sequences which code for murine TLR9 polypeptides and which hybridize to a nucleic acid molecule consisting of SEQ ID NO:1 or SEQ ID NO:2 under stringent conditions.

In screening for murine TLR nucleic acids, a Southern blot may be performed using the foregoing stringent conditions, together with a radioactive probe. After washing the membrane to which the DNA is finally transferred, the membrane can be placed against X-ray film to detect the radioactive signal. Corresponding non-radioactive methods are also well known in the art and can be used to similar effect.

The murine TLR nucleic acids of the invention also include degenerate nucleic acids which include alternative codons to those present in the native materials. For example, serine residues are encoded by the codons AGC, AGT, and TCA, TCC, TCG and TCT. Each of the six codons is equivalent for the purposes of encoding a serine residue. Thus, it will be apparent to one of ordinary skill in the art that any of the serine-encoding nucleotide triplets may be employed to direct the protein synthesis apparatus, *in vitro* or *in vivo*, to incorporate a serine residue into an elongating murine TLR polypeptide. Similarly, nucleotide sequence triplets which encode other amino acid residues include, but are not limited to: CCA, CCC, CCG and CCT (proline codons); CGA, CGC, CGG, CGT, AGA and AGG (arginine codons); ACA, ACC, ACG and ACT (threonine codons); AAC and AAT (asparagine codons); and ATA, ATC and ATT (isoleucine codons). As is well known by those of ordinary skill in the art, other specific amino acid residues may be encoded similarly by multiple nucleotide sequences. Thus, the invention embraces degenerate nucleic acids that differ from the biologically isolated nucleic acids in codon sequence due to the degeneracy of the genetic code. The above-noted codon degeneracy notwithstanding, it is well appreciated by those skilled in the art that there are certain codon usage preferences in specific host organisms, such that in practice it may be preferred to select or to avoid certain degenerate codons in a particular host.

The invention also provides modified nucleic acid molecules which include additions, substitutions and deletions of one or more nucleotides. The modified nucleic acid molecules according to this aspect of the invention exclude fully native human TLR9 nucleic acid molecules (GenBank Accession No. AF245704 (SEQ ID NO:4) or GenBank Accession No. NM_017442 (SEQ ID NO:5)). In preferred embodiments, these modified nucleic acid molecules and/or the polypeptides they encode retain at least one activity or function of the unmodified nucleic acid molecule and/or the polypeptides, such as signaling activity, etc. In certain embodiments, the modified nucleic acid molecules encode modified polypeptides, preferably polypeptides having conservative amino acid substitutions as are described elsewhere herein. The modified nucleic acid molecules are structurally related to the unmodified nucleic acid molecules and in preferred embodiments are sufficiently structurally related to the unmodified nucleic acid molecules so that the modified and unmodified nucleic acid molecules hybridize under stringent conditions known to one of skill in the art.

For example, modified nucleic acid molecules which encode polypeptides having single amino acid changes can be prepared. Each of these nucleic acid molecules can have one, two or three nucleotide substitutions exclusive of nucleotide changes corresponding to the degeneracy of the genetic code as described herein. Likewise, modified nucleic acid molecules which encode polypeptides having two amino acid changes can be prepared which have, e.g., 2-6 nucleotide changes. Numerous modified nucleic acid molecules like these will be readily envisioned by one of skill in the art, including for example, substitutions of nucleotides in codons encoding amino acids 2 and 3, 2 and 4, 2 and 5, 2 and 6, and so on. In the foregoing example, each combination of two amino acids is included in the set of modified nucleic acid molecules, as well as all nucleotide substitutions which code for the amino acid substitutions. Additional nucleic acid molecules that encode polypeptides having additional substitutions (i.e., 3 or more), additions or deletions (e.g., by introduction of a stop codon or a splice site(s)) also can be prepared and are embraced by the invention as readily envisioned by one of ordinary skill in the art. Any of the foregoing nucleic acids or polypeptides can be tested by routine experimentation for retention of structural relation or activity to the nucleic acids and/or polypeptides disclosed herein.

The invention also provides isolated fragments of SEQ ID NO:1 and SEQ ID NO:2. The fragments can be used as probes in Southern blot assays to identify such nucleic acids, or

they can be used in amplification assays such as those employing PCR. Smaller fragments are those comprising 12, 13, 14, 15, 16, 17, 18, 20, 22, 25, 30, 40, 50, or 75 nucleotides, and every integer therebetween, and are useful, e.g., as primers for nucleic acid amplification procedures. As known to those skilled in the art, larger probes such as 200, 250, 300, 400 or more nucleotides are preferred for certain uses such as Southern blots, while smaller fragments will be preferred for uses such as PCR. Fragments also can be used to produce fusion proteins for generating antibodies or determining binding of the polypeptide fragments. Likewise, fragments can be employed to produce non-fused fragments of the murine TLR9 polypeptides, useful, for example, in the preparation of antibodies, in immunoassays, and the like. The foregoing nucleic acid fragments further can be used as antisense molecules to inhibit the expression of murine TLR9 nucleic acids and polypeptides, particularly for therapeutic purposes as described in greater detail below.

The invention also includes functionally equivalent variants of the murine TLR9, which include variant nucleic acids and polypeptides which retain one or more of the functional properties of the murine TLR9. Preferably such variants include the murine-specific N-terminal domain (e.g., amino acids 1-819 or amino acids 1-837 of SEQ ID NO:3).

For example, variants include a fusion protein which includes the extracellular and transmembrane domains of the murine TLR9 (i.e., amino acids 1-837) which retains the ability to interact with extracellular molecules in a manner similar to intact murine TLR9.

Alternative variants include, for example, a fusion protein which includes the cytoplasmic domain of murine TLR9 (i.e., amino acids 838-1032) which retains the ability to interact with intracellular molecules in a manner similar to intact murine TLR9. Still other functionally equivalent variants include truncations, deletions, point mutations, or additions of amino acids to the sequence of SEQ ID NO:3 which retain functions of SEQ ID NO:3. For example, the FLAG peptide sequence (DYKDDDDK) can be added at the N-terminal end, or green fluorescent protein (GFP) can be added at the C-terminal end. All such addition variant polypeptides are preferably made by translation of modified nucleic acids based on SEQ ID NO:1 or SEQ ID NO:2 with corresponding appropriate coding nucleic acid sequence appended thereto with maintenance of the proper reading frame.

Functionally equivalent variants also include a murine TLR9 which has had a portion (e.g., of the N-terminus) removed or replaced by a similar domain from another TLR (e.g., a

“domain-swapping” variant). Examples of such domain-swapping variants include at least two types: those involving swapping a TLR9 domain from one species with a TLR9 domain from another species, and those involving swapping a TLR domain from TLR9 with a TLR domain from another TLR. In certain preferred embodiments the swapping involves corresponding domains between the different TLR molecules. It is believed that certain such domain-swapping variants are not functionally equivalent in a literal sense, insofar as they can function in a manner superior to either or both intact parent TLR molecules from which a particular domain-swapping variant derives. For example, the TLR/IL-1R signaling mediated by human TLR9 could be limited, not by the capacity of its extracellular domain to interact with CpG ODN, but rather by the capacity of its cytoplasmic domain to engage the TLR/IL-1R signaling pathway. In such a circumstance, it could be advantageous to substitute a more potent cytoplasmic domain from a TLR9 from another species. Such a domain-swapping variant, e.g., chimeric hTLR9/mTLR9, could be used in screening assays for CpG immunagonist/antagonists to increase the sensitivity of the assay, without changing the species specificity.

Other functionally equivalent variants will be known to one of ordinary skill in the art, as will be methods for preparing such variants. The activity of a functionally equivalent variant can be determined using the methods provided herein, and in references that have described assays using other TLRs and TLRs of other species. Such variants are useful, *inter alia*, for evaluating bioavailability of drugs, in assays for identification of compounds which bind to and/or regulate the signaling function of the murine TLR9, and for determining the portions of the murine TLR9 which are required for signaling activity.

Variants which are non-functional also can be prepared as described above. Such variants are useful, for example, as negative controls in experiments testing TLR9 signaling activity. Examples of non-functional variants include those incorporating a mutation of proline at aa 915 to histidine (P915H) which renders both mTLR9 and hTLR9 nonfunctional with respect to signaling. Further examples of non-functional variants include those incorporating a mutation of the D-X-Y core of the MBD motif to A-L-A, as discussed above, to render both mTLR9 and hTLR9 nonfunctional with respect to CpG DNA binding.

A murine TLR9 nucleic acid, in one embodiment, is operably linked to a gene expression sequence which can direct the expression of the murine TLR9 nucleic acid within

a eukaryotic or prokaryotic cell. A "gene expression sequence" is any regulatory nucleotide sequence, such as a promoter sequence or promoter-enhancer combination, which facilitates the efficient transcription and translation of the nucleic acid to which it is operably linked.

With respect to murine TLR9 nucleic acid, the "gene expression sequence" is any regulatory nucleotide sequence, such as a promoter sequence or promoter-enhancer combination, which facilitates the efficient transcription and translation of the murine TLR9 nucleic acid to which it is operably linked. The gene expression sequence may, for example, be a mammalian or viral promoter, such as a constitutive or inducible promoter. Constitutive mammalian promoters include, but are not limited to, the promoters for the following genes: hypoxanthine phosphoribosyl transferase (HPRT), adenosine deaminase, pyruvate kinase, β -actin promoter, and other constitutive promoters. Exemplary viral promoters which function constitutively in eukaryotic cells include, for example, promoters from the simian virus (e.g., SV40), papilloma virus, adenovirus, human immunodeficiency virus (HIV), Rous sarcoma virus (RSV), cytomegalovirus (CMV), the long terminal repeats (LTR) of Moloney murine leukemia virus and other retroviruses, and the thymidine kinase (TK) promoter of herpes simplex virus. Other constitutive promoters are known to those of ordinary skill in the art. The promoters useful as gene expression sequences of the invention also include inducible promoters. Inducible promoters are expressed in the presence of an inducing agent. For example, the metallothionein (MT) promoter is induced to promote transcription and translation in the presence of certain metal ions. Other inducible promoters are known to those of ordinary skill in the art.

In general, the gene expression sequence shall include, as necessary, 5' non-transcribing and 5' non-translating sequences involved with the initiation of transcription and translation, respectively, such as a TATA box, capping sequence, CAAT sequence, and the like. Especially, such 5' non-transcribing sequences will include a promoter region which includes a promoter sequence for transcriptional control of the operably joined murine TLR9 nucleic acid. The gene expression sequences optionally include enhancer sequences or upstream activator sequences as desired.

Generally a nucleic acid coding sequence and a gene expression sequence are said to be "operably linked" when they are covalently linked in such a way as to place the transcription and/or translation of the nucleic acid coding sequence under the influence or

control of the gene expression sequence. Thus the murine TLR9 nucleic acid sequence and the gene expression sequence are said to be "operably linked" when they are covalently linked in such a way as to place the transcription and/or translation of the murine TLR9 coding sequence under the influence or control of the gene expression sequence. If it is desired that the murine TLR9 sequence be translated into a functional protein, two DNA sequences are said to be operably linked if induction of a promoter in the 5' gene expression sequence results in the transcription of the murine TLR9 sequence and if the nature of the linkage between the two DNA sequences does not (1) result in the introduction of a frame-shift mutation, (2) interfere with the ability of the promoter region to direct the transcription of the murine TLR9 sequence, or (3) interfere with the ability of the corresponding RNA transcript to be translated into a protein. Thus, a gene expression sequence would be operably linked to a murine TLR9 nucleic acid sequence if the gene expression sequence were capable of effecting transcription of that murine TLR9 nucleic acid sequence such that the resulting transcript might be translated into the desired protein or polypeptide.

The murine TLR9 nucleic acid molecules and the murine TLR9 polypeptides (including the murine TLR9 inhibitors described below) of the invention can be delivered to the eukaryotic or prokaryotic cell alone or in association with a vector. In its broadest sense, a "vector" is any vehicle capable of facilitating: (1) delivery of a nucleic acid or polypeptide to a target cell, (2) uptake of a nucleic acid or polypeptide by a target cell, or (3) expression of a nucleic acid molecule or polypeptide in a target cell. In this particular setting, a "vector" is any vehicle capable of facilitating: (1) delivery of a murine TLR9 nucleic acid or polypeptide to a target cell, (2) uptake of a murine TLR9 nucleic acid or polypeptide by a target cell, or (3) expression of a murine TLR9 nucleic acid molecule or polypeptide in a target cell. Preferably, the vectors transport the murine TLR9 nucleic acid or polypeptide into the target cell with reduced degradation relative to the extent of degradation that would result in the absence of the vector. Optionally, a "targeting ligand" can be attached to the vector to selectively deliver the vector to a cell which expresses on its surface the cognate receptor (e.g., a receptor, an antigen recognized by an antibody) for the targeting ligand. In this manner, the vector (containing a murine TLR9 nucleic acid or a murine TLR9 polypeptide) can be selectively delivered to a specific cell. In general, the vectors useful in the invention are divided into two classes: biological vectors and chemical/physical vectors. Biological

vectors are more useful for delivery/uptake of murine TLR9 nucleic acids to/by a target cell. Chemical/physical vectors are more useful for delivery/uptake of murine TLR9 nucleic acids or murine TLR9 proteins to/by a target cell.

Biological vectors include, but are not limited to, plasmids, phagemids, viruses, other vehicles derived from viral or bacterial sources that have been manipulated by the insertion or incorporation of the nucleic acid sequences of the invention, and free nucleic acid fragments which can be linked to the nucleic acid sequences of the invention. Viral vectors are a preferred type of biological vector and include, but are not limited to, nucleic acid sequences from the following viruses: retroviruses, such as Moloney murine leukemia virus; Harvey murine sarcoma virus; murine mammary tumor virus; Rous sarcoma virus; adenovirus; adeno-associated virus; SV40-type viruses; polyoma viruses; poxviruses; Epstein-Barr viruses; papilloma viruses; herpes virus; vaccinia virus; and polio virus. One can readily employ other vectors not named but known in the art.

Preferred viral vectors are based on non-cytopathic eukaryotic viruses in which non-essential genes have been replaced with the gene of interest. Non-cytopathic viruses include retroviruses, the life cycle of which involves reverse transcription of genomic viral RNA into DNA with subsequent proviral integration into host cellular DNA. In general, the retroviruses are replication-deficient (i.e., capable of directing synthesis of the desired proteins, but incapable of manufacturing an infectious particle). Such genetically altered retroviral expression vectors have general utility for the high-efficiency transduction of genes *in vivo*. Standard protocols for producing replication-deficient retroviruses (including the steps of incorporation of exogenous genetic material into a plasmid, transfection of a packaging cell line with plasmid, production of recombinant retroviruses by the packaging cell line, collection of viral particles from tissue culture media, and infection of the target cells with viral particles) are provided in Kriegler, M., *"Gene Transfer and Expression, A Laboratory Manual,"* W.H. Freeman Co., New York (1990) and Murray, E.J., ed., *"Methods in Molecular Biology,"* vol. 7, Humana Press, Inc., Clifton, New Jersey (1991).

Another preferred virus for certain applications is the adeno-associated virus (AAV), a double-stranded DNA virus. The AAV can be engineered to be replication-deficient and is capable of infecting a wide range of cell types and species. It further has advantages, such as heat and lipid solvent stability; high transduction frequencies in cells of diverse lineages; and

lack of superinfection inhibition thus allowing multiple series of transductions. Reportedly, the AAV can integrate into human cellular DNA in a site-specific manner, thereby minimizing the possibility of insertional mutagenesis and variability of inserted gene expression. In addition, wild-type AAV infections have been followed in tissue culture for greater than 100 passages in the absence of selective pressure, implying that the AAV genomic integration is a relatively stable event. The AAV can also function in an extrachromosomal fashion.

Expression vectors containing all the necessary elements for expression are commercially available and known to those skilled in the art. See, e.g., *Molecular Cloning: A Laboratory Manual*, J. Sambrook, et al., eds., Second Edition, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York, 1989. Cells are genetically engineered by the introduction into the cells of heterologous DNA (RNA) encoding a murine TLR9 polypeptide or fragment or variant thereof. That heterologous DNA (RNA) is placed under operable control of transcriptional elements to permit the expression of the heterologous DNA in the host cell.

Preferred systems for mRNA expression in mammalian cells are those such as pRc/CMV (available from Invitrogen, Carlsbad, CA) that contain a selectable marker such as a gene that confers G418 resistance (which facilitates the selection of stably transfected cell lines) and the human CMV enhancer-promoter sequences. Additionally, suitable for expression in primate or canine cell lines is the pCEP4 vector (Invitrogen), which contains an Epstein Barr virus (EBV) origin of replication, facilitating the maintenance of plasmid as a multicopy extrachromosomal element. Another expression vector is the pEF-BOS plasmid containing the promoter of polypeptide Elongation Factor 1 α , which stimulates efficiently transcription *in vitro*. The plasmid is described by Mishizuma and Nagata (*Nucleic Acids Res* 18:5322 (1990)), and its use in transfection experiments is disclosed by, for example, Demoulin (*Mol Cell Biol* 16:4710-4716 (1996)). Still another preferred expression vector is an adenovirus, described by Stratford-Perricaudet, which is defective for E1 and E3 proteins (*J Clin Invest* 90:626-630 (1992)).

In addition to the biological vectors, chemical/physical vectors may be used to deliver a nucleic acid or polypeptide to a target cell and facilitate uptake thereby. As used herein, a "chemical/physical vector" refers to a natural or synthetic molecule, other than those derived

from bacteriological or viral sources, capable of delivering an isolated nucleic acid or polypeptide to a cell. As used herein with respect to a murine TLR9 nucleic acid or polypeptide, a "chemical/physical vector" refers to a natural or synthetic molecule, other than those derived from bacteriological or viral sources, capable of delivering the isolated murine TLR9 nucleic acid or polypeptide to a cell.

A preferred chemical/physical vector of the invention is a colloidal dispersion system. Colloidal dispersion systems include lipid-based systems including oil-in-water emulsions, micelles, mixed micelles, and liposomes. A preferred colloidal system of the invention is a liposome. Liposomes are artificial membrane vesicles which are useful as a delivery vector *in vivo* or *in vitro*. It has been shown that large unilamellar vesicles (LUV), which range in size from 0.2 - 4.0 μm can encapsulate large macromolecules. RNA, DNA, and intact virions can be encapsulated within the aqueous interior and be delivered to cells in a biologically active form (Fraley et al., *Trends Biochem Sci* 6:77 (1981)). In order for a liposome to be an efficient nucleic acid transfer vector, one or more of the following characteristics should be present: (1) encapsulation of the nucleic acid of interest at high efficiency with retention of biological activity; (2) preferential and substantial binding to a target cell in comparison to non-target cells; (3) delivery of the aqueous contents of the vesicle to the target cell cytoplasm at high efficiency; and (4) accurate and effective expression of genetic information.

Liposomes may be targeted to a particular tissue by coupling the liposome to a specific ligand such as a monoclonal antibody, sugar, glycolipid, or protein. Ligands which may be useful for targeting a liposome to a particular cell will depend on the particular cell or tissue type. Additionally when the vector encapsulates a nucleic acid, the vector may be coupled to a nuclear targeting peptide, which will direct the murine TLR9 nucleic acid to the nucleus of the host cell.

Liposomes are commercially available from Gibco BRL, for example, as LIPOFECTINTM and LIPOFECTACETM, which are formed of cationic lipids such as N-[1-(2, 3 dioleyloxy)-propyl]-N, N, N-trimethylammonium chloride (DOTMA) and dimethyl dioctadecylammonium bromide (DDAB). Methods for making liposomes are well known in the art and have been described in many publications.

Other exemplary compositions that can be used to facilitate uptake by a target cell of nucleic acids in general, and nucleic acids encoding the murine TLR9 in particular, include

calcium phosphate and other chemical mediators of intracellular transport, microinjection compositions, electroporation and homologous recombination compositions (e.g., for integrating a murine TLR9 nucleic acid into a preselected location within a target cell chromosome).

5 The invention also embraces so-called expression kits, which allow the artisan to prepare a desired expression vector or vectors. Such expression kits include at least separate portions of the previously discussed coding sequences. Other components may be added, as desired, as long as the previously mentioned sequences, which are required, are included.

10 It will also be recognized that the invention embraces the use of the murine TLR9 cDNA sequences in expression vectors to transfect host cells and cell lines, be these prokaryotic (e.g., *E. coli*), or eukaryotic (e.g., 293 fibroblast cells (ATCC, CRL-1573), MonoMac-6, THP-1, U927, CHO cells, COS cells, yeast expression systems and recombinant baculovirus expression in insect cells). Especially useful are mammalian cells such as human, pig, goat, primate, rodent, guinea pig, etc. They may be of a wide variety of tissue types, and include primary cells and cell lines. The expression vectors require that the
15 pertinent sequence, i.e., those nucleic acids described *supra*, be operably linked to a promoter.

20 The invention also provides isolated murine TLR9 polypeptides which include the amino acid sequences of SEQ ID NO:3 and fragments thereof, encoded by the murine TLR9 nucleic acids described above. Murine TLR9 polypeptides also embrace alleles, functionally equivalent variants and analogs (those non-allelic polypeptides which vary in amino acid sequence from the disclosed murine TLR9 polypeptides by 1, 2, 3, 4, 5, or more amino acids) provided that such polypeptides retain TLR9 activity. Non-functional variants also are embraced by the invention; these are useful as antagonists of TLR9 signaling function, as negative controls in assays, and the like. Such alleles, variants, analogs and fragments are
25 useful, for example, alone or as fusion proteins for a variety of purposes including as a component of assays.

30 Fragments of a polypeptide preferably are those fragments which retain a distinct functional capability of the intact polypeptide, in particular as a receptor of various molecules. Accordingly, fragments of a TLR9 polypeptide preferably are those fragments which retain a distinct functional capability of the TLR9 polypeptide, in particular as a receptor of various molecules. Of particular interest are fragments that bind to ISNAs, including, for example,

fragments that bind CpG nucleic acids. Other functional capabilities which can be retained in a fragment of a polypeptide include signal transduction (e.g., TLR/IL-1R signaling by murine TLR9), interaction with antibodies and interaction with other polypeptides (such as would be found in a protein complex). Those skilled in the art are well versed in methods that can be applied for selecting fragments which retain a functional capability of the murine TLR9. Confirmation of the functional capability of the fragment can be carried out by synthesis of the fragment and testing of the capability according to standard methods. For example, to test the signaling activity of a murine TLR9 fragment, one inserts or expresses the fragment in a cell in which signaling can be measured. Such methods, which are standard in the art, are described further herein.

The invention embraces variants of the murine TLR9 polypeptides described above. As used herein, a "variant" of a polypeptide is a polypeptide which contains one or more modifications to the primary amino acid sequence of a polypeptide. Accordingly, a "variant" of a murine TLR9 polypeptide is a polypeptide which contains one or more modifications to the primary amino acid sequence of a murine TLR9 polypeptide. Modifications which create a murine TLR9 variant can be made to a murine TLR9 polypeptide for a variety of reasons, including 1) to reduce or eliminate an activity of a murine TLR9 polypeptide, such as signaling; 2) to enhance a property of a murine TLR9 polypeptide, such as signaling, binding affinity for nucleic acid ligand or other ligand molecule, protein stability in an expression system, or the stability of protein-protein binding; 3) to provide a novel activity or property to a murine TLR9 polypeptide, such as addition of an antigenic epitope or addition of a detectable moiety, e.g., luciferase, FLAG peptide, GFP; 4) to establish that an amino acid substitution does or does not affect molecular signaling activity; or 5) reduce immunogenicity of a murine TLR9 polypeptide. Modifications to a murine TLR9 polypeptide are typically made to the nucleic acid which encodes the murine TLR9 polypeptide, and can include deletions, point mutations, truncations, amino acid substitutions and additions of amino acids or non-amino acid moieties. Alternatively, modifications can be made directly to the polypeptide, such as by cleavage, addition of a linker molecule, addition of a detectable moiety (for example, biotin, fluorophore, radioisotope, enzyme, or peptide), addition of a fatty acid, and the like.

Modifications also embrace fusion proteins comprising all or part of the murine TLR9

amino acid sequence. One of skill in the art will be familiar with methods for predicting the effect on protein conformation of a change in protein sequence, and can thus "design" a variant murine TLR9 according to known methods. One example of such a method is described by Dahiyat and Mayo in *Science* 278:82-87 (1997), whereby proteins can be designed *de novo*. The method can be applied to a known protein to vary only a portion of the polypeptide sequence. By applying the computational methods of Dahiyat and Mayo, specific variants of a murine TLR9 polypeptide can be proposed and tested to determine whether the variant retains a desired conformation.

Variants include murine TLR9 polypeptides which are modified specifically to alter a feature of the polypeptide unrelated to its physiological activity. For example, cysteine residues can be substituted or deleted to prevent unwanted disulfide linkages. Similarly, certain amino acids can be changed to enhance expression of a murine TLR9 polypeptide by eliminating proteolysis by proteases in an expression system (e.g., dibasic amino acid residues in yeast expression systems in which KEX2 protease activity is present).

Mutations of a nucleic acid which encode a murine TLR9 polypeptide preferably preserve the amino acid reading frame of the coding sequence, and preferably do not create regions in the nucleic acid which are likely to hybridize to form secondary structures, such as hairpins or loops, which can be deleterious to expression of the variant polypeptide.

Mutations can be made by selecting an amino acid substitution, or by random mutagenesis of a selected site in a nucleic acid which encodes the polypeptide. Variant polypeptides are then expressed and tested for one or more activities to determine which mutation provides a variant polypeptide with a desired property. Further mutations can be made to variants (or to non-variant murine TLR9 polypeptides) which are silent as to the amino acid sequence of the polypeptide, but which provide preferred codons for translation in a particular host. The preferred codons for translation of a nucleic acid in, e.g., *E. coli*, are well known to those of ordinary skill in the art. Still other mutations can be made to the noncoding sequences of a murine TLR9 gene or cDNA clone to enhance expression of the polypeptide.

The activity of variants of murine TLR9 polypeptides can be tested by cloning the gene encoding the variant murine TLR9 polypeptide into a prokaryotic or eukaryotic (e.g., mammalian) expression vector, introducing the vector into an appropriate host cell,

expressing the variant murine TLR9 polypeptide, and testing for a functional capability of the murine TLR9 polypeptides as disclosed herein. For example, the variant murine TLR9 polypeptide can be tested for ability to provide signaling, as set forth below in the examples. Preparation of other variant polypeptides may favor testing of other activities, as will be known to one of ordinary skill in the art.

The skilled artisan will also realize that conservative amino acid substitutions may be made in murine TLR9 polypeptides to provide functionally equivalent variants of the foregoing polypeptides, i.e., variants which retain the functional capabilities of the murine TLR9 polypeptides. As used herein, a "conservative amino acid substitution" refers to an amino acid substitution which does not alter the relative charge or size characteristics of the polypeptide in which the amino acid substitution is made. Variants can be prepared according to methods for altering polypeptide sequence known to one of ordinary skill in the art such as are found in references which compile such methods, e.g., *Molecular Cloning: A Laboratory Manual*, J. Sambrook, et al., eds., Second Edition, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York, 1989, or *Current Protocols in Molecular Biology*, F.M. Ausubel, et al., eds., John Wiley & Sons, Inc., New York. Exemplary functionally equivalent variants of the murine TLR9 polypeptides include conservative amino acid substitutions of SEQ ID NO:3. Conservative substitutions of amino acids include substitutions made amongst amino acids within the following groups: (a) M, I, L, V; (b) F, Y, W; (c) K, R, H; (d) A, G; (e) S, T; (f) Q, N; and (g) E, D.

Conservative amino acid substitutions in the amino acid sequence of murine TLR9 polypeptide to produce functionally equivalent variants of murine TLR9 typically are made by alteration of the nucleic acid sequence encoding murine TLR9 polypeptides (e.g., SEQ ID NO:1 and SEQ ID NO:2). Such substitutions can be made by a variety of methods known to one of ordinary skill in the art. For example, amino acid substitutions may be made by PCR-directed mutation, site-directed mutagenesis according to the method of Kunkel (Kunkel, *Proc Natl Acad Sci USA* 82:488-492 (1985)), or by chemical synthesis of a gene encoding a murine TLR9 polypeptide. The activity of functionally equivalent fragments of murine TLR9 polypeptides can be tested by cloning the gene encoding the altered murine TLR9 polypeptide into a bacterial or mammalian expression vector, introducing the vector into an appropriate host cell, expressing the altered murine TLR9 polypeptide, and testing for the ability of the

murine TLR9 polypeptide to mediate a signaling event. Peptides which are chemically synthesized can be tested directly for function.

A variety of methodologies well known to the skilled practitioner can be utilized to obtain isolated murine TLR9 polypeptide molecules. The polypeptide may be purified from cells which naturally produce the polypeptide by chromatographic means or immunological recognition. Alternatively, an expression vector may be introduced into cells to cause production of the polypeptide. In another method, mRNA transcripts may be microinjected or otherwise introduced into cells to cause production of the encoded polypeptide. Translation of mRNA in cell-free extracts such as the reticulocyte lysate system also may be used to produce polypeptide. Those skilled in the art also can readily follow known methods for isolating murine TLR9 polypeptides. These include, but are not limited to, immunochromatography, HPLC, size-exclusion chromatography, ion-exchange chromatography and immune-affinity chromatography.

The invention as described herein has a number of uses, some of which are described elsewhere herein. For example, the invention permits isolation of the murine TLR9 polypeptide molecules by, e.g., expression of a recombinant nucleic acid to produce large quantities of polypeptide which may be isolated using standard protocols. As another example, the isolation of the murine TLR9 gene makes it possible for murine TLR9 to be used in methods for assaying molecular interactions involving TLR9.

As discussed further in the Examples below, it has been discovered according to one aspect of the invention that responsiveness to ISNA can be reconstituted in ISNA-unresponsive cells by introducing into ISNA-unresponsive cells an expression vector that directs the expression of murine TLR9 (and certain homologues and variants thereof). Cells so reconstituted also exhibit responses to substances other than phosphorothioate ISNA, e.g., *E. coli* DNA, phosphodiester CpG-ODN, and even methylated CpG-ODN.

Also as discussed further in the Examples below, it has been discovered according to certain aspects of the instant invention that TLR9 not only confers upon cells the ability to signal in response to binding ISNA, but also confers both sequence specificity and species specificity to such signaling responses. Thus murine TLR9 signaling in response to CpG-ODN 1668, reportedly an optimal murine ISNA, was found to be significantly stronger than the corresponding murine TLR9 signaling response to CpG-ODN 2006, reportedly an optimal

human ISNA. The converse was also found to be true, i.e., human TLR9 signaling in response to CpG-ODN 2006 was found to be significantly stronger than the corresponding human TLR9 signaling response to CpG-ODN 1668. Furthermore, it has been discovered according to the instant invention that certain types of cells preferentially express TLR9. For example, TLR9 is strongly expressed in B cells and plasmacytoid dendritic cells (CD123+ DC), but only weakly by T cells, monocyte-derived dendritic cells (MDDC), and CD14+ monocytes. In contrast, TLR2 and TLR4 are strongly expressed by MDDC and CD14+ monocytes, but relatively weakly by B cells, CD123+ DC, and T cells.

The invention also embraces agents which bind selectively to the murine TLR9 nucleic acid molecules or polypeptides as well as agents which bind to variants and fragments of the polypeptides and nucleic acids as described herein. The agents include polypeptides which bind to murine TLR9, and antisense nucleic acids, both of which are described in greater detail below. The agents can inhibit or increase murine TLR9-mediated signaling activity (antagonists and agonists, respectively).

Some of the agents are inhibitors. A murine TLR9 inhibitor is an agent that inhibits murine TLR9-mediated signaling across a cell membrane.

As used herein "TLR9 signaling" refers to an ability of a TLR9 polypeptide to activate the TLR/IL-1R (TIR) signaling pathway, also referred to herein as the TLR signal transduction pathway. Without meaning to be held to any particular theory, it is believed that the TLR/IL-1R signaling pathway involves signaling via the molecules myeloid differentiation marker 88 (MyD88) and tumor necrosis factor (TNF) receptor-associated factor 6 (TRAF6), leading to activation of kinases of the I κ B kinase complex and the c-jun NH₂-terminal kinases (e.g., JNK 1/2). Häcker H et al., *J Exp Med* 192:595-600 (2000). A molecule which inhibits TLR9 activity (an antagonist) is one which inhibits TLR9-mediated activation of the TLR/IL-1R signaling pathway, and a molecule which increases TLR9 signaling (an agonist) is one which increases TLR9-mediated activation of the TLR/IL-1R signaling pathway. Changes in TLR9 activity can be measured by assays such as those disclosed herein, including expression of genes under control of κ B-sensitive promoters and enhancers. Such naturally occurring genes include the genes encoding IL-1 β , IL-6, IL-8, the p40 subunit of interleukin 12 (IL-12p40), and the costimulatory molecules CD80 and CD86. Other genes can be placed under the control of such regulatory elements (see below) and thus

serve to report the level of TLR9 signaling. Additional nucleotide sequence can be added to SEQ ID NO:1 or SEQ ID NO:2, preferably to the 5' or the 3' end of SEQ ID NO:2, to yield a nucleotide sequence encoding a chimeric polypeptide that includes a detectable or reporter moiety, e.g., FLAG, luciferase (luc), green fluorescent protein (GFP) and others known by those skilled in the art. These are discussed in greater detail in the Examples below.

In one embodiment the murine TLR9 inhibitor is an antisense oligonucleotide that selectively binds to a murine TLR9 nucleic acid molecule, to reduce the expression of murine TLR9 (or TLR9 of another species) in a cell. This is desirable in virtually any medical condition wherein a reduction of TLR9 signaling activity is desirable.

As used herein, the term "antisense oligonucleotide" or "antisense" describes an oligonucleotide that is an oligoribonucleotide, oligodeoxyribonucleotide, modified oligoribonucleotide, or modified oligodeoxyribonucleotide which hybridizes under physiological conditions to DNA comprising a particular gene or to an mRNA transcript of that gene and, thereby, inhibits the transcription of that gene and/or the translation of that mRNA. The antisense molecules are designed so as to interfere with transcription or translation of a target gene upon hybridization with the target gene or transcript. Those skilled in the art will recognize that the exact length of the antisense oligonucleotide and its degree of complementarity with its target will depend upon the specific target selected, including the sequence of the target and the particular bases which comprise that sequence. It is preferred that the antisense oligonucleotide be constructed and arranged so as to bind selectively with the target under physiological conditions, i.e., to hybridize substantially more to the target sequence than to any other sequence in the target cell under physiological conditions.

Based upon SEQ ID NO:1 and SEQ ID NO:2, or upon allelic or homologous genomic and/or cDNA sequences, one of skill in the art can easily choose and synthesize any of a number of appropriate antisense molecules for use in accordance with the present invention. In order to be sufficiently selective and potent for inhibition, such antisense oligonucleotides should comprise at least 10 and, more preferably, at least 15 consecutive bases which are complementary to the target, although in certain cases modified oligonucleotides as short as 7 bases in length have been used successfully as antisense oligonucleotides. Wagner RW et al., *Nat Biotechnol* 14:840-844 (1996). Most preferably, the antisense oligonucleotides comprise

a complementary sequence of 20-30 bases. Although oligonucleotides may be chosen which are antisense to any region of the gene or mRNA transcripts, in preferred embodiments the antisense oligonucleotides correspond to N-terminal or 5' upstream sites such as translation initiation, transcription initiation or promoter sites. In addition, 3'-untranslated regions may be targeted. Targeting to mRNA splicing sites has also been used in the art but may be less preferred if alternative mRNA splicing occurs. In addition, the antisense is targeted, preferably, to sites in which mRNA secondary structure is not expected (see, e.g., Sainio et al., *Cell Mol Neurobiol* 14(5):439-457 (1994)) and at which polypeptides are not expected to bind. Thus, the present invention also provides for antisense oligonucleotides which are complementary to allelic or homologous cDNAs and genomic DNAs corresponding to murine TLR9 nucleic acid containing SEQ ID NO:1 or SEQ ID NO:2.

In one set of embodiments, the antisense oligonucleotides of the invention may be composed of "natural" deoxyribonucleotides, ribonucleotides, or any combination thereof. That is, the 5' end of one native nucleotide and the 3' end of another native nucleotide may be covalently linked, as in natural systems, via a phosphodiester internucleoside linkage. These oligonucleotides may be prepared by art-recognized methods which may be carried out manually or by an automated synthesizer. They also may be produced recombinantly by vectors.

In preferred embodiments, however, the antisense oligonucleotides of the invention also may include "modified" oligonucleotides. That is, the oligonucleotides may be modified in a number of ways which do not prevent them from hybridizing to their target but which enhance their stability or targeting or which otherwise enhance their therapeutic effectiveness.

The term "modified oligonucleotide" as used herein describes an oligonucleotide in which (1) at least two of its nucleotides are covalently linked via a synthetic internucleoside linkage (i.e., a linkage other than a phosphodiester linkage between the 5' end of one nucleotide and the 3' end of another nucleotide) and/or (2) a chemical group not normally associated with nucleic acids has been covalently attached to the oligonucleotide. Preferred synthetic internucleoside linkages are phosphorothioates, alkylphosphonates, phosphorodithioates, phosphate esters, alkylphosphonothioates, phosphoramidates, carbamates, carbonates, phosphate triesters, acetamides, carboxymethyl esters and peptides.

The term "modified oligonucleotide" also encompasses oligonucleotides with a

covalently modified base and/or sugar. For example, modified oligonucleotides include oligonucleotides having backbone sugars which are covalently attached to low molecular weight organic groups other than a hydroxyl group at the 3' position and other than a phosphate group at the 5' position. Thus modified oligonucleotides may include a 2'-O-alkylated ribose group. In addition, modified oligonucleotides may include sugars such as arabinose instead of ribose. The present invention, thus, contemplates pharmaceutical preparations containing modified antisense molecules that are complementary to and hybridizable with, under physiological conditions, nucleic acids encoding murine TLR9 polypeptides, together with pharmaceutically acceptable carriers.

Agents which bind murine TLR9 also include binding peptides and other molecules which bind to the murine TLR9 polypeptide and complexes containing the murine TLR9 polypeptide. When the binding molecules are inhibitors, the molecules bind to and inhibit the activity of murine TLR9. When the binding molecules are activators, the molecules bind to and increase the activity of murine TLR9. To determine whether a murine TLR9 binding agent binds to murine TLR9 any known binding assay may be employed. For example, the binding agent may be immobilized on a surface and then contacted with a labeled murine TLR9 polypeptide. The amount of murine TLR9 which interacts with the murine TLR9 binding agent or the amount which does not bind to the murine TLR9 binding agent may then be quantitated to determine whether the murine TLR9 binding agent binds to murine TLR9.

The murine TLR9 binding agents include molecules of numerous size and type that bind selectively or preferentially to murine TLR9 polypeptides, and complexes of both murine TLR9 polypeptides and their binding partners. These molecules may be derived from a variety of sources. For example, murine TLR9 binding agents can be provided by screening degenerate peptide libraries which can be readily prepared in solution, in immobilized form or as phage display libraries. Combinatorial libraries also can be synthesized of peptides containing one or more amino acids. Libraries further can be synthesized of peptoids and non-peptide synthetic moieties.

Phage display can be particularly effective in identifying binding peptides useful according to the invention. Briefly, one prepares a phage library (using, e.g., m13, fd, or lambda phage), displaying inserts from 4 to about 80 amino acid residues using conventional procedures. The inserts may represent, for example, a completely degenerate or biased array.

One then can select phage-bearing inserts which bind to the murine TLR9 polypeptide. This process can be repeated through several cycles of reselection of phage that bind to the murine TLR9 polypeptide. Repeated rounds lead to enrichment of phage bearing particular sequences. DNA sequence analysis can be conducted to identify the sequences of the expressed polypeptides. The minimal linear portion of the sequence that binds to the murine TLR9 polypeptide can be determined. One can repeat the procedure using a biased library containing inserts containing part or all of the minimal linear portion plus one or more additional degenerate residues upstream or downstream thereof. Yeast two-hybrid screening methods also may be used to identify polypeptides that bind to the murine TLR9 polypeptides. Thus, the murine TLR9 polypeptides of the invention, or a fragment thereof, can be used to screen peptide libraries, including phage display libraries, to identify and select peptide binding partners of the murine TLR9 polypeptides of the invention. Such molecules can be used, as described, for screening assays, for purification protocols, for interfering directly with the functioning of murine TLR9 and for other purposes that will be apparent to those of ordinary skill in the art.

The invention also embraces agents which bind selectively to certain regulatory sequences associated with the murine TLR9 nucleic acid molecules described herein. The agents include polypeptides which bind to transcription and translation regulatory sequences of murine TLR9, and antisense nucleic acids, both of which are described in greater detail below. The agents can inhibit or increase murine TLR9 expression, as well as signaling activity (antagonists and agonists, respectively). Agents which bind selectively to regulatory sequences associated with the murine TLR9 nucleic acid molecules can be identified using methods familiar to those of skill in the art. For example, a promoter region including at least 100, 200, 300, 400, 500, or more nucleotides upstream (5') of the coding region of murine TLR9 can be identified by isolating, from appropriate genomic DNA, such nucleotide sequences using the sequences of SEQ ID NO:1 or SEQ ID NO:2 as primers or as probes, and then inserting the promoter region DNA into an appropriate expression vector so as to control the expression of TLR9 or some other reporter gene, introducing the TLR9 promoter vector into an appropriate host cell, and screening for TLR9 or reporter expression by those cells following their incubation in the presence and absence of various test agents. A reporter gene other than TLR9 can include, for example, an enzyme, a cytokine, a cell surface antigen,

luciferase, chloramphenicol acetyl transferase (CAT), etc. An agent that inhibits expression of TLR9 or the reporter under the control of the TLR9 promoter is classified as a TLR9 expression inhibitor. Conversely, an agent that augments expression of TLR9 or reporter under the control of the TLR9 promoter is classified as a TLR9 expression enhancer. It was discovered according to the instant invention, for example, that the cytokine IL-4 inhibits the expression of TLR9. In this manner it is possible to identify agents that can be administered in conjunction with ISNA, for example by local administration, to enhance response to the ISNA. Such an enhancing effect might be desirable, for example, in the setting of immunization or vaccination. Conversely, it is possible to identify agents that can be administered in conjunction with a ISNA, for example by local administration, to inhibit response to the ISNA. Such an inhibiting response might be desirable, for example, in the setting of gene replacement therapy.

Therefore the invention generally provides efficient methods of identifying pharmacological agents or lead compounds for agents useful in the treatment of conditions associated with TLR9 activity and the compounds and agents so identified. Generally, the screening methods involve assaying for compounds which inhibit or enhance signaling through murine TLR9. Such methods are adaptable to automated, high throughput screening of compounds. Examples of such high throughput screening methods are described in U.S. patents 6,103,479; 6,051,380; 6,051,373; 5,998,152; 5,876,946; 5,708,158; 5,443,791; 5,429,921; and 5,143,854.

A variety of assays for pharmacological agents are provided, including labeled *in vitro* protein binding assays, signaling assays using detectable molecules, etc. For example, protein binding screens are used to rapidly examine the binding of candidate pharmacological agents to a murine TLR9. The candidate pharmacological agents can be derived from, for example, combinatorial peptide or nucleic acid libraries. Convenient reagents for such assays are known in the art. An exemplary cell-based assay of signaling involves contacting a cell having a murine TLR9 with a candidate pharmacological agent under conditions whereby the induction of a detectable molecule can occur. Specific conditions are well known in the art and are described, for example, in Häcker H et al., *J Exp Med* 192:595-600 (2000), and references cited therein. A reduced degree of induction of the detectable molecule in the presence of the candidate pharmacological agent indicates that the candidate pharmacological

agent reduces the signaling activity of murine TLR9. An increased degree of induction of the detectable molecule in the presence of the candidate pharmacological agent indicates that the candidate pharmacological agent increases the signaling activity of murine TLR9.

Murine TLR9 used in the methods of the invention can be added to an assay mixture as an isolated polypeptide (where binding of a candidate pharmaceutical agent is to be measured) or as a cell or other membrane-encapsulated space which includes a murine TLR9 polypeptide. In the latter assay configuration, the cell or other membrane-encapsulated space can contain the murine TLR9 as a polypeptide or as a nucleic acid (e.g., a cell transfected with an expression vector containing a murine TLR9). In the assays described herein, the murine TLR9 polypeptide can be produced recombinantly, isolated from biological extracts, or synthesized *in vitro*. Murine TLR9 polypeptides encompass chimeric proteins comprising a fusion of a murine TLR9 polypeptide with another polypeptide, e.g., a polypeptide capable of providing or enhancing protein-protein binding, enhancing signaling capability, facilitating detection, or enhancing stability of the murine TLR9 polypeptide under assay conditions. A polypeptide fused to a murine TLR9 polypeptide or fragment thereof may also provide means of readily detecting the fusion protein, e.g., by immunological recognition or by fluorescent labeling.

The assay mixture also comprises a candidate pharmacological agent. Typically, a plurality of assay mixtures are run in parallel with different agent concentrations to obtain a different response to the various concentrations. Typically, one of these concentrations serves as a negative control, i.e., at zero concentration of agent or at a concentration of agent below the limits of assay detection. Candidate pharmaceutical agents encompass numerous chemical classes, although typically they are organic compounds. Preferably, the candidate pharmacological agents are small organic compounds, i.e., those having a molecular weight of more than 50 yet less than about 2500. Polymeric candidate agents can have higher molecular weights, e.g., oligonucleotides in the range of about 2500 to about 12,500. Candidate agents comprise functional chemical groups necessary for structural interactions with polypeptides, and may include at least an amine, carbonyl, hydroxyl or carboxyl group, preferably at least two of the functional chemical groups and more preferably at least three of the functional chemical groups. The candidate agents can comprise cyclic carbon or heterocyclic structure and/or aromatic or polyaromatic structures substituted with one or more

of the above-identified functional groups. Candidate agents also can be biomolecules such as nucleic acids, peptides, saccharides, fatty acids, sterols, isoprenoids, purines, pyrimidines, derivatives or structural analogs of the above, or combinations thereof and the like. Where the agent is a nucleic acid, the agent typically is a DNA or RNA molecule, although modified nucleic acids having non-natural bonds or subunits are also contemplated.

Candidate agents are obtained from a wide variety of sources, including libraries of natural, synthetic, or semisynthetic compounds, or any combination thereof. For example, numerous means are available for random and directed synthesis of a wide variety of organic compounds and biomolecules, including expression of randomized oligonucleotides, synthetic organic combinatorial libraries, phage display libraries of random peptides, and the like. Alternatively, libraries of natural compounds in the form of bacterial, fungal, plant and animal extracts are available or readily produced. Additionally, natural and synthetically produced libraries and compounds can be readily modified through conventional chemical, physical, and biochemical means. Further, known pharmacological agents may be subjected to directed or random chemical modifications such as acylation, alkylation, esterification, amidification, etc., to produce structural analogs of the agents.

Therefore, a source of candidate agents are libraries of molecules based on known TLR9 ligands, e.g., CpG oligonucleotides shown herein to interact with TLR9, in which the structure of the ligand is changed at one or more positions of the molecule to contain more or fewer chemical moieties or different chemical moieties. The structural changes made to the molecules in creating the libraries of analog inhibitors can be directed, random, or a combination of both directed and random substitutions and/or additions. One of ordinary skill in the art in the preparation of combinatorial libraries can readily prepare such libraries based on existing TLR9 ligands.

A variety of other reagents also can be included in the mixture. These include reagents such as salts, buffers, neutral proteins (e.g., albumin), detergents, etc. which may be used to facilitate optimal protein-protein and/or protein-nucleic acid binding. Such a reagent may also reduce non-specific or background interactions of the reaction components. Other reagents that improve the efficiency of the assay such as protease inhibitors, nuclease inhibitors, antimicrobial agents, and the like may also be used.

The mixture of the foregoing assay materials is incubated under conditions whereby,

but for the presence of the candidate pharmacological agent, the murine TLR9 mediates TLR/IL-1R signaling. For determining the binding of a candidate pharmaceutical agent to a murine TLR9, the mixture is incubated under conditions which permit binding. The order of addition of components, incubation temperature, time of incubation, and other parameters of the assay may be readily determined. Such experimentation merely involves optimization of the assay parameters, not the fundamental composition of the assay. Incubation temperatures typically are between 4°C and 40°C. Incubation times preferably are minimized to facilitate rapid, high throughput screening, and typically are between 1 minute and 10 hours.

After incubation, the level of signaling or the level of specific binding between the murine TLR9 polypeptide and the candidate pharmaceutical agent is detected by any convenient method available to the user. For cell-free binding type assays, a separation step is often used to separate bound from unbound components. The separation step may be accomplished in a variety of ways. For example, separation can be accomplished in solution, or, conveniently, at least one of the components is immobilized on a solid substrate, from which the unbound components may be easily separated. The solid substrate can be made of a wide variety of materials and in a wide variety of shapes, e.g., microtiter plate, microbead, dipstick, resin particle, etc. The substrate preferably is chosen to maximize signal-to-noise ratios, primarily to minimize background binding, as well as for ease of separation and cost.

Separation may be effected for example, by removing a bead or dipstick from a reservoir, emptying or diluting a reservoir such as a microtiter plate well, rinsing a bead, particle, chromatographic column or filter with a wash solution or solvent. The separation step preferably includes multiple rinses or washes. For example, when the solid substrate is a microtiter plate, the wells may be washed several times with a washing solution, which typically includes those components of the incubation mixture that do not participate in specific bindings such as salts, buffer, detergent, non-specific protein, etc. Where the solid substrate is a magnetic bead, the beads may be washed one or more times with a washing solution and isolated using a magnet.

Detection may be effected in any convenient way for cell-based assays such as measurement of an induced polypeptide within, on the surface of, or secreted by the cell. Examples of detection methods useful in such cell-based assays include fluorescence-activated cell sorting (FACS) analysis, bioluminescence, fluorescence, enzyme-linked

immunosorbent assay (ELISA), reverse transcriptase-polymerase chain reaction (RT-PCR), and the like.

A variety of methods may be used to detect the label, depending on the nature of the label and other assay components. For example, the label may be detected while bound to the solid substrate or subsequent to separation from the solid substrate. Labels may be directly detected through optical or electron density, radioactive emissions, nonradiative energy transfers, etc., or indirectly detected with antibody conjugates, streptavidin-biotin conjugates, etc. Methods for detecting the labels are well known in the art.

The murine TLR9 binding agent may also be an antibody or a functionally active antibody fragment. Antibodies are well known to those of ordinary skill in the science of immunology. As used herein, the term "antibody" means not only intact antibody molecules but also fragments of antibody molecules retaining specific target binding ability. Such fragments are also well known in the art and are regularly employed both *in vitro* and *in vivo*. In particular, as used herein, the term "antibody" means not only intact immunoglobulin molecules but also the well-known active fragments F(ab')₂ and Fab. F(ab')₂ and Fab fragments which lack the Fc fragment of intact antibody clear more rapidly from the circulation and may have less non-specific tissue binding than an intact antibody (Wahl RL et al., *J Nucl Med* 24:316-325 (1983)).

Monoclonal antibodies may be made by any of the methods known in the art utilizing murine TLR9, or a fragment thereof, as an immunogen. Alternatively the antibody may be a polyclonal antibody specific for murine TLR9 which inhibits murine TLR9 activity. The preparation and use of polyclonal antibodies are also known to one of ordinary skill in the art.

Significantly, as is well known in the art, only a small portion of an antibody molecule, the paratope, is involved in the binding of the antibody to its epitope (see, in general, Clark, W.R. (1986) *The Experimental Foundations of Modern Immunology*, Wiley & Sons, Inc., New York; Roitt, I. (1991) *Essential Immunology*, 7th Ed., Blackwell Scientific Publications, Oxford). The pFc' and Fc regions, for example, are effectors of the complement cascade but are not involved in antigen binding. An antibody from which the pFc' region has been enzymatically cleaved, or which has been produced without the pFc' region, designated an F(ab')₂ fragment, retains both of the antigen binding sites of an intact antibody. Similarly, an antibody from which the Fc region has been enzymatically cleaved, or which has been

produced without the Fc region, designated an Fab fragment, retains one of the antigen binding sites of an intact antibody molecule. Proceeding further, Fab fragments consist of a covalently bound antibody light chain and a portion of the antibody heavy chain denoted Fd. The Fd fragments are the major determinant of antibody specificity (a single Fd fragment may be associated with up to ten different light chains without altering antibody specificity) and Fd fragments retain epitope-binding ability in isolation.

Within the antigen-binding portion of an antibody, as is well-known in the art, there are complementarity determining regions (CDRs), which directly interact with the epitope of the antigen, and framework regions (FRs), which maintain the tertiary structure of the paratope (see, in general, Clark, 1986; Roitt, 1991). In both the heavy chain Fd fragment and the light chain of IgG immunoglobulins, there are four framework regions (FR1 through FR4) separated respectively by three complementarity determining regions (CDR1 through CDR3). The CDRs, and in particular the CDR3 regions, and more particularly the heavy chain CDR3, are largely responsible for antibody specificity.

The sequences of the antigen-binding Fab' portion of the anti-murine TLR9 monoclonal antibodies identified as being useful according to the invention in the assays provided above, as well as the relevant FR and CDR regions, can be determined using amino acid sequencing methods that are routine in the art. It is well established that non-CDR regions of a mammalian antibody may be replaced with corresponding regions of non-specific or hetero-specific antibodies while retaining the epitope specificity of the original antibody. This technique is useful for the development and use of "humanized" antibodies in which non-human CDRs are covalently joined to human FR and/or Fc/pFc' regions to produce a functional antibody. Techniques to humanize antibodies are particularly useful when non-human animal (e.g., murine) antibodies which inhibit murine TLR9 activity are identified. These non-human animal antibodies can be humanized for use in the treatment of a human subject in the methods according to the invention. Examples of methods for humanizing a murine antibody are provided in U.S. patents 4,816,567, 5,225,539, 5,585,089, 5,693,762 and 5,859,205. Other antibodies, including fragments of intact antibodies with antigen-binding ability, are often referred to as "chimeric" antibodies.

Thus, as will be apparent to one of ordinary skill in the art, the present invention also provides for F(ab')₂ and Fab fragments of an anti-murine TLR9 monoclonal antibody;

chimeric antibodies in which the Fc and/or FR and/or CDR1 and/or CDR2 and/or light chain CDR3 regions of an anti-murine TLR9 antibody have been replaced by homologous human or non-human sequences; chimeric F(ab')₂ fragment antibodies in which the FR and/or CDR1 and/or CDR2 and/or light chain CDR3 regions of an anti-murine TLR9 antibody have been replaced by homologous human or non-human sequences; and chimeric Fab fragment antibodies in which the FR and/or CDR1 and/or CDR2 and/or light chain CDR3 regions have been replaced by homologous human or non-human sequences.

According to the invention murine TLR9 inhibitors also include "dominant negative" polypeptides derived from SEQ ID NO:3. A dominant negative polypeptide is an inactive variant of a polypeptide, which, by interacting with the cellular machinery, displaces an active polypeptide from its interaction with the cellular machinery or competes with the active polypeptide, thereby reducing the effect of the active polypeptide. For example, a dominant negative receptor which binds a ligand but does not transmit a signal in response to binding of the ligand can reduce the biological effect of expression of the receptor. As shown in the Examples below, TLR9 polypeptides which incorporate the substitution of histidine for proline at aa 915 (P915H mutation) are functionally inactive and are dominant negative with respect to the native TLR9 polypeptide.

The end result of the expression of a dominant negative murine TLR9 polypeptide of the invention in a cell is a reduction in TLR9 activity such as signaling through the TIR pathway. One of ordinary skill in the art can assess the potential for a dominant negative variant of a murine TLR9 polypeptide and, using standard mutagenesis techniques, create one or more dominant negative variant polypeptides. For example, given the teachings contained herein of a murine TLR9 polypeptide, one of ordinary skill in the art can modify the sequence of the murine TLR9 polypeptide by site-specific mutagenesis, scanning mutagenesis, partial gene deletion or truncation, and the like. See, e.g., U.S. Patent No. 5,580,723 and *Molecular Cloning: A Laboratory Manual*, J. Sambrook, et al., eds., Second Edition, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York, 1989. The skilled artisan then can test the population of mutagenized polypeptides for diminution in murine TLR9 activity and/or for retention of such an activity. Other similar methods for creating and testing dominant negative variants of a murine TLR9 polypeptide will be apparent to one of ordinary skill in the art.

Each of the compositions according to this aspect of the invention is useful for a variety of therapeutic and non-therapeutic purposes. For example, the murine TLR9 nucleic acids of the invention are useful as oligonucleotide probes. Such oligonucleotide probes can be used herein to identify genomic or cDNA library clones possessing an identical or substantially similar nucleic acid sequence. A suitable oligonucleotide or set of oligonucleotides, which is capable of hybridizing under stringent hybridization conditions to the desired sequence, a variant or fragment thereof, or an anti-sense complement of such an oligonucleotide or set of oligonucleotides, can be synthesized by means well known in the art (see, for example, *Synthesis and Application of DNA and RNA*, S.A. Narang, ed., 1987, Academic Press, San Diego, CA) and employed as a probe to identify and isolate the desired sequence, variant or fragment thereof by techniques known in the art. Techniques of nucleic acid hybridization and clone identification are disclosed by Sambrook, et al., *Molecular Cloning: A Laboratory Manual*, J. Sambrook, et al., eds., Second Edition, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York, 1989.

To facilitate the detection of a desired nucleic acid sequence, or variant or fragment thereof, whether for cloning purposes or for the mere detection of the presence of the sequence, the above-described probes may be labeled with a detectable group. Such a detectable group may be any material having a detectable physical or chemical property. Such materials have been well developed in the field of nucleic acid hybridization and, in general, many labels useful in such methods can be applied to the present invention. Particularly useful are radioactive labels. Any radioactive label may be employed which provides for an adequate signal and has a sufficient half-life. If single stranded, the oligonucleotide may be radioactively labeled using kinase reactions. Alternatively, oligonucleotides are also useful as nucleic acid hybridization probes when labeled with a non-radioactive marker such as biotin, an enzyme or a fluorescent group. See, for example, Leary JJ et al., *Proc Natl Acad Sci USA* 80:4045 (1983); Renz M et al., *Nucleic Acids Res* 12:3435 (1984); and Renz M, *EMBO J* 6:817 (1983).

Additionally, complements of the murine TLR9 nucleic acids can be useful as antisense oligonucleotides, e.g., by delivering the antisense oligonucleotide to an animal to induce a murine TLR9 "knockout" phenotype. The administration of antisense RNA probes to block gene expression is discussed in Lichtenstein C, *Nature* 333:801-802 (1988).

Alternatively, the murine TLR9 nucleic acid of the invention can be used to prepare a non-human transgenic animal. A "transgenic animal" is an animal having cells that contain DNA which has been artificially inserted into a cell, which DNA becomes part of the genome of the animal which develops from that cell. Preferred transgenic animals are primates, mice, rats, cows, pigs, horses, goats, sheep, dogs and cats. Animals suitable for transgenic experiments can be obtained from standard commercial sources such as Charles River (Wilmington, MA), Taconic (Germantown, NY), Harlan (Indianapolis, IN), etc. Transgenic animals having a particular property associated with a particular disease can be used to study the effects of a variety of drugs and treatment methods on the disease, and thus serve as genetic models for the study of a number of human diseases. The invention, therefore, contemplates the use of murine TLR9 knockout and transgenic animals as models for the study of disorders involving TLR9-mediated signaling. A variety of methods known to one of ordinary skill in the art are available for the production of transgenic animals associated with this invention.

Inactivation or replacement of the endogenous TLR9 gene can be achieved by a homologous recombination system using embryonic stem cells. The resultant transgenic non-human mammals having a TLR9^{-/-} knockout phenotype may be made transgenic for the murine TLR9 and used as a model for screening compounds as modulators (agonists or antagonists/inhibitors) of the murine TLR9. In this manner, such therapeutic drugs can be identified.

Additionally, a normal or mutant version of murine TLR9 can be inserted into the germ line to produce transgenic animals which constitutively or inducibly express the normal or mutant form of murine TLR9. These animals are useful in studies to define the role and function of murine TLR9 in cells.

Generally, doses of active compounds would be from about 0.01 mg/kg per day to 1000 mg/kg per day. It is expected that doses ranging from 50-500 mg/kg will be suitable and in one or several administrations per day. Lower doses will result from other forms of administration, such as intravenous administration. In the event that a response in a subject is insufficient at the initial doses applied, higher doses (or effectively higher doses by a different, more localized delivery route) may be employed to the extent that patient tolerance permits. Multiple doses per day are contemplated to achieve appropriate systemic levels of

compound, although fewer doses typically will be given when compounds are prepared as slow release or sustained release medications.

The antagonists, agonists, nucleic acids, and polypeptides of murine TLR9 useful according to the invention may be combined, optionally, with a pharmaceutically acceptable carrier. Thus the invention also provides pharmaceutical compositions and a method for preparing the pharmaceutical compositions which contain compositions of this aspect of the invention. The pharmaceutical compositions include any one or combination of the antagonists, agonists, nucleic acids and polypeptides of murine TLR9 useful according to the invention and, optionally, a pharmaceutically acceptable carrier. Each pharmaceutical composition is prepared by selecting an antagonist, agonist, nucleic acid or polypeptide of murine TLR9 useful according to the invention, as well as any combination thereof, and, optionally, combining it with a pharmaceutically acceptable carrier.

The term "pharmaceutically acceptable carrier" as used herein means one or more compatible solid or liquid fillers, diluents or encapsulating substances which are suitable for administration into a human. The term "carrier" denotes an organic or inorganic ingredient, natural or synthetic, with which the active ingredient is combined to facilitate the application. The components of the pharmaceutical compositions also are capable of being co-mingled with the molecules of the present invention, and with each other, in a manner such that there is no interaction which would substantially impair the desired pharmaceutical efficacy.

The pharmaceutical compositions may contain suitable buffering agents, including, without limitation: acetic acid in a salt; citric acid in a salt; and phosphoric acid in a salt.

The pharmaceutical compositions also may contain, optionally, suitable preservatives, such as benzalkonium chloride, chlorobutanol, parabens, and thimerosal.

When administered, the pharmaceutical preparations of the invention are applied in pharmaceutically acceptable amounts and in pharmaceutically acceptable compositions. Such preparations may routinely contain salts, buffering agents, preservatives, compatible carriers, and optionally other therapeutic agents. When used in medicine, the salts should be pharmaceutically acceptable, but non-pharmaceutically acceptable salts may conveniently be used to prepare pharmaceutically acceptable salts thereof and are not excluded from the scope of the invention. Such pharmacologically and pharmaceutically acceptable salts include, but are not limited to, those prepared from the following acids: hydrochloric, hydrobromic,

sulfuric, nitric, phosphoric, maleic, acetic, salicylic, citric, formic, malonic, succinic, and the like. Also, pharmaceutically acceptable salts can be prepared as alkaline metal or alkaline earth salts, such as sodium, potassium or calcium salts.

A variety of administration routes are available. The particular mode selected will depend, of course, upon the particular compound selected, the severity of the condition being treated, and the dosage required for therapeutic efficacy. The methods of the invention, generally speaking, may be practiced using any mode of administration that is medically acceptable, meaning any mode that produces effective levels of the active compounds without causing clinically unacceptable adverse effects. Such modes of administration include oral, rectal, topical, nasal, intradermal, or parenteral routes. The term "parenteral" includes, without limitation, subcutaneous, transdermal, intravenous, intra-arterial, intrathecal, intramuscular, intraperitoneal, mucosal (apart from gastrointestinal mucosa), pulmonary, intralesional, and infusion.

The pharmaceutical compositions may conveniently be presented in unit dosage form and may be prepared by any of the methods well-known in the art of pharmacy. All methods include the step of bringing the active agent into association with a carrier which constitutes one or more accessory ingredients. In general, the compositions are prepared by uniformly and intimately bringing the active compound into association with a liquid carrier, a finely divided solid carrier, or both, and then, if necessary, shaping the product.

Compositions suitable for oral administration may be presented as discrete units, such as capsules, tablets, lozenges, each containing a predetermined amount of the active compound. Other compositions include suspensions in aqueous liquids or non-aqueous liquids such as a syrup, elixir or an emulsion.

Compositions suitable for parenteral administration conveniently comprise a sterile aqueous preparation of the antagonists, agonists, nucleic acids, or polypeptides of murine TLR9, which is preferably isotonic with the blood of the recipient. This aqueous preparation may be formulated according to known methods using suitable dispersing or wetting agents and suspending agents. The sterile injectable preparation also may be a sterile injectable solution or suspension in a non-toxic parenterally acceptable diluent or solvent, for example, as a solution in 1,3-butane diol. Among the acceptable vehicles and solvents that may be employed are water, Ringer's solution, and isotonic sodium chloride solution. In addition,

sterile, fixed oils are conventionally employed as a solvent or suspending medium. For this purpose any bland fixed oil may be employed including synthetic mono- or di-glycerides. In addition, fatty acids such as oleic acid may be used in the preparation of injectables. Carrier formulation suitable for oral, subcutaneous, intravenous, intrathecal, intramuscular, etc. administrations can be found in Remington's Pharmaceutical Sciences, Mack Publishing Co., Easton, PA.

Other delivery systems can include time-release, delayed release or sustained release delivery systems such as the biological/chemical vectors is discussed above. Such systems can avoid repeated administrations of the active compound, increasing convenience to the subject and the physician. Many types of release delivery systems are available and known to those of ordinary skill in the art. Use of a long-term sustained release implant may be desirable. Long-term release, as used herein, means that the implant is constructed and arranged to delivery therapeutic levels of the active ingredient for at least 30 days, and preferably 60 days. Long-term sustained release implants are well-known to those of ordinary skill in the art and include some of the release systems described above.

In another aspect the invention involves the identification of cDNAs encoding mouse TLR7 and mouse TLR8, referred to herein as murine TLR7 and murine TLR8 and, equivalently, mTLR7 and mTLR8, respectively. The nucleotide sequence of the cDNA for murine TLR7 is presented as SEQ ID NO:173, the coding region of the cDNA for murine TLR7 is presented as SEQ ID NO:174, and the amino acid sequence of the murine TLR7 is presented as SEQ ID NO:175. The closely related human TLR7 (equivalently, hTLR7) was previously deposited in GenBank under accession numbers AF245702 and AF240467. The nucleotide sequence of the cDNA for murine TLR7 presented as SEQ ID NO:173 is 3357 nucleotides long and includes the ORF spanning bases 117-3266, presented as SEQ ID NO:174, which spans 3150 nucleotides (excluding the stop codon). The amino acid sequence of the murine TLR7 presented as SEQ ID NO:175 is 1050 amino acids long.

The nucleotide sequence of the cDNA for murine TLR8 is presented as SEQ ID NO:190, the coding region of the cDNA for murine TLR8 is presented as SEQ ID NO:191, and the amino acid sequence of the murine TLR8 is presented as SEQ ID NO:192. The closely related human TLR8 (equivalently, hTLR8) was previously deposited in GenBank under accession numbers AF245703 and AF246971.

Like both human and murine TLR9, human TLR7 and human TLR8 each contains one CXXC motif and one MBD motif. The hTLR7 CXXC motif contains amino acids 258-273, and the hTLR8 CXXC motif contains amino acids 255-270.

5	CXXC motif:	GNCXXCXXXXXXXXCXXC	SEQ ID NO:196
	hTLR9:	GNCRRCDHAPNPCMEC	SEQ ID NO:197
	mTLR9:	GNCRRCDHAPNPCMIC	SEQ ID NO:198
	hTLR7:	GNCPRCYNAPFPCAPC	SEQ ID NO:199
10	mTLR7:	GNCPRCYNVPYPCTPC	SEQ ID NO:200
	hTLR8:	GNCPRCFNAPFPCVPC	SEQ ID NO:201
	mTLR8:	GNCPRCYNAPFPCTPC	SEQ ID NO:202

Also like human and murine TLR9, human TLR7 and TLR8 also have a single MBD motif. The the hTLR7 MBD motif spans amino acids 545-575, and the hTLR8 MBD motif amino acids spans 533-563.

MBD motif

	MBD-1	R-XXXXXXXX-R-X-D-X-Y-XXXXXXXXXX-R-S-XXXXXX-Y	SEQ ID NO:125
20	hTLR9	Q-XXXXXXXX-K-X-D-X-Y-XXXXXXXXXX-R-L-XXXXXX-Y	SEQ ID NO:126
	mTLR9	Q-XXXXXXXX-K-X-D-X-Y-XXXXXXXXXX-Q-L-XXXXXX-Y	SEQ ID NO:127
	hTLR7	R-XXXXXXXX-R-X-D-X-L-XXXXXXXXXX-K-L-XXXXXX-S	SEQ ID NO:203
	mTLR7	R-XXXXXXXX-R-X-D-X-L-XXXXXXXXXX-S-L-XXXXXX-S	SEQ ID NO:204
	hTLR8	K-XXXXXXXX-R-X-D-X-D-XXXXXXXXXX-D-L-XXXXXX-Y	SEQ ID NO:205
25	mTLR8	K-XXXXXXXX-R-X-D-X-D-XXXXXXXXXX-D-L-XXXXXX-H	SEQ ID NO:206
	hTLR7	R-YLDFSNN-R-L-D-L-L-HSTAFEELH-K-L-EVLDIS-S	SEQ ID NO:212
	mTLR7	R-YLDFSNN-R-L-D-L-L-YSTAFEELQ-S-L-EVLDSL-S	SEQ ID NO:213
30	hTLR8	K-YLDLTNN-R-L-D-F-D-NASALTELS-D-L-EVLDSL-Y	SEQ ID NO:214
	mTLR8	K-YLDLTNN-R-L-D-F-D-DNNAFSDLH-D-L-EVLDSL-H	SEQ ID NO:215

The core D-X-Y in the MBD motif is involved in CpG binding of the MBD-1 protein and is conserved in TLR9 but only partially conserved in TLR8 and TLR7 (Y to D or L). The other mismatches are highly or moderately conserved; example R to K, Q, or D. These changes could explain MBD-1 as a methyl-CpG binder and TLR9 as a binder for CpG-DNA. The modification in the core sequence (D-X-Y) in hTLR7 (D-X-L) and TLR8 (D-X-D) is likely a structural basis for the recognition of different nucleic acid motifs. Combined with the presence of a CXXC domain TLR7 and TLR8 appear certainly to be nucleic acid binding receptors relevant to the innate immune system and thus clinical value.

The invention involves in one aspect murine TLR7 and murine TLR8 nucleic acids and polypeptides, as well as therapeutics relating thereto. The invention also embraces isolated functionally equivalent variants, useful analogs and fragments of the foregoing murine TLR7 and murine TLR8 nucleic acids and polypeptides; complements of the foregoing murine TLR7 and murine TLR8 nucleic acids; and molecules which selectively bind the foregoing murine TLR7 and murine TLR8 nucleic acids and polypeptides.

The murine TLR7 and murine TLR8 nucleic acids and polypeptides of the invention are isolated. The term "isolated," with respect to murine TLR7 and murine TLR8 nucleic acids and polypeptides, has the same meaning as used elsewhere herein.

As used herein a murine TLR7 nucleic acid refers to an isolated nucleic acid molecule which codes for a murine TLR7 polypeptide. Such nucleic acid molecules code for murine TLR7 polypeptides which include the sequence of SEQ ID NO:175 and fragments thereof. The nucleic acid molecules include the nucleotide sequences of SEQ ID NO:173, SEQ ID NO:174, and nucleotide sequences which differ from the sequences of SEQ ID NO:173 and SEQ ID NO:174 in codon sequence due to the degeneracy of the genetic code.

Also as used herein a murine TLR8 nucleic acid refers to an isolated nucleic acid molecule which codes for a murine TLR8 polypeptide. Such nucleic acid molecules code for murine TLR8 polypeptides which include the sequences of SEQ ID NO:193, and fragments thereof. The nucleic acid molecules include the nucleotide sequences of SEQ ID NO:190, SEQ ID NO:191, and nucleotide sequences which differ from the sequences of SEQ ID NO:190 and SEQ ID NO:191 in codon sequence due to the degeneracy of the genetic code.

The murine TLR7 and murine TLR8 nucleic acids of the invention also include alleles

as well as fragments of the foregoing nucleic acids. Such fragments can be used, for example, as probes in hybridization assays and as primers in a polymerase chain reaction. Preferred murine TLR7 nucleic acids include the nucleic acid sequence of SEQ ID NO:173 and SEQ ID NO:174. Preferred murine TLR8 nucleic acids include the nucleic acid sequence of SEQ ID NO:190 and SEQ ID NO:191. Complements of the foregoing nucleic acids also are embraced by the invention.

As used herein a murine TLR7 nucleic acid or murine TLR7 polypeptide also embraces homologues and alleles of murine TLR7. Likewise, as used herein a murine TLR8 nucleic acid or murine TLR8 polypeptide also embraces homologues and alleles of murine TLR8. Homologues and alleles of murine TLR7 and murine TLR8 comply with the degrees of nucleotide and amino acid identity as previously set forth herein in reference to homologues and alleles of murine TLR9.

Alleles of the murine TLR7 and murine TLR8 nucleic acids of the invention can be identified by conventional techniques. For example, alleles of murine TLR7 can be isolated by hybridizing a probe which includes at least a fragment of SEQ ID NO:173 or SEQ ID NO:174 under stringent conditions with a cDNA library and selecting positive clones. Thus, an aspect of the invention is those nucleic acid sequences which code for murine TLR7 polypeptides and which hybridize to a nucleic acid molecule consisting of SEQ ID NO:173 or SEQ ID NO:174 under stringent conditions. Likewise, an aspect of the invention is those nucleic acid sequences which code for murine TLR8 polypeptides and which hybridize to a nucleic acid molecule consisting of SEQ ID NO:190 or SEQ ID NO:191 under stringent conditions. Stringent conditions in this context has the same meaning as described elsewhere herein, including the use of a suitable hybridization buffer and a temperature of about 65°C.

In screening for murine TLR7 or murine TLR8 nucleic acids, a Southern blot may be performed using the stringent conditions previously described herein, together with a radioactive probe. After washing the membrane to which the DNA is finally transferred, the membrane can be placed against X-ray film to detect the radioactive signal. Corresponding non-radioactive methods are also well known in the art and can be used to similar effect.

The murine TLR7 and murine TLR8 nucleic acids of the invention also include degenerate nucleic acids which include alternative codons to those present in the native materials, as previously described herein.

The invention also provides modified nucleic acid molecules which include additions, substitutions and deletions of one or more nucleotides. The modified nucleic acid molecules according to this aspect of the invention exclude fully native human TLR7 (SEQ ID NO:168, SEQ ID NO:169, GenBank Accession No. AF245702, and GenBank Accession No. AF240467) and fully native human TLR8 nucleic acid molecules (SEQ ID NO:182, SEQ ID NO:183, GenBank Accession No. AF245703, and GenBank Accession No. AF246971). In preferred embodiments, these modified nucleic acid molecules and/or the polypeptides they encode retain at least one activity or function of the unmodified nucleic acid molecule and/or the polypeptides, such as signaling activity, etc. In certain embodiments, the modified nucleic acid molecules encode modified polypeptides, preferably polypeptides having conservative amino acid substitutions as are described elsewhere herein. The modified nucleic acid molecules are structurally related to the unmodified nucleic acid molecules and in preferred embodiments are sufficiently structurally related to the unmodified nucleic acid molecules so that the modified and unmodified nucleic acid molecules hybridize under stringent conditions known to one of skill in the art.

The invention also provides isolated fragments of nucleotide sequences for murine TLR7 (SEQ ID NO:173 and SEQ ID NO:174) and for murine TLR8 (SEQ ID NO:190 and SEQ ID NO:191). The fragments can be used as probes in Southern blot assays to identify such nucleic acids, or can be used in amplification assays such as those employing PCR. Smaller fragments are those comprising 12, 13, 14, 15, 16, 17, 18, 20, 22, 25, 30, 40, 50, or 75 nucleotides, and every integer therebetween, and are useful, e.g., as primers for nucleic acid amplification procedures. As known to those skilled in the art, larger probes such as 200, 250, 300, 400 or more nucleotides are preferred for certain uses such as Southern blots, while smaller fragments will be preferred for uses such as PCR. Fragments also can be used to produce fusion proteins for generating antibodies or determining binding of the polypeptide fragments. Likewise, fragments can be employed to produce non-fused fragments of the murine TLR7 and murine TLR8 polypeptides, useful, for example, in the preparation of antibodies, in immunoassays, and the like. The foregoing nucleic acid fragments further can be used as antisense molecules to inhibit the expression of murine TLR7 and murine TLR8 nucleic acids and polypeptides.

The invention also includes functionally equivalent variants of the murine TLR7 and murine TLR8, which include variant nucleic acids and polypeptides which retain one or more of the functional properties of the murine TLR7 and murine TLR8. Preferably such variants include the murine-specific N-terminal domain.

Functionally equivalent variants also include a murine TLR7 or murine TLR8 which has had a portion (e.g., of the N-terminus) removed or replaced by a similar domain from another TLR (e.g., a "domain-swapping" variant). Examples of such domain-swapping variants include those involving swapping a TLR7 domain from another species and swapping a TLR domain from another TLR.

Other functionally equivalent variants will be known to one of ordinary skill in the art, as will be methods for preparing such variants. The activity of a functionally equivalent variant can be determined using the methods provided herein, and in references that have described assays using other TLRs and TLRs of other species. Such variants are useful, *inter alia*, for evaluating bioavailability of drugs, in assays for identification of compounds which bind to and/or regulate the signaling function of the murine TLR7 and murine TLR8, and for determining the portions of the murine TLR7 and murine TLR8 which are required for signaling activity.

Variants which are non-functional also can be prepared as described above. Such variants are useful, for example, as negative controls in experiments testing TLR7 and TLR8 signaling activity. Examples of non-functional variants include those incorporating a truncation or mutation of amino acids deemed critical to ligand binding or signaling activity.

In certain embodiments a murine TLR7 or murine TLR8 nucleic acid is operably linked to a gene expression sequence which can direct the expression of the murine TLR7 or murine TLR8 nucleic acid within a eukaryotic or prokaryotic cell. The terms "gene expression sequence" and "operably linked" are as previously described herein.

The murine TLR7 and murine TLR8 nucleic acid molecules and the murine TLR7 and murine TLR8 polypeptides of the invention can be delivered to a eukaryotic or prokaryotic cell alone or in association with a vector. As applied to murine TLR7 and murine TLR8 nucleic acid molecules, a "vector" is any vehicle capable of facilitating: (1) delivery of a murine TLR7 or murine TLR8 nucleic acid or polypeptide to a target cell, (2) uptake of a murine TLR7 or murine TLR8 nucleic acid or polypeptide by a target cell, or (3) expression

of a murine TLR7 or murine TLR8 nucleic acid molecule or polypeptide in a target cell.

In addition to the biological vectors, chemical/physical vectors may be used to deliver a murine TLR7 or murine TLR8 nucleic acid or polypeptide to a target cell and facilitate uptake thereby. As used herein with respect to a murine TLR7 or murine TLR8 nucleic acid or polypeptide, a "chemical/physical vector" refers to a natural or synthetic molecule, other than those derived from bacteriological or viral sources, capable of delivering the isolated murine TLR7 or murine TLR8 nucleic acid or polypeptide to a cell.

Other exemplary compositions that can be used to facilitate uptake by a target cell of the murine TLR7 or murine TLR8 nucleic acids include calcium phosphate and other chemical mediators of intracellular transport, microinjection compositions, electroporation and homologous recombination compositions (e.g., for integrating a murine TLR7 or murine TLR8 nucleic acid into a preselected location within a target cell chromosome).

It will also be recognized that the invention embraces the use of the murine TLR7 and murine TLR8 cDNA sequences in expression vectors to transfect host cells and cell lines, be these prokaryotic (e.g., *E. coli*), or eukaryotic (e.g., 293 fibroblast cells (ATCC, CRL-1573), MonoMac-6, THP-1, U927, CHO cells, COS cells, yeast expression systems and recombinant baculovirus expression in insect cells). Especially useful are mammalian cells such as human, pig, goat, primate, rodent, guinea pig, etc. They may be of a wide variety of tissue types, and include primary cells and cell lines. The expression vectors require that the pertinent sequence, i.e., those nucleic acids described *supra*, be operably linked to a promoter.

The invention also provides isolated murine TLR7 and isolated murine TLR8 polypeptides which include the amino acid sequences of SEQ ID NO:175, SEQ ID NO:192, and fragments thereof, encoded by the murine TLR7 and murine TLR8 nucleic acids described above. Murine TLR7 and murine TLR8 polypeptides also embrace alleles, functionally equivalent variants and analogs (those non-allelic polypeptides which vary in amino acid sequence from the disclosed murine TLR7 and murine TLR8 polypeptides by 1, 2, 3, 4, 5, or more amino acids) provided that such polypeptides retain murine TLR7 or murine TLR8 activity. Non-functional variants also are embraced by the invention; these are useful as antagonists of TLR7 and TLR8 signaling function, as negative controls in assays, and the like. Such alleles, variants, analogs and fragments are useful, for example, alone or as fusion proteins for a variety of purposes including as a component of assays.

The invention also embraces variants of the murine TLR7 and murine TLR8 polypeptides described above. Modifications which create a murine TLR7 variant or murine TLR8 variant can be made to a murine TLR7 or murine TLR8 polypeptide for a variety of reasons, including 1) to reduce or eliminate an activity of a murine TLR7 or murine TLR8 polypeptide, such as signaling; 2) to enhance a property of a murine TLR7 or murine TLR8 polypeptide, such as signaling, binding affinity for nucleic acid ligand or other ligand molecule, protein stability in an expression system, or the stability of protein-protein binding; 3) to provide a novel activity or property to a murine TLR7 or murine TLR8 polypeptide, such as addition of an antigenic epitope or addition of a detectable moiety, e.g., luciferase, FLAG peptide, GFP; 4) to establish that an amino acid substitution does or does not affect molecular signaling activity; or 5) reduce immunogenicity. Modifications to a murine TLR7 or murine TLR8 polypeptide are typically made to the nucleic acid which encodes the murine TLR7 or murine TLR8 polypeptide, and can include deletions, point mutations, truncations, amino acid substitutions and additions of amino acids or non-amino acid moieties. Alternatively, modifications can be made directly to the polypeptide, such as by cleavage, addition of a linker molecule, addition of a detectable moiety (for example, biotin, fluorophore, radioisotope, enzyme, or peptide), addition of a fatty acid, and the like.

Modifications also embrace fusion proteins comprising all or part of the murine TLR7 or murine TLR8 amino acid sequence.

Variants include murine TLR7 and murine TLR8 polypeptides which are modified specifically to alter a feature of each polypeptide unrelated to its physiological activity. For example, cysteine residues can be substituted or deleted to prevent unwanted disulfide linkages. Similarly, certain amino acids can be changed to enhance expression of a murine TLR7 or murine TLR8 polypeptide by eliminating proteolysis by proteases in an expression system (e.g., dibasic amino acid residues in yeast expression systems in which KEX2 protease activity is present).

Mutations of a nucleic acid which encode a murine TLR7 or murine TLR8 polypeptide preferably preserve the amino acid reading frame of the coding sequence, and preferably do not create regions in the nucleic acid which are likely to hybridize to form secondary structures, such as hairpins or loops, which can be deleterious to expression of the variant polypeptide. Methods of making mutations of murine TLR7 or murine TLR8 are as

described elsewhere herein with reference to making mutations of murine TLR9.

The activity of variants of murine TLR7 and murine TLR8 polypeptides can be tested by cloning the gene encoding the variant murine TLR7 or murine TLR8 polypeptide into a prokaryotic or eukaryotic (e.g., mammalian) expression vector, introducing the vector into an appropriate host cell, expressing the variant murine TLR7 or murine TLR8 polypeptide, and testing for a functional capability of the murine TLR7 or murine TLR8 polypeptides as disclosed herein.

The skilled artisan will also realize that conservative amino acid substitutions may be made in murine TLR7 and murine TLR8 polypeptides to provide functionally equivalent variants of the foregoing polypeptides, i.e., variants which retain the functional capabilities of the murine TLR7 and murine TLR8 polypeptides.

A variety of methodologies well known to the skilled practitioner can be utilized to obtain isolated murine TLR7 and murine TLR8 polypeptide molecules, as previously described in reference to murine TLR9 polypeptides.

The invention as described herein has a number of uses, some of which are described elsewhere herein. For example, the invention permits isolation of the murine TLR7 and the murine TLR8 polypeptide molecules by, e.g., expression of a recombinant nucleic acid to produce large quantities of polypeptide which may be isolated using standard protocols. As another example, the isolation of the murine TLR7 gene makes it possible for murine TLR7 to be used in methods for assaying molecular interactions involving TLR7.

The invention also embraces agents which bind selectively to the murine TLR7 or murine TLR8 nucleic acid molecules or polypeptides as well as agents which bind to variants and fragments of the polypeptides and nucleic acids as described herein. The agents include polypeptides which bind to murine TLR7 or murine TLR8, and antisense nucleic acids, both of which are described in greater detail below. Some agents can inhibit or increase murine TLR7-mediated signaling activity (antagonists and agonists, respectively), and some can inhibit or increase murine TLR8-mediated signaling activity.

In one embodiment the murine TLR7 inhibitor is an antisense oligonucleotide that selectively binds to a murine TLR7 nucleic acid molecule, to reduce the expression of murine TLR7 (or TLR7 of another species) in a cell. This is desirable in virtually any medical condition wherein a reduction of TLR7 signaling activity is desirable. Based upon SEQ ID

NO:173 and SEQ ID NO:174, or upon allelic or homologous genomic and/or cDNA sequences, one of skill in the art can easily choose and synthesize any of a number of appropriate antisense molecules for use in accordance with the present invention.

In one embodiment the murine TLR8 inhibitor is an antisense oligonucleotide that selectively binds to a murine TLR8 nucleic acid molecule, to reduce the expression of murine TLR8 (or TLR8 of another species) in a cell. This is desirable in virtually any medical condition wherein a reduction of TLR8 signaling activity is desirable. Based upon SEQ ID NO:190 and SEQ ID NO:191, or upon allelic or homologous genomic and/or cDNA sequences, one of skill in the art can easily choose and synthesize any of a number of appropriate antisense molecules for use in accordance with the present invention.

Antisense oligonucleotides for murine TLR7 or murine TLR8 can include "natural" and "modified" oligonucleotides as previously described herein.

Agents which bind murine TLR7 or murine TLR8 also include binding peptides and other molecules which bind to the murine TLR7 or murine TLR8 polypeptide and complexes containing the murine TLR7 or murine TLR8 polypeptide, respectively. When the binding molecules are inhibitors, the molecules bind to and inhibit the activity of murine TLR7 or murine TLR8. When the binding molecules are activators, the molecules bind to and increase the activity of murine TLR7 or murine TLR8. To determine whether a murine TLR7 or murine TLR8 binding agent binds to murine TLR7 or murine TLR8, any known binding assay may be employed. For example, the binding agent may be immobilized on a surface and then contacted with a labeled murine TLR7 or murine TLR8 polypeptide. The amount of murine TLR7 or murine TLR8 which interacts with the murine TLR7 or murine TLR8 binding agent, or the amount which does not bind to the murine TLR7 or murine TLR8 binding agent, may then be quantitated to determine whether the murine TLR7 or murine TLR8 binding agent binds to murine TLR7 or murine TLR8.

The murine TLR7 or murine TLR8 binding agents include molecules of numerous size and type that bind selectively or preferentially to murine TLR7 or murine TLR8 polypeptides, and to complexes involving murine TLR7 or murine TLR8 polypeptides and their binding partners. These molecules may be derived from a variety of sources. For example, murine TLR7 or murine TLR8 binding agents can be provided by screening degenerate peptide libraries which can be readily prepared in solution, in immobilized form or

as phage display libraries. Combinatorial libraries also can be synthesized of peptides containing one or more amino acids. Libraries further can be synthesized of peptoids and non-peptide synthetic moieties.

Exemplary methods useful for identifying murine TLR7 and murine TLR8 binding peptides are analogous to those described herein with reference to methods for identifying murine TLR9 binding peptides murine, and thus are not repeated here.

Therefore the invention generally provides efficient methods of identifying pharmacological agents or lead compounds for agents useful in the treatment of conditions associated with TLR7 and TLR8 activity, and the compounds and agents so identified.

Generally, the screening methods involve assaying for compounds which inhibit or enhance the expression of or signaling through murine TLR7 or murine TLR8. Such methods are adaptable to automated, high throughput screening of compounds.

A variety of assays for pharmacological agents are provided, including labeled *in vitro* protein binding assays, signaling assays using detectable molecules, etc. For example, protein binding screens are used to rapidly examine the binding of candidate pharmacological agents to a murine TLR7 or murine TLR8. The candidate pharmacological agents can be derived from, for example, combinatorial peptide or nucleic acid libraries. Convenient reagents for such assays are known in the art. An exemplary cell-based assay of signaling involves contacting a cell having a murine TLR7 or murine TLR8 with a candidate pharmacological agent under conditions whereby the induction of a detectable molecule can occur. A reduced degree of induction of the detectable molecule in the presence of the candidate pharmacological agent indicates that the candidate pharmacological agent reduces the signaling activity of murine TLR7 or murine TLR8. An increased degree of induction of the detectable molecule in the presence of the candidate pharmacological agent indicates that the candidate pharmacological agent increases the signaling activity of murine TLR7 or murine TLR8.

Murine TLR7 and murine TLR8 used in the methods of the invention can be added to an assay mixture as an isolated polypeptide (where binding of a candidate pharmaceutical agent is to be measured) or as a cell or other membrane-encapsulated space which includes a murine TLR7 or murine TLR8 polypeptide. In the latter assay configuration, the cell or other membrane-encapsulated space can contain the murine TLR7 or murine TLR8 as a

polypeptide or as a nucleic acid (e.g., a cell transfected with an expression vector containing a nucleic acid molecule encoding murine TLR7). In the assays described herein, the murine TLR7 or murine TLR8 polypeptide can be produced recombinantly, isolated from biological extracts, or synthesized *in vitro*. Murine TLR7 or murine TLR8 polypeptides encompass
5 chimeric proteins comprising a fusion of a murine TLR7 or murine TLR8 polypeptide with another polypeptide, e.g., a polypeptide capable of providing or enhancing protein-protein binding, enhancing signaling capability, facilitating detection, or enhancing stability of the murine TLR7 or murine TLR8 polypeptide under assay conditions. A polypeptide fused to a murine TLR7 or murine TLR8 polypeptide or fragment thereof may also provide means of
10 readily detecting the fusion protein, e.g., by immunological recognition or by fluorescent labeling.

The assay mixture also comprises a candidate pharmacological agent, as previously described in reference to murine TLR9. Candidate pharmacologic agents are obtained from a wide variety of sources, including libraries of natural, synthetic, or semisynthetic compounds, or any combination thereof. Presently, natural ligands of murine TLR7 and murine TLR8 are
15 unknown, but they appear not to include CpG-ODN.

A variety of other reagents also can be included in the assay mixture. These include reagents such as salts, buffers, neutral proteins (e.g., albumin), detergents, etc. which may be used to facilitate optimal protein-protein and/or protein-nucleic acid binding. Such a reagent
20 may also reduce non-specific or background interactions of the reaction components. Other reagents that improve the efficiency of the assay such as protease inhibitors, nuclease inhibitors, antimicrobial agents, and the like may also be used.

The mixture of the foregoing assay materials is incubated under conditions whereby, but for the presence of the candidate pharmacological agent, the murine TLR7 or murine
25 TLR8 mediates TLR7-mediated or TLR8-mediated signaling, preferably TLR/IL-1R signaling. For determining the binding of a candidate pharmaceutical agent to a murine TLR7 or murine TLR8, the mixture is incubated under conditions which permit binding. The order of addition of components, incubation temperature, time of incubation, and other parameters of the assay may be readily determined. Such experimentation merely involves optimization
30 of the assay parameters, not the fundamental composition of the assay. Incubation temperatures typically are between 4°C and 40°C. Incubation times preferably are minimized

to facilitate rapid, high throughput screening, and typically are between 1 minute and 10 hours.

After incubation, the level of signaling or the level of specific binding between the murine TLR7 or murine TLR8 polypeptide and the candidate pharmaceutical agent is detected by any convenient method available to the user, as described elsewhere herein.

The murine TLR7 or murine TLR8 binding agent may also be an antibody or a functionally active antibody fragment. Antibodies, including monoclonal antibodies and antibody fragments, are well known to those of ordinary skill in the science of immunology and are as described elsewhere herein. Monoclonal antibodies may be made by any of the methods known in the art utilizing murine TLR7 or murine TLR8, or a fragment thereof, as an immunogen. Alternatively the antibody may be a polyclonal antibody specific for murine TLR7 or murine TLR8 which inhibits murine TLR7 or murine TLR8 activity. The preparation and use of polyclonal antibodies are also known to one of ordinary skill in the art.

The sequences of the antigen-binding Fab' portion of the anti-murine TLR7 or anti-murine TLR8 monoclonal antibodies identified as being useful according to the invention in the assays provided above, as well as the relevant FR and CDR regions, can be determined using amino acid sequencing methods that are routine in the art. Such sequence information can be used to generate humanized and chimeric antibodies, as well as various fusion proteins and binding fragments, as described elsewhere herein.

Thus, as will be apparent to one of ordinary skill in the art, the present invention also provides for F(ab')₂ and Fab fragments of an anti-murine TLR7 or anti-murine TLR8 monoclonal antibody; chimeric antibodies in which the Fc and/or FR and/or CDR1 and/or CDR2 and/or light chain CDR3 regions of an anti-murine TLR7 or anti-murine TLR8 antibody have been replaced by homologous human or non-human sequences; chimeric F(ab')₂ fragment antibodies in which the FR and/or CDR1 and/or CDR2 and/or light chain CDR3 regions of an anti-murine TLR7 or anti-murine TLR8 antibody have been replaced by homologous human or non-human sequences; and chimeric Fab fragment antibodies in which the FR and/or CDR1 and/or CDR2 and/or light chain CDR3 regions have been replaced by homologous human or non-human sequences.

According to the invention murine TLR7 and murine TLR8 inhibitors also include "dominant negative" polypeptides derived from SEQ ID NO:175 or SEQ ID NO:192,

respectively. The end result of the expression of a dominant negative murine TLR7 or dominant negative murine TLR8 polypeptide of the invention in a cell is a reduction in TLR7 or murine TLR8 activity such as signaling through the TIR pathway. One of ordinary skill in the art can assess the potential for a dominant negative variant of a murine TLR7 or dominant negative murine TLR8 polypeptide and, using standard mutagenesis techniques, create one or more dominant negative variant polypeptides.

Each of the compositions according to this aspect of the invention is useful for a variety of therapeutic and non-therapeutic purposes. For example, the murine TLR7 and murine TLR8 nucleic acids of the invention are useful as oligonucleotide probes. Such oligonucleotide probes can be used herein to identify genomic or cDNA library clones possessing an identical or substantially similar nucleic acid sequence. Methods of hybridization, synthesis of probes, and detection are generally as described elsewhere herein.

Additionally, complements of the murine TLR7 and murine TLR8 nucleic acids can be useful as antisense oligonucleotides, e.g., by delivering the antisense oligonucleotide to an animal to induce a murine TLR7 or murine TLR8 "knockout" phenotype.

Alternatively, the murine TLR7 and murine TLR8 nucleic acids of the invention can be used to prepare a non-human transgenic animal. The invention, therefore, contemplates the use of murine TLR7 and murine TLR8 knockout and transgenic animals as models for the study of disorders involving TLR7- and murine TLR8-mediated signaling. A variety of methods known to one of ordinary skill in the art are available for the production of transgenic animals associated with this invention.

Inactivation or replacement of the endogenous TLR7 or TLR8 gene can be achieved by a homologous recombination system using embryonic stem cells. The resultant transgenic non-human mammals having a TLR7^{-/-} or TLR8^{-/-} knockout phenotype may be made transgenic for the murine TLR7 or murine TLR8 and used as a model for screening compounds as modulators (agonists or antagonists/inhibitors) of the murine TLR7 or murine TLR8. In this manner, such therapeutic drugs can be identified.

Additionally, a normal or mutant version of murine TLR7 or murine TLR8 can be inserted into the germ line to produce transgenic animals which constitutively or inducibly express the normal or mutant form of murine TLR7 or murine TLR8. These animals are useful in studies to define the role and function of murine TLR7 or murine TLR8 in cells.

The antagonists, agonists, nucleic acids, and polypeptides of murine TLR7 and murine TLR8 useful according to the invention may be combined, optionally, with a pharmaceutically acceptable carrier. Thus the invention also provides pharmaceutical compositions and a method for preparing the pharmaceutical compositions which contain compositions of this aspect of the invention. The pharmaceutical compositions include one or any combination of the antagonists, agonists, nucleic acids and polypeptides of murine TLR7 and murine TLR8 useful according to the invention and, optionally, a pharmaceutically acceptable carrier. Each pharmaceutical composition is prepared by selecting an antagonist, agonist, nucleic acid or polypeptide of murine TLR7 and murine TLR8 useful according to the invention, as well as any combination thereof, and, optionally, combining it with a pharmaceutically acceptable carrier.

A variety of administration routes are available, as described previously herein. The particular mode selected will depend, of course, upon the particular compound selected, the severity of the condition being treated, and the dosage required for therapeutic efficacy.

Likewise, a variety of formulations are contemplated, including, by analogy those discussed above in reference to murine TLR9, unit dose solids, liquids, extended release formulations, etc.

Screening Assays

In another aspect the invention provides methods for screening candidate compounds that act as ISNA mimics, agonists or antagonists in ISNA-induced immunomodulation via TLR7, TLR8, and TLR9. Preferably the screening method can be adapted to accommodate high throughput screening assays, as can be achieved, for example, through the use of multiwell arrays of samples in conjunction with robotic or automated array handling devices.

Immunostimulatory nucleic acids include but are not limited to CpG nucleic acids.

A "CpG nucleic acid" or a "CpG immunostimulatory nucleic acid" as used herein is a nucleic acid containing at least one unmethylated CpG dinucleotide (cytosine-guanine dinucleotide sequence, i.e. "CpG DNA" or DNA containing a 5' cytosine followed by 3' guanine and linked by a phosphate bond) and activates a component of the immune system.

The entire CpG nucleic acid can be unmethylated or portions may be unmethylated but at least the C of the 5' CG 3' must be unmethylated.

In one embodiment a CpG nucleic acid is represented by at least the formula:



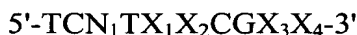
wherein X_1 and X_2 are nucleotides, N is any nucleotide, and N_1 and N_2 are nucleic acid sequences composed of from about 0-25 N's each. In some embodiments X_1 is adenine, guanine, or thymine and/or X_2 is cytosine, adenine, or thymine. In other embodiments X_1 is cytosine and/or X_2 is guanine.

In other embodiments the CpG nucleic acid is represented by at least the formula:



wherein X_1 , X_2 , X_3 , and X_4 are nucleotides; N is any nucleotide; and N_1 and N_2 are nucleic acid sequences composed of from about 0-25 N's each. In some embodiments, X_1X_2 are nucleotides selected from the group consisting of: GpT, GpG, GpA, ApA, ApT, ApG, CpT, CpA, CpG, TpA, TpT, and TpG; and X_3X_4 are nucleotides selected from the group consisting of: TpT, CpT, ApT, TpG, ApG, CpG, TpC, ApC, CpC, TpA, ApA, and CpA. In some embodiments, X_1X_2 are GpA or GpT and X_3X_4 are TpT. In other embodiments X_1 or X_2 or both are purines and X_3 or X_4 or both are pyrimidines or X_1X_2 are GpA and X_3 or X_4 or both are pyrimidines.

In another embodiment the CpG nucleic acid is represented by at least the formula:



wherein X_1 , X_2 , X_3 , and X_4 are nucleotides; N is any nucleotide; and N_1 and N_2 are nucleic acid sequences composed of from about 0-25 N's each. In some embodiments, X_1X_2 are nucleotides selected from the group consisting of: GpT, GpG, GpA, ApA, ApT, ApG, CpT, CpA, CpG, TpA, TpT, and TpG; and X_3X_4 are nucleotides selected from the group consisting of: TpT, CpT, ApT, TpG, ApG, CpG, TpC, ApC, CpC, TpA, ApA, and CpA. In some embodiments, X_1X_2 are GpA or GpT and X_3X_4 are TpT. In other embodiments X_1 or X_2 or both are purines and X_3 or X_4 or both are pyrimidines or X_1X_2 are GpA and X_3 or X_4 or both are pyrimidines.

Examples of CpG nucleic acids according to the invention include but are not limited to those listed in Table 1, such as SEQ ID NOs:21-29, 31-42, 44, 46-50, 52-62, 64-75, 77-88, 90-117, 119-124.

Table 1. Exemplary CpG nucleic acids

	AACGTTCT	SEQ ID NO:21
	AAGCGAAAAATGAAATTGACT	SEQ ID NO:22
	ACCATGGACGAACTGTTTCCCCTC	SEQ ID NO:23
5	ACCATGGACGACCTGTTTCCCCTC	SEQ ID NO:24
	ACCATGGACGAGCTGTTTCCCCTC	SEQ ID NO:25
	ACCATGGACGATCTGTTTCCCCTC	SEQ ID NO:26
	ACCATGGACGGTCTGTTTCCCCTC	SEQ ID NO:27
	ACCATGGACGTA CTGTTTCCCCTC	SEQ ID NO:28
10	ACCATGGACGTTCTGTTTCCCCTC	SEQ ID NO:29
	AGATTTCTAGGAATTCAATC	SEQ ID NO:30
	AGCGGGGGCGAGCGGGGGCG	SEQ ID NO:31
	AGCTATGACGTTCCAAGG	SEQ ID NO:32
	ATCGACTCTCGAGCGTTCTC	SEQ ID NO:33
15	ATGACGTTCTGACGTT	SEQ ID NO:34
	ATGGAAGGTCCAACGTTCTC	SEQ ID NO:35
	ATGGAAGGTCCAGCGTTCTC	SEQ ID NO:36
	ATGGAAGGTCCAGCGTTCTC	SEQ ID NO:37
	ATGGAAGGTCCATCGTTCTC	SEQ ID NO:38
20	CAACGTT	SEQ ID NO:39
	CACGTTGAGGGGGCAT	SEQ ID NO:40
	CAGGCATAACGTTCCGTAG	SEQ ID NO:41
	CCAACGTT	SEQ ID NO:42
	CTCCTAGTGGGGGTGTCCTAT	SEQ ID NO:43
25	CTGATTTCCCGAAATGATG	SEQ ID NO:44
	CTGCTGAGACTGGAG	SEQ ID NO:45
	GAGAACGATGGACCTTCCAT	SEQ ID NO:46
	GAGAACGCTCCAGCACTGAT	SEQ ID NO:47
	GAGAACGCTCGACCTTCCAT	SEQ ID NO:48
30	GAGAACGCTCGACCTTCGAT	SEQ ID NO:49
	GAGAACGCTGGACCTTCCAT	SEQ ID NO:50
	GAGCAAGCTGGACCTTCCAT	SEQ ID NO:51
	GATTGCCTGACGTCAGAGAG	SEQ ID NO:52
	GCATGACGTTGAGCT	SEQ ID NO:53
35	GCGGCGGGCGGCGCGCGCC	SEQ ID NO:54
	GCGTGCGTTGTCGTTGTCGTT	SEQ ID NO:55
	GCTAGACGTTAGCGT	SEQ ID NO:56
	GCTAGACGTTAGTGT	SEQ ID NO:57
	GCTAGATGTTAGCGT	SEQ ID NO:58
40	GCTTGATGACTCAGCCGGAA	SEQ ID NO:59
	GGAATGACGTTCCCTGTG	SEQ ID NO:60
	GGGGTCAACGTTGACGGGG	SEQ ID NO:61
	GGGGTCAGTCTTGACGGGG	SEQ ID NO:62
	GTATTTCCCAGAAAAGGAAC	SEQ ID NO:63
45	GTCCATTTCCCGTAAATCTT	SEQ ID NO:64
	GTCGCT	SEQ ID NO:65
	GTCGTT	SEQ ID NO:66
	TACCGCGTGCGACCCTCT	SEQ ID NO:67
	TATGCATATTCTGTAAAGTG	SEQ ID NO:68
50	TCAACGTC	SEQ ID NO:69
	TCAACGTT	SEQ ID NO:70
	TCAAGCTT	SEQ ID NO:71
	TCAGCGCT	SEQ ID NO:72
	TCAGCGTGCGCC	SEQ ID NO:73
55	TCATCGAT	SEQ ID NO:74
	TCCACGACGTTTTCGACGTT	SEQ ID NO:75
	TCCAGGACTTCTCTCAGGTT	SEQ ID NO:76
	TCCATAACGTTCCCTGATGCT	SEQ ID NO:77
	TCCATAGCGTTCCCTAGCGTT	SEQ ID NO:78
60	TCCATCACGTTGCTGATGCT	SEQ ID NO:79
	TCCATGACGGTCCCTGATGCT	SEQ ID NO:80
	TCCATGACGTTCCCTGATGCT	SEQ ID NO:81

	TCCATGACGTGCCTGATGCT	SEQ ID NO:82
	TCCATGACGTTCCCTGACGTT	SEQ ID NO:83
	TCCATGACGTTCCCTGATGCT	SEQ ID NO:84
	TCCATGAGCTTCCTGATGCT	SEQ ID NO:85
5	TCCATGCCGGTCCTGATGCT	SEQ ID NO:86
	TCCATGCGTGCGTGCGTTTT	SEQ ID NO:87
	TCCATGCGTTGCGTTGCGTT	SEQ ID NO:88
	TCCATGCTGGTCCCTGATGCT	SEQ ID NO:89
	TCCATGGCGGTCCTGATGCT	SEQ ID NO:90
10	TCCATGTGATCCTGATGCT	SEQ ID NO:91
	TCCATGTGCTCCTGATGCT	SEQ ID NO:92
	TCCATGTGCTCCTGATGCT	SEQ ID NO:93
	TCCATGTGCTCCTGCTGAT	SEQ ID NO:94
	TCCATGTGCTCCCTGATGCT	SEQ ID NO:95
15	TCCATGTGCTTCCTGATGCT	SEQ ID NO:96
	TCCATGTGCTTCCTGTCGTT	SEQ ID NO:97
	TCCATGTGCTTTTTGTCGTT	SEQ ID NO:98
	TCCTGACGTTCCCTGACGTT	SEQ ID NO:99
	TCCTGTGCTTCCTGTCGTT	SEQ ID NO:100
20	TCCTGTGCTTCCTGTCGTT	SEQ ID NO:101
	TCCTGTGCTTTTTGTCGTT	SEQ ID NO:102
	TCCTGTGCTTCCTGTCGTT	SEQ ID NO:103
	TGATCGGGCGGGCGAGC	SEQ ID NO:104
	TGCTGCTGTCTCCGCTTCTT	SEQ ID NO:105
25	TGCTGCTGTCTCCGCTTCTTCTTGCC	SEQ ID NO:106
	TGCTGCTGTCTGCCCTTCTT	SEQ ID NO:107
	TGCTGCTGTGTGCTTCTTCTT	SEQ ID NO:108
	TGCTGCTGCTGCTT	SEQ ID NO:109
	TGCTGCTGTGCTTGTGCTT	SEQ ID NO:110
30	TGCTGCTGTGCTTTTTGTCGTT	SEQ ID NO:111
	TGCTGCTTTTTGTCGTTTTGTCGTT	SEQ ID NO:112
	TCTCCAGCGCGCCCAT	SEQ ID NO:113
	TCTCCAGCGGCGCAT	SEQ ID NO:114
	TCTCCAGCGTGCGCCAT	SEQ ID NO:115
35	TCTTCGAA	SEQ ID NO:116
	TGCAGATTGCGCAATCTGCA	SEQ ID NO:117
	TGCTGCTTTTGTGCTTTTGTGCTT	SEQ ID NO:118
	TGTCGCT	SEQ ID NO:119
	TGTCGTT	SEQ ID NO:120
40	TGTCGTTGTGCTT	SEQ ID NO:121
	TGTCGTTGTGCTTGTGCTT	SEQ ID NO:122
	TGTCGTTGTGCTTGTGCTTGTGCTT	SEQ ID NO:123
	TGTCGTTTGTGCTTGTGCTT	SEQ ID NO:124

45 Other ISNAs include but are not limited to T-rich nucleic acids, poly G nucleic acids, and nucleic acids having phosphate modified backbones, such as phosphorothioate backbones.

A "T rich nucleic acid" or "T rich immunostimulatory nucleic acid" is a nucleic acid which includes at least one poly T sequence and/or which has a nucleotide composition of
50 greater than 25% T nucleotide residues and which activates a component of the immune system. A nucleic acid having a poly-T sequence includes at least four Ts in a row, such as 5'TTTT3'. Preferably the T rich nucleic acid includes more than one poly T sequence. In preferred embodiments the T rich nucleic acid may have 2, 3, 4, etc poly T sequences. One of

the most highly immunostimulatory T rich oligonucleotides discovered according to the invention is a nucleic acid composed entirely of T nucleotide residues. Other T rich nucleic acids have a nucleotide composition of greater than 25% T nucleotide residues, but do not necessarily include a poly T sequence. In these T rich nucleic acids the T nucleotide residues may be separated from one another by other types of nucleotide residues, i.e., G, C, and A. In some embodiments the T rich nucleic acids have a nucleotide composition of greater than 30%, 40%, 50%, 60%, 70%, 80%, 90%, and 99%, T nucleotide residues and every integer % in between. Preferably the T rich nucleic acids have at least one poly T sequence and a nucleotide composition of greater than 25% T nucleotide residues.

In one embodiment the T rich nucleic acid is represented by at least the formula:



wherein X_1 , X_2 , X_3 , and X_4 are nucleotides. In one embodiment $X_1 X_2$ is TT and/or $X_3 X_4$ is TT. In another embodiment $X_1 X_2$ are any one of the following nucleotides TA, TG, TC, AT, AA, AG, AC, CT, CC, CA, CG, GT, GG, GA, and GC; and $X_3 X_4$ are any one of the following nucleotides TA, TG, TC, AT, AA, AG, AC, CT, CC, CA, CG, GT, GG, GA, and GC.

In some embodiments it is preferred that the T-rich nucleic acid does not contain poly C (CCCC), poly A (AAAA), poly G (GGGG), CpG motifs, or multiple GGs. In other embodiments the T-rich nucleic acid includes these motifs. Thus in some embodiments of the invention the T rich nucleic acids include CpG dinucleotides and in other embodiments the T rich nucleic acids are free of CpG dinucleotides. The CpG dinucleotides may be methylated or unmethylated.

Poly G containing nucleic acids are also immunostimulatory. A variety of references, including Pisetsky and Reich, 1993 *Mol. Biol. Reports*, 18:217-221; Krieger and Herz, 1994, *Ann. Rev. Biochem.*, 63:601-637; Macaya et al., 1993, *PNAS*, 90:3745-3749; Wyatt et al., 1994, *PNAS*, 91:1356-1360; Rando and Hogan, 1998, In *Applied Antisense Oligonucleotide Technology*, ed. Krieg and Stein, p. 335-352; and Kimura et al., 1994, *J. Biochem.* 116, 991-994 also describe the immunostimulatory properties of poly G nucleic acids.

Poly G nucleic acids preferably are nucleic acids having the following formulas:



wherein X₁, X₂, X₃, and X₄ are nucleotides. In preferred embodiments at least one of X₃ and X₄ are a G. In other embodiments both of X₃ and X₄ are a G. In yet other embodiments the preferred formula is 5' GGGNNGGG 3', or 5' GGGNNGGGNNGGG 3' wherein N represents between 0 and 20 nucleotides. In other embodiments the Poly G nucleic acid is free of unmethylated CG dinucleotides. In other embodiments the poly G nucleic acid includes at least one unmethylated CG dinucleotide.

Nucleic acids having modified backbones, such as phosphorothioate backbones, also fall within the class of immunostimulatory nucleic acids. U.S. Patents Nos. 5,723,335 and 5,663,153 issued to Hutcherson, et al. and related PCT publication WO95/26204 describe immune stimulation using phosphorothioate oligonucleotide analogues. These patents describe the ability of the phosphorothioate backbone to stimulate an immune response in a non-sequence specific manner.

The ISNAs may be double-stranded or single-stranded. Generally, double-stranded molecules may be more stable *in vivo*, while single-stranded molecules may have increased activity. The terms "nucleic acid" and "oligonucleotide" refer to multiple nucleotides (i.e. molecules comprising a sugar (e.g. ribose or deoxyribose) linked to a phosphate group and to an exchangeable organic base, which is either a substituted pyrimidine (e.g. cytosine (C), thymine (T) or uracil (U)) or a substituted purine (e.g., adenine (A) or guanine (G)) or a modified base. As used herein, the terms refer to oligoribonucleotides as well as oligodeoxyribonucleotides. The terms shall also include polynucleosides (i.e., a polynucleotide minus the phosphate) and any other organic base-containing polymer. The terms "nucleic acid" and "oligonucleotide" also encompass nucleic acids or oligonucleotides with a covalently modified base and/or sugar. For example, they include nucleic acids having backbone sugars which are covalently attached to low molecular weight organic groups other than a hydroxyl group at the 3' position and other than a phosphate group at the 5' position. Thus modified nucleic acids may include a 2'-O-alkylated ribose group. In addition, modified nucleic acids may include sugars such as arabinose instead of ribose. Thus the nucleic acids may be heterogeneous in backbone composition thereby containing any possible combination of polymer units linked together such as peptide- nucleic acids (which have amino acid backbone with nucleic acid bases). In some embodiments the nucleic acids are homogeneous in backbone composition.

The substituted purines and pyrimidines of the ISNAs include standard purines and pyrimidines such as cytosine as well as base analogs such as C-5 propyne substituted bases. Wagner RW et al., *Nat Biotechnol* 14:840-844 (1996). Purines and pyrimidines include but are not limited to adenine, cytosine, guanine, thymine, 5-methylcytosine, 2-aminopurine, 2-amino-6-chloropurine, 2,6-diaminopurine, hypoxanthine, and other naturally and non-naturally occurring nucleobases, substituted and unsubstituted aromatic moieties.

The ISNA is a linked polymer of bases or nucleotides. As used herein with respect to linked units of a nucleic acid, "linked" or "linkage" means two entities are bound to one another by any physicochemical means. Any linkage known to those of ordinary skill in the art, covalent or non-covalent, is embraced. Such linkages are well known to those of ordinary skill in the art. Natural linkages, which are those ordinarily found in nature connecting the individual units of a nucleic acid, are most common. The individual units of a nucleic acid may be linked, however, by synthetic or modified linkages.

Whenever a nucleic acid is represented by a sequence of letters it will be understood that the nucleotides are in 5' to 3' order from left to right and that "A" denotes adenine, "C" denotes cytosine, "G" denotes guanine, "T" denotes thymidine, and "U" denotes uracil unless otherwise noted.

Immunostimulatory nucleic acid molecules useful according to the invention can be obtained from natural nucleic acid sources (e.g., genomic nuclear or mitochondrial DNA or cDNA), or are synthetic (e.g., produced by oligonucleotide synthesis). Nucleic acids isolated from existing nucleic acid sources are referred to herein as native, natural, or isolated nucleic acids. The nucleic acids useful according to the invention may be isolated from any source, including eukaryotic sources, prokaryotic sources, nuclear DNA, mitochondrial DNA, etc. Thus, the term nucleic acid encompasses both synthetic and isolated nucleic acids.

The term "isolated" as used herein with reference to an ISNA means substantially free of or separated from components which it is normally associated with in nature, e.g., nucleic acids, proteins, lipids, carbohydrates or *in vivo* systems to an extent practical and appropriate for its intended use. In particular, the nucleic acids are sufficiently pure and are sufficiently free from other biological constituents of host cells so as to be useful in, for example, producing pharmaceutical preparations. Because an isolated nucleic acid of the invention may be admixed with a pharmaceutically-acceptable carrier in a pharmaceutical preparation,

the nucleic acid may comprise only a small percentage by weight of the preparation. The nucleic acid is nonetheless substantially pure in that it has been substantially separated from the substances with which it may be associated in living systems.

The ISNAs can be produced on a large scale in plasmids, (see *Molecular Cloning: A Laboratory Manual*, J. Sambrook, et al., eds., Second Edition, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York, 1989) and separated into smaller pieces or administered whole. After being administered to a subject the plasmid can be degraded into oligonucleotides. One skilled in the art can purify viral, bacterial, eukaryotic, etc. nucleic acids using standard techniques, such as those employing restriction enzymes, exonucleases or endonucleases.

For use in the instant invention, the ISNAs can be synthesized *de novo* using any of a number of procedures well known in the art. For example, the β -cyanoethyl phosphoramidite method (Beaucage SL and Caruthers MH, *Tetrahedron Let* 22:1859 (1981)); nucleoside H-phosphonate method (Garegg et al., *Tetrahedron Let* 27:4051-4054 (1986); Froehler et al., *Nucl Acid Res* 14:5399-5407 (1986); Garegg et al., *Tetrahedron Let* 27:4055-4058 (1986); Gaffney et al., *Tetrahedron Let* 29:2619-2622 (1988)). These chemistries can be performed by a variety of automated oligonucleotide synthesizers available in the market.

ISNAs having modified backbones, such as phosphorothioate backbones, also fall within the class of immunostimulatory nucleic acids. U.S. Patents Nos. 5,723,335 and 5,663,153 issued to Hutcherson, et al. and related PCT publication WO95/26204 describe immune stimulation using phosphorothioate oligonucleotide analogues. These patents describe the ability of the phosphorothioate backbone to stimulate an immune response in a non-sequence specific manner.

The ISNA may be any size of at least 6 nucleotides but in some embodiments are in the range of between 6 and 100 or in some embodiments between 8 and 35 nucleotides in size. Immunostimulatory nucleic acids can be produced on a large scale in plasmids. These may be administered in plasmid form or alternatively they can be degraded into oligonucleotides before administration.

"Palindromic sequence" shall mean an inverted repeat (i.e., a sequence such as ABCDEE'D'C'B'A' in which A and A', B and B', etc., are bases capable of forming the usual Watson-Crick base pairs and which includes at least 6 nucleotides in the palindrome. *In vivo*,

such sequences may form double-stranded structures. In one embodiment the nucleic acid contains a palindromic sequence. In some embodiments when the nucleic acid is a CpG nucleic acid, a palindromic sequence used in this context refers to a palindrome in which the CpG is part of the palindrome, and optionally is the center of the palindrome. In another embodiment the nucleic acid is free of a palindrome. A nucleic acid that is free of a
5 palindrome does not have any regions of 6 nucleotides or greater in length which are palindromic. A nucleic acid that is free of a palindrome can include a region of less than 6 nucleotides which are palindromic.

A "stabilized ISNA" shall mean a nucleic acid molecule that is relatively resistant to
10 *in vivo* degradation (e.g. via an exo- or endo-nuclease). Stabilization can be a function of length or secondary structure. Nucleic acids that are tens to hundreds of kbs long are relatively resistant to *in vivo* degradation. For shorter nucleic acids, secondary structure can stabilize and increase their effect. For example, if the 3' end of an oligonucleotide has self-complementarity to an upstream region, so that it can fold back and form a sort of stem loop
15 structure, then the oligonucleotide becomes stabilized and therefore exhibits more activity.

Some stabilized ISNAs of the instant invention have a modified backbone. It has been demonstrated that modification of the oligonucleotide backbone provides enhanced activity of the ISNAs when administered *in vivo*. Nucleic acids, including at least two phosphorothioate linkages at the 5' end of the oligonucleotide and multiple phosphorothioate linkages at the 3'
20 end, preferably 5, may provide maximal activity and protect the oligonucleotide from degradation by intracellular exo- and endo-nucleases. Other modified oligonucleotides include phosphodiester modified oligonucleotide, combinations of phosphodiester and phosphorothioate oligonucleotide, methylphosphonate, methylphosphorothioate, phosphorodithioate, and combinations thereof. Each of these combinations and their
25 particular effects on immune cells is discussed in more detail in U.S. Patent Nos. 6,194,388 and 6,207,646, the entire contents of which is hereby incorporated by reference. It is believed that these modified oligonucleotides may show more stimulatory activity due to enhanced nuclease resistance, increased cellular uptake, increased protein binding, and/or altered intracellular localization. Both phosphorothioate and phosphodiester nucleic acids are active
30 in immune cells.

Other stabilized ISNAs include: nonionic DNA analogs, such as alkyl- and aryl-

phosphates (in which the charged phosphonate oxygen is replaced by an alkyl or aryl group), phosphodiester and alkylphosphotriesters, in which the charged oxygen moiety is alkylated. Oligonucleotides which contain diol, such as tetraethyleneglycol or hexaethyleneglycol, at either or both termini have also been shown to be substantially resistant to nuclease degradation.

For use *in vivo*, ISNAs are preferably relatively resistant to degradation (e.g., via endo- and exo-nucleases). Secondary structures, such as stem loops, can stabilize nucleic acids against degradation. Alternatively, nucleic acid stabilization can be accomplished via phosphate backbone modifications. One type of stabilized nucleic acid has at least a partial phosphorothioate modified backbone. Phosphorothioates may be synthesized using automated techniques employing either phosphoramidate or H-phosphonate chemistries. Aryl- and alkyl-phosphonates can be made, e.g., as described in U.S. Patent No. 4,469,863; and alkylphosphotriesters (in which the charged oxygen moiety is alkylated as described in U.S. Patent No. 5,023,243 and European Patent No. 092,574) can be prepared by automated solid phase synthesis using commercially available reagents. Methods for making other DNA backbone modifications and substitutions have been described. Uhlmann E and Peyman A, *Chem Rev* 90:544 (1990); Goodchild J, *Bioconjugate Chem* 1:165 (1990).

Other sources of immunostimulatory nucleic acids useful according to the invention include standard viral and bacterial vectors, many of which are commercially available. In its broadest sense, a "vector" is any nucleic acid material which is ordinarily used to deliver and facilitate the transfer of nucleic acids to cells. The vector as used herein may be an empty vector or a vector carrying a gene which can be expressed. In the case when the vector is carrying a gene the vector generally transports the gene to the target cells with reduced degradation relative to the extent of degradation that would result in the absence of the vector.

In this case the vector optionally includes gene expression sequences to enhance expression of the gene in target cells such as immune cells, but it is not required that the gene be expressed in the cell.

A basis for certain of the screening assays is the presence of a functional TLR 7, TLR 8, or TLR9 in a cell. The functional TLR in some instances is naturally expressed by the cell. In other instances, expression of the functional TLR can involve introduction or reconstitution of a species-specific TLR9 into a cell or cell line that otherwise lacks the TLR

or lacks responsiveness to ISNA, resulting in a cell or cell line capable of activating the TLR/IL-1R signaling pathway in response to contact with an ISNA. Examples of cell lines lacking TLR9 or ISNA responsiveness include, but are not limited to, 293 fibroblasts (ATCC CRL-1573), MonoMac-6, THP-1, U937, CHO, and any TLR9 knock-out. The introduction of the species-specific TLR into the cell or cell line is preferably accomplished by transient or stable transfection of the cell or cell line with a TLR-encoding nucleic acid sequence operatively linked to a gene expression sequence (as described above).

The species-specific TLR, including TLR7, TLR8, and TLR9, is not limited to a murine TLR, but rather can include a TLR derived from murine or non-murine sources.

Examples of non-murine sources include, but are not limited to, human, bovine, canine, feline, ovine, porcine, and equine. Other species include chicken and fish, e.g., aquaculture species.

The species-specific TLR, including TLR7, TLR8, and TLR9, also is not limited to native TLR polypeptides. In certain embodiments the TLR can be, e.g., a chimeric TLR in which the extracellular domain and the cytoplasmic domains are derived from TLR polypeptides from different species. Such chimeric TLR polypeptides, as described above, can include, for example, a human TLR extracellular domain and a murine TLR cytoplasmic domain, each domain derived from the corresponding TLR7, TLR8, or TLR9 of each species.

In alternative embodiments, such chimeric TLR polypeptides can include chimeras created with different TLR splice variants or allotypes. Other chimeric TLR polypeptides useful for the purposes of screening ISNA mimics, agonists and antagonists can include chimeric polypeptides created with a TLR of a first type, e.g., TLR9, and another TLR, e.g., TLR7 or TLR8, of the same or another species as the TLR of the first type. Also contemplated are chimeric polypeptides which incorporate sequences derived from more than two polypeptides, e.g., an extracellular domain, a transmembrane domain, and a cytoplasmic domain all derived from different polypeptide sources, provided at least one such domain derives from a TLR7, TLR8, or TLR9 polypeptide. As a further example, also contemplated are constructs such as include an extracellular domain of one TLR9, an intracellular domain of another TLR9, and a non-TLR reporter such as luciferase, GFP, etc. Those of skill in the art will recognize how to design and generate DNA sequences coding for such chimeric TLR polypeptides.

The screening assays can have any of a number of possible readout systems based

upon either TLR/IL-1R signaling pathway or other assays useful for assaying response to ISNAs. It has been reported that immune cell activation by CpG immunostimulatory sequences is dependent in some way on endosomal processing. It is not yet known whether TLR9 is directly involved in this endosomal pathway, or if there is some intermediary between TLR9 and the endosome.

In preferred embodiments, the readout for the screening assay is based on the use of native genes or, alternatively, cotransfected or otherwise co-introduced reporter gene constructs which are responsive to the TLR/IL-1R signal transduction pathway involving MyD88, TRAF6, p38, and/or ERK. Häcker H et al., *EMBO J* 18:6973-6982 (1999). These pathways activate kinases including κ B kinase complex and c-Jun N-terminal kinases. Thus reporter genes and reporter gene constructs particularly useful for the assays can include a reporter gene operatively linked to a promoter sensitive to NF- κ B. Examples of such promoters include, without limitation, those for NF- κ B, IL-1 β , IL-6, IL-8, IL-12 p40, CD80, CD86, and TNF- α . The reporter gene operatively linked to the TLR7-, TLR8-, or TLR9-sensitive promoter can include, without limitation, an enzyme (e.g., luciferase, alkaline phosphatase, β -galactosidase, chloramphenicol acetyltransferase (CAT), etc.), a bioluminescence marker (e.g., green-fluorescent protein (GFP, U.S. patent 5,491,084), etc.), a surface-expressed molecule (e.g., CD25), and a secreted molecule (e.g., IL-8, IL-12 p40, TNF- α). In preferred embodiments the reporter is selected from IL-8, TNF- α , NF- κ B-luciferase (NF- κ B-luc; Häcker H et al., *EMBO J* 18:6973-6982 (1999)), IL-12 p40-luc (Murphy TL et al., *Mol Cell Biol* 15:5258-5267 (1995)), and TNF-luc (Häcker H et al., *EMBO J* 18:6973-6982 (1999)). In assays relying on enzyme activity readout, substrate can be supplied as part of the assay, and detection can involve measurement of chemiluminescence, fluorescence, color development, incorporation of radioactive label, drug resistance, or other marker of enzyme activity. For assays relying on surface expression of a molecule, detection can be accomplished using FACS analysis or functional assays. Secreted molecules can be assayed using enzyme-linked immunosorbent assay (ELISA) or bioassays. Many such readout systems are well known in the art and are commercially available.

In another aspect the invention provides a screening method for identifying an immunostimulatory nucleic acid molecule (ISNA). The method entails contacting a functional TLR selected from the group consisting of TLR7, TLR8, and TLR9 with a test

nucleic acid molecule; detecting the presence or absence of a response mediated by a TLR signal transduction pathway in the presence of the test nucleic acid molecule arising as a result of an interaction between the functional TLR and the test nucleic acid molecule; and determining the test nucleic acid molecule is an ISNA when the presence of a response mediated by the TLR signal transduction pathway is detected. "Functional TLR" and a "cell expressing functional TLR" are as described elsewhere herein. A response mediated by a TLR signal transduction pathway includes induction of a gene under control of a promoter responsive to the TLR/IL-1R signaling pathway, including but not limited to promoters responsive to NF- κ B. The biological response thus can include, e.g., secretion of IL-8 and luciferase activity in a cell transfected with NF- κ B-luc, IL-12 p40-luc, or TNF-luc. A test nucleic acid molecule can include a DNA, RNA, or modified nucleic acid molecule as described herein. In some embodiments the test nucleic acid molecule is a CpG nucleic acid.

Preferably, the test nucleic acid molecule is a sequence variant of a reference ISNA, containing at least one alternative base, at least one alternative internucleotide backbone linkage, or at least one alternative sugar moiety as compared to the particular reference ISNA. In a preferred embodiment the test nucleic acid molecule is a member of a library of such test nucleic acid molecules.

According to one embodiment of this method, comparison can be made to a reference ISNA. The reference ISNA may be any ISNA, including a CpG nucleic acid. In preferred embodiments the screening method is performed using a plurality of test nucleic acids. Preferably comparison of test and reference responses is based on comparison of quantitative measurements of responses in each instance.

The method can be used to select a subset of test nucleic acid molecules based on their ability to induce a similar specific response mediated by the TLR signal transduction pathway. For instance, the method can be used to classify test CpG nucleic acids as predominantly B-cell activating CpG nucleic acids, or as predominantly IFN- α inducing CpG nucleic acids. Other new classes of ISNAs may be identified and characterized using the method.

Application of this method permits the identification of ISNAs, delineation of sequence specificity of a given TLR, and also optimization of ISNA sequences. Identification of ISNAs involves screening candidate ISNAs as above and selecting any ISNA that induces

a response as defined. Delineation of sequence specificity involves screening candidate ISNAs as above with reference to a particular TLR9, selecting any ISNAs that induce a response as defined, and categorizing ISNAs that do and do not induce a response on the basis of their sequence. Optimization of ISNA sequences involves an iterative application of the method as described, further including the steps of selecting the best sequence at any given stage or round in the screening and substituting it as a benchmark or reference in a subsequent round of screening. This latter process can further include selection of parameters to modify in choosing and generating candidate ISNAs to screen.

In another aspect the invention provides screening method for identifying species specificity of an ISNA. The method involves contacting a functional TLR selected from the group consisting of TLR7, TLR8, and TLR9 of a first species with a test ISNA; contacting a functional TLR selected from the group consisting of TLR7, TLR8, and TLR9 of a second species with the test ISNA; measuring a response mediated by a TLR signal transduction pathway associated with the contacting the functional TLR of the first species with the test ISNA; measuring a response mediated by the TLR signal transduction pathway associated with the contacting the functional TLR of the second species with the test ISNA; and comparing (a) the response mediated by a TLR signal transduction pathway associated with the contacting the functional TLR of the first species with the test ISNA with (b) the response mediated by the TLR signal transduction pathway associated with the contacting the functional TLR of the second species with the test ISNA. The functional TLR may be expressed by a cell or it may be part of a cell-free system. The functional TLR may be part of a complex, with either another TLR or with another protein, e.g., MyD88, IRAK, TRAF6, I κ B, NF- κ B, or functional homologues and derivatives thereof. Thus for example a given ODN can be tested against a panel of 293 fibroblast cells transfected with TLR7, TLR8, or TLR9 from various species and optionally cotransfected with a reporter construct (e.g., NF- κ B-luc) sensitive to TLR/IL-1R activation pathways. Thus in another aspect, the invention provides a method for screening species selectivity with respect to a given nucleic acid sequence.

As mentioned above, the invention in one aspect provides a screening method for comparing TLR signaling activity or a test compound against corresponding TLR signaling activity of a reference ISNA. The methods generally involve contacting a functional TLR

selected from the group consisting of TLR7, TLR8, and TLR9 with a reference ISNA and detecting a reference response mediated by a TLR signal transduction pathway; contacting a functional TLR selected from the group consisting of TLR7, TLR8, and TLR9 with a test compound and detecting a test response mediated by a TLR signal transduction pathway; and
5 comparing the test response with the reference response to compare the TLR signaling activity of the test compound with the ISNA. Assays in which the test compound and the reference ISNA contact the TLR independently may be used to identify test compounds that are ISNA mimics. Assays in which the test compound and the reference ISNA contact the TLR concurrently may be used to identify test compounds that are ISNA agonists and ISNA
10 antagonists.

An ISNA mimic as used herein is a compound which causes a response mediated by a TLR signal transduction pathway. As used herein the term "response mediated by a TLR signal transduction pathway" refers to a response which is characteristic of an ISNA-TLR interaction. As demonstrated herein responses which are characteristic of ISNA-TLR
5 interactions include the induction of a gene under control of an ISNA-specific promoter such as a NF- κ B promoter, increases in Th1 cytokine levels, etc. The gene under the control of the NF- κ B promoter may be a gene which naturally includes an NF- κ B promoter or it may be a gene in a construct in which an NF- κ B promoter has been inserted. Genes which naturally include the NF- κ B promoter include but are not limited to IL-8, IL-12 p40, NF- κ B-luc, IL-12
20 p40-luc, and TNF-luc. Increases in Th1 cytokine levels is another measure characteristic of an ISNA-TLR interaction. Increases in Th1 cytokine levels may result from increased production or increased stability or increased secretion of the Th1 cytokines in response to the ISNA-TLR interaction. Th1 cytokines include but are not limited to IL-2, IFN- γ , and IL-12. Other responses which are characteristic of an ISNA-TLR interaction include but are not
25 limited to a reduction in Th2 cytokine levels. Th2 cytokines include but are not limited to IL-4, IL-5, and IL-10.

The response which is characteristic of an ISNA-TLR interaction may be a direct response or an indirect response. A direct response is a response that arises directly as a result of the ISNA-TLR interaction. An indirect response is a response which involves the
30 modulation of other parameters prior to its occurrence.

An ISNA agonist as used herein is a compound which causes an enhanced response to

an ISNA mediated by a TLR signal transduction pathway. Thus an ISNA agonist as used herein is a compound which causes an increase in at least one aspect of an immune response that is ordinarily induced by the reference ISNA. For example, an immune response that is ordinarily induced by an ISNA can specifically include TLR7-, TLR8-, or TLR9-mediated signal transduction in response to immunostimulatory CpG nucleic acid. An ISNA agonist will in some embodiments compete with ISNA for binding to TLR7, TLR8, or TLR9. In other embodiments an ISNA agonist will bind to a site on TLR7, TLR8, or TLR9 that is distinct from the site for binding ISNA. In yet other embodiments an ISNA agonist will act via another molecule or pathway distinct from TLR7, TLR8, or TLR9.

An ISNA antagonist as used herein is a compound which causes a decreased response to an ISNA mediated by a TLR signal transduction pathway. Thus an ISNA antagonist as used herein is a compound which causes a decrease in at least one aspect of an immune response that is ordinarily induced by the reference ISNA. For example, an immune response that is ordinarily induced by an ISNA can specifically include TLR7-, TLR8-, or TLR9-mediated signal transduction in response to immunostimulatory CpG nucleic acid. An ISNA antagonist will in some embodiments compete with ISNA for binding to TLR7, TLR8, or TLR9. In other embodiments an ISNA antagonist will bind to a site on TLR7, TLR8, or TLR9 that is distinct from the site for binding ISNA. In yet other embodiments an ISNA antagonist will act via another molecule or pathway distinct from TLR7, TLR8, or TLR9.

The screening methods for comparing TLR signaling activity of a test compound with signaling activity of an ISNA involve contacting at least one test compound with a functional TLR selected from TLR7, TLR8, and TLR9 under conditions which, in the absence of a test compound, permit a reference ISNA to induce at least one aspect of an immune response. The functional TLR may be expressed by a cell or it may be part of a cell-free system. A cell expressing a functional TLR is a cell that either naturally expresses the TLR, or is a cell into which has been introduced a TLR expression vector, or is a cell manipulated to express TLR in a manner that allows the TLR to be expressed by the cell and to transduce a signal under conditions which normally permit signal transduction by the signal transducing portion of the TLR. The TLR can be a native TLR or it can be a fragment or variant thereof, as described above. According to these methods, the test compound is contacted with a functional TLR or TLR-expressing cell before, after, or simultaneously with contacting a reference ISNA with

the functional TLR or TLR-expressing cell. A response of the functional TLR or TLR-expressing cell is measured and compared with the corresponding response that results or would result under the same conditions in the absence of the test compound. Where it is appropriate, the response in the absence of the test compound can be determined as a concurrent or historical control. Examples of such responses include, without limitation, a response mediated through the TLR signal transduction pathway, secretion of a cytokine, cell proliferation, and cell activation. In a preferred embodiment, the measurement of a response involves the detection of IL-8 secretion (e.g., by ELISA). In another preferred embodiment, the measurement of the response involves the detection of luciferase activity (e.g., NF- κ B-luc, IL-12 p40-luc, or TNF-luc).

Examples of reference ISNAs include, without limitation, those listed in Table 1 (above). In some preferred embodiments the reference ISNA is a CpG nucleic acid.

Test compounds can include but are not limited to peptide nucleic acids (PNAs), antibodies, polypeptides, carbohydrates, lipids, hormones, and small molecules. Test compounds can further include variants of a reference ISNA incorporating any one or combination of the substitutions described above. Test compounds can be generated as members of a combinatorial library of compounds.

In preferred embodiments, the methods for screening test compounds, test nucleic acid molecules, test ISNAs, and candidate pharmacological agents can be performed on a large scale and with high throughput by incorporating, e.g., an array-based assay system and at least one automated or semi-automated step. For example, the assays can be set up using multiwell plates in which cells are dispensed in individual wells and reagents are added in a systematic manner using a multiwell delivery device suited to the geometry of the multiwell plate. Manual and robotic multiwell delivery devices suitable for use in a high throughput screening assay are well known by those skilled in the art. Each well or array element can be mapped in a one-to-one manner to a particular test condition, such as the test compound. Readouts can also be performed in this multiwell array, preferably using a multiwell plate reader device or the like. Examples of such devices are well known in the art and are available through commercial sources. Sample and reagent handling can be automated to further enhance the throughput capacity of the screening assay, such that dozens, hundreds, thousands, or even millions of parallel assays can be performed in a day or in a week. Fully

robotic systems are known in the art for applications such as generation and analysis of combinatorial libraries of synthetic compounds. See, for example, U.S. patents 5,443,791 and 5,708,158.

5 The invention will be more fully understood by reference to the following examples. These examples, however, are merely intended to illustrate the embodiments of the invention and are not to be construed to limit the scope of the invention.

Examples

10 **Example 1. Method of cloning the mouse TLR9**

Alignment of human TLR9 protein sequence with mouse EST database using tfasta yielded 7 hits with mouse EST sequences aa197442, ai451215, aa162495, aw048117, ai463056, aw048548, and aa273731. Two primers were designed that bind to aa197442 EST sequence for use in a RACE-PCR to amplify 5' and 3' ends of the mouse TLR9 cDNA. The library used for the RACE PCR was a mouse spleen marathon-ready cDNA commercially available from Clontech. A 5' fragment with a length of 1800 bp obtained by this method was cloned into Promega pGEM-T Easy vector. After sequencing of the 5' end, additional primers were designed for amplification of the complete mouse TLR9 cDNA. The primer for the 5' end was obtained from the sequence of the 5' RACE product whereas the primer for the 3' end was selected from the mouse EST sequence aa273731.

Three independent PCR reactions were set up using a murine macrophage RAW264.7 (ATCC TIB-71) cDNA as a template, and the resulting amplification products were cloned into the pGEM-T Easy vector. The inserts were fully sequenced, translated into protein and aligned to the human protein sequence. One out of three clones was error-free based on alignment comparison (clone mtlr932e.pep). The cDNA sequence for mTLR9 is SEQ ID NO:1, is presented in Table 2. The ATG start codon occurs at base 40, and a TAG termination codon occurs at base 3136. SEQ ID NO:2 (Table 3), corresponding to bases 40-3135 of SEQ ID NO:1, is the coding region for the polypeptide of SEQ ID NO:3.

30 **Table 2. cDNA Sequence for Murine TLR9 (5' to 3'; SEQ ID NO:1)**

tgtcagaggg	agcctcgga	gaatcctcca	tctcccaaca	tggttctccg	tccaaggact	60
ctgcacccct	tgccctcct	ggtacaggct	gcagtgtctg	ctgagactct	ggccctgggt	120

	accctgcctg	ccttcctacc	ctgtgagctg	aagcctcatg	gcctgggtgga	ctgcaattgg	180
	ctgttcctga	agtctgtacc	ccgtttctct	gcggcagcat	cctgctccaa	catcacccgc	240
	ctctccttga	tctccaaccg	tatccaccac	ctgcacaact	ccgacttcgt	ccacctgtcc	300
	aacctgcggc	agctgaacct	caagtggaa	tgtccaccca	ctggccttag	ccccctgcac	360
5	ttctcttgcc	acatgacct	tgagcccaga	accttcctgg	ctatgcgtac	actggaggag	420
	ctgaacctga	gctataatgg	tatcaccact	gtgccccgac	tgcccagctc	cctgggtgaat	480
	ctgagcctga	gccacaccaa	catcctgggt	ctagatgcta	acagcctcgc	cggcctatac	540
	agcctgcgcg	ttctcttcat	ggacgggaac	tgctactaca	agaacccttg	cacaggagcg	600
	gtgaagggtga	ccccaggcgc	cctcctgggg	ctgagcaatc	tcacccatct	gtctctgaag	660
10	tataacaacc	tcacaaaggt	gccccgcaa	ctgcccccca	gcctggagta	cctcctgggtg	720
	tcctataaacc	tcattgtcaa	gctggggcct	gaagacctgg	ccaatctgac	ctcccttcga	780
	gtacttgatg	tgggtgggaa	ttgccgtcgc	tgcgaccatg	cccccaatcc	ctgtatagaa	840
	tgtggccaaa	agtccctcca	cctgcaccct	gagaccttcc	atcacctgag	ccatctggaa	900
	ggcctgggtgc	tgaaggacag	ctctctccat	acactgaact	cttcctgggt	ccaagggtctg	960
15	gtcaacctct	cgggtgctgga	cctaagcgag	aactttctct	atgaaagcat	caaccacacc	1020
	aatgcctttc	agaacctaac	ccgcctgcgc	aagctcaacc	tgctcttcaa	ttaccgcaag	1080
	aaggatcct	ttgcccgcc	ccacctggca	agttccttca	agaacctgg	gtcactgcag	1140
	gagctgaaca	tgaacggcat	cttcttccgc	tcgctcaaca	agtacacgct	cagatggctg	1200
	gccgatctgc	ccaaactcca	cactctgcac	cttcaaata	acttcatcaa	ccaggcacag	1260
20	ctcagcatct	ttggtacctt	ccgagccctt	cgttttgtgg	acttgtcaga	caatcgcatc	1320
	agtgggcctt	caacgctgtc	agaagccacc	cctgaagagg	cagatgatgc	agagcaggag	1380
	gagctgttgt	ctgcggatcc	tcacccagct	ccactgagca	cccctgcttc	taagaacttc	1440
	atggacaggt	gtaagaactt	caagttcacc	atggacctgt	ctcggaacaa	cctgggtgact	1500
	atcaagccag	agatgtttgt	caatctctca	cgcctccagt	gtcttagcct	gagccacaa	1560
25	tccattgcac	aggctgtcaa	tggctctcag	ttcctgccgc	tgactaatct	gcagggtgctg	1620
	gacctgtccc	ataacaaact	ggacttgtac	cactggaaat	cgttcagtga	gctaccacag	1680
	ttgcaggccc	tggacctgag	ctacaacagc	cagcccttta	gcatgaagg	tataggccac	1740
	aatttcagtt	ttgtggccca	tctgtccatg	ctacacagcc	ttagcctggc	acacaatgac	1800
	attcataccc	gtgtgtcctc	acatctcaac	agcaactcag	tgaggtttct	tgacttcagc	1860
30	ggcaacggta	tgggccgcat	gtgggatgag	gggggccttt	atctccattt	cttccaaggc	1920
	ctgagtggcc	tgctgaagct	ggacctgtct	caaaataacc	tgcatatcct	ccggccccag	1980
	aaccttgaca	acctcccaa	gagcctgaag	ctgctgagcc	tccgagacaa	ctacctatct	2040
	ttctttaact	ggaccagtct	gtccttcctg	cccaacctgg	aagtcctaga	cctggcaggc	2100
	aaccagctaa	aggccctgac	caatggcacc	ctgcctaata	gcaccctcct	ccagaaactg	2160
35	gatgtcagca	gcaacagtat	cgtctctgtg	gtcccagcct	tcttcgctct	ggcggtcgag	2220
	ctgaaagagg	tcaacctcag	ccacaacatt	ctcaagacgg	tggatcgctc	ctgggttgagg	2280
	cccattgtga	tgaacctgac	agttctagac	gtgagaagca	accctctgca	ctgtgcctgt	2340
	ggggcagcct	tcgtagactt	actgttgagg	gtgcagacca	aggtgcctgg	cctgggcta	2400
	ggtgtgaagt	gtggcagccc	cggccagctg	cagggccgta	gcatcttcgc	acaggacctg	2460
40	cggctgtgcc	tggatgaggt	cctctcttgg	gactgctttg	gcctttcact	cttggctgtg	2520
	gccgtgggca	tgggtgggtg	tatactgcac	catctctgcg	gctgggacgt	ctgggtactgt	2580
	tttcatctgt	gcctggcatg	gctacctttg	ctggccccga	gccgacgcag	cgcaccaagt	2640

	ctccccctatg	atgccttcgt	ggtgttcgat	aaggcacaga	gcgcagttgc	ggactgggtg	2700
	tataacgagc	tgcgggtgcg	gctggaggag	cggcgcggtc	gccgagccct	acgcttgtgt	2760
	ctggaggacc	gagattggct	gcctggccag	acgctcttcg	agaacctctg	ggcttccatc	2820
	tatgggagcc	gcaagactct	atttgtgctg	gccacacagg	accgcgtcag	tggcctcctg	2880
5	cgcaccagct	tcctgctggc	tcagcagcgc	ctgttggaag	accgcaagga	cgtggtggtg	2940
	ttggtgatcc	tgcgtccgga	tccccaccgc	tcccgctatg	tgcgactgcg	ccagcgtctc	3000
	tgccgccaga	gtgtgctctt	ctggccccag	cagcccaacg	ggcagggggg	cttctgggcc	3060
	cagctgagta	cagccctgac	tagggacaac	cgccacttct	ataaccagaa	cttctgccgg	3120
	ggacctacag	cagaatagct	cagagcaaca	gctggaaaca	gctgcatctt	catgcctggt	3180
10	tcccagagtg	ctctgcctgc					3200

Table 3. Coding region for murine TLR9 (SEQ ID NO:2)

	atggttctcc	gtcgaaggac	tctgcacccc	ttgtccctcc	tggtacaggc	tgcagtgctg	60
	gctgagactc	tggccctggg	taccctgcct	gccttcctac	cctgtgagct	gaagcctcat	120
5	ggcctgggtg	actgcaattg	gctgttcctg	aagtctgtac	cccgtttctc	tgcggcagca	180
	tcctgctcca	acatcacccg	cctctccttg	atctccaacc	gtatccacca	cctgcacaac	240
	tccgacttcg	tccacctgtc	caacctgcgg	cagctgaacc	tcaagtggaa	ctgtccaccc	300
	actggcctta	gccccctgca	cttctcttgc	cacatgacca	ttgagcccag	aaccttcctg	360
	gctatgcgta	cactggagga	gctgaacctg	agctataatg	gtatcaccac	tgtgccccga	420
20	ctgcccagct	ccctggtgaa	tctgagcctg	agccacacca	acatcctggg	tctagatgct	480
	aacagcctcg	ccggcctata	cagcctgcgc	gttctcttca	tggacgggaa	ctgctactac	540
	aagaacccct	gcacaggagc	ggtgaagggtg	acccagggcg	ccctcctggg	cctgagcaat	600
	ctcacccatc	tgtctctgaa	gtataacaac	ctcacaaagg	tgccccgcca	actgcccccc	660
	agcctggagt	acctcctggg	gtcctataac	ctcattgtca	agctgggggc	tgaagacctg	720
25	gccaatctga	cctcccttcg	agtacttgat	gtgggtggga	attgccgtcg	ctgcgaccat	780
	gcccccaatc	cctgtataga	atgtggccaa	aagtccctcc	acctgcaccc	tgagaccttc	840
	catcacctga	gccatctgga	aggcctgggtg	ctgaaggaca	gctctctcca	tacactgaac	900
	tcttctctgg	tccaaggtct	ggtcaacctc	tcggtgctgg	acctaagcga	gaactttctc	960
	tatgaaagca	tcaaccacac	caatgccttt	cagaacctaa	cccgcctgcg	caagctcaac	1020
30	ctgtccttca	attaccgcaa	gaagggtatcc	tttgcccgcc	tccacctggc	aagttccttc	1080
	aagaacctgg	tgtcactgca	ggagctgaac	atgaacggca	tcttcttccg	ctcgctcaac	1140
	aagtacacgc	tcagatggct	ggccgatctg	cccaaactcc	acactctgca	tcttcaaagt	1200
	aacttcatca	accaggcaca	gctcagcatc	tttggtacct	tccgagccct	tcgctttgtg	1260
	gacttgtcag	acaatcgcat	cagtgggctt	tcaacgctgt	cagaagccac	ccctgaagag	1320
35	gcagatgatg	cagagcagga	ggagctgttg	tctgcggatc	ctcacccagc	tccactgagc	1380
	accctgctt	ctaagaactt	catggacagg	tgtaagaact	tcaagttcac	catggacctg	1440
	tctcggaaca	acctggtgac	tatcaagcca	gagatgtttg	tcaatctctc	acgcctccag	1500
	tgtcttagcc	tgagccacaa	ctccattgca	caggctgtca	atggctctca	gttctctgcg	1560
	ctgactaatc	tgcaggtgct	ggacctgtcc	cataacaaac	tggacttgta	ccactggaaa	1620
40	tcgttcagtg	agctaccaca	gttgacggcc	ctggacctga	gctacaacag	ccagcccttt	1680
	agcatgaagg	gtataggcca	caatttcagt	tttgtggccc	atctgtccat	gctacacagc	1740

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cttagcctgg cacacaatga cattcatacc cgtgtgtcct cacatctcaa cagcaactca 1800
gtgaggtttc ttgacttcag cggcaacggt atggggccgca tgtgggatga ggggggcctt 1860
tatctccatt tcttccaagg cctgagtggc ctgctgaagc tggacctgtc tcaaaataac 1920
ctgcatatcc tccggcccca gaaccttgac aacctcccca agagcctgaa gctgctgagc 1980
5 ctccgagaca actacctatc tttctttaac tggaccagtc tgtccttcct gcccacactg 2040
gaagtcctag acctggcagg caaccagcta aaggccctga ccaatggcac cctgcctaata 2100
ggcaccctcc tccagaaact ggatgtcagc agcaacagta tctgtctgtg ggtcccagcc 2160
ttcttcgctc tggcggtcga gctgaaagag gtcaacctca gccacaacat tctcaagacg 2220
gtggatcgct cctggtttgg gccattgtg atgaacctga cagttctaga cgtgagaagc 2280
10 aacctctgac actgtgcctg tggggcagcc ttcgtagact tactgttgga ggtgcagacc 2340
aaggtgcctg gcctggctaa tgggtgtgaag tgtggcagcc ccggccagct gcagggccgt 2400
agcatcttcg cacaggacct gcggctgtgc ctggatgagg tctctctctg ggactgcttt 2460
ggcctttcac tcttggtgtg ggccgtgggc atggtggtgc ctatactgca ccatctctgc 2520
ggctgggacg tctgggtactg ttttcatctg tgcctggcat ggctaccttt gctggcccg 2580
15 agccgacgca gcgcccagc tctccctat gatgccttcg tgggtgttcga taaggcacag 2640
agcgcagttg cggactgggt gtataacgag ctgcgggtgc ggctggagga gcggcgcggt 2700
cgccgagccc tacgcttgtg tctggaggac cgagattggc tgcctggcca gacgctcttc 2760
gagaacctct gggcttccat ctatgggagc cgcaagactc tatttgtgct ggcccacacg 2820
gaccgcgtca gtggcctcct gcgcaccagc ttcctgctgg ctcagcagcg cctggttgaa 2880
20 gaccgcaagg acgtgggtgt gttggtgatc ctgcgtccgg atgccaccg ctcccgtat 2940
gtgcgactgc gccagcgtct ctgccgccag agtgtgctct tctggcccca gcagcccaac 3000
gggcaggggg gcttctgggc ccagctgagt acagccctga ctagggacaa ccgccacttc 3060
tataaccaga acttctgccc gggacctaca gcagaa 3096

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The deduced amino acid sequence for murine TLR9 (SEQ ID NO:3), comprising 1032 amino acid residues, is shown in Table 4 below in the aligned sequence comparison as mtlr932e.pep. The deduced amino acid sequence for human TLR9 (SEQ ID NO:6), comprising 1032 amino acid residues, is shown in Table 4 below in the aligned sequence comparison as htlr9.pro.

Table 4. Amino Acid Sequence of Murine and Human TLR9

```

. . . . . : . . . . . : . . . . . : . . . . . : . . . . . : 60
htlr9.pro  MGFCRSALHPLSLLVQAIMLAMTLALGTLPAFLPCELQPHGLVNCNWLFLKSVPHFSSMAA 60
mtlr932e.pep MVLRRRTLHPLSLLVQAAVLAETLALGTLPAFLPCELKPHGLVDCNWLFLKSVPFRFSAAA 60
35
. . . . . : . . . . . : . . . . . : . . . . . : . . . . . : 120
htlr9.pro  PRGNVTSLSLSSNRIHHLHDSDFAHLPRLHNLKWNCPVGLSPMHFPCHMTIEPSTFL 120
mtlr932e.pep SCSNITRLSLISNRIHHLHNSDFVHLSNLRQLNLKWNCPPTGLSPLHFSCHMTIEPRTFL 120
40
. . . . . : . . . . . : . . . . . : . . . . . : . . . . . : 180

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	htlr9.pro	AVPTLEELNLSYNNIMTVPALPKSLISLSLSHTNIIIMLDSASLAGLHALRFLFMDGNCYY	180
	mtlr932e.pep	AMRTLEELNLSYNGITTVPRLPSSLVNLSLSHTNIIIVLDANSLAGLYSLRVLFMDGNCYY	180
		. : . : . : . : . : . : . : . : . :	240
5	htlr9.pro	KNPCRQALEVAPGALLGLGNLTHLSLKYNNLTVPVPRNLPSSLEYLLLSYNRIVKLAPEDL	240
	mtlr932e.pep	KNPCTGAVKVTPGALLGLSNLTHLSLKYNNLTQVPRQLPPSLEYLLVSYNLIVKLGPEDL	240
		. : . : . : . : . : . : . : . : . :	300
	htlr9.pro	ANLTALRVLDVGGNCRCDHAPNCPMECPRHFPQLHPDFTSHLSRLEGLVLKDSSSLSWLN	300
10	mtlr932e.pep	ANLTSLRVLDVGGNCRCDHAPNPCIIEGQKSLHLHPETFHHLSHLEGLVLKDSSSLHTLN	300
		. : . : . : . : . : . : . : . : . :	360
	aa197442.pep	LNLSEFNRYKKVVSFARLHLASSF	22
	htlr9.pro	ASWFRGLGNLRVLDLSENFLYKCIITKTAFQGLTQLRKLNLSEFNRYKKVVSFAHLSLAPSF	360
15	mtlr932e.pep	SSWFQGLVNLVLDLSENFLYESINHTNAFQNLTRLRKLNLSEFNRYKKVVSFARLHLASSF	360
		. : . : . : . : . : . : . : . : . :	420
	mousepep1	C	1
	aa197442.pep	KNLVSLQELNMNGIFFRLLNKYTLRWLADLPKLHTLHLQMNFINQAQLSIFGTFRALRFV	82
20	htlr9.pro	GSILVALKELDMHGIFFRSLDETTLRPLARLPMLQTLRLQMNFINQAQLGIFRAFPGLRYV	420
	mtlr932e.pep	KNLVSLQELNMNGIFFRSLNKYTLRWLADLPKLHTLHLQMNFINQAQLSIFGTFRALRFV	420
		. : . : . : . : . : . : . : . : . :	480
	mousepep1	DLSDNRISGPSTLSEA	17
25	humanpep1	PAPVDTPSSEDFRPNCL	16
	aa197442.pep	DLSDNRISGPSTLSEATPEEADDAEQEELLSADPHAPLSTPASKNFMDRCKNFKFNMDL	142
	htlr9.pro	DLSDNRISGASELT-ATMGEADGGEKVWLQPGDLAPAPVDTPSSEDFRPNCLSTLNFTLDL	479
	mtlr932e.pep	DLSDNRISGPSTLSEATPEEADDAEQEELLSADPHAPLSTPASKNFMDRCKNFKFTMDL	480
		. : . : . : . : . : . : . : . : . :	540
30	aa197442.pep	SRNNLVTTITAEMFVNLSRLQCLSLSHNSIAQAVNGS	178
	htlr9.pro	SRNNLVTVQPEMFQQLSHLQCLRLSHNCISQAVNGSQFLPLTGLQVLDLSRNKLDLYHEH	539
	mtlr932e.pep	SRNNLVTIKPEMFVNLSRLQCLSLSHNSIAQAVNGSQFLPLTNLQVLDLSHNKLDLYHWK	540
		. : . : . : . : . : . : . : . : . :	600
35	aa162495.pep	YNSQPFMSMGIGHNFSFVTHLSMLQSLSLAHNDIHTRVSSHLNSNS	46
	htlr9.pro	SFTELPRLAALDLSYNSQPFMGQGVGHNFSFVAHLRTRLRHLSLAHNNIHSQVSQQLCSTS	599
	mtlr932e.pep	SFSELPQLQALDLSYNSQPFMSMGIGHNFSFVAHLSMLHSLSLAHNDIHTRVSSHLNSNS	600
		. : . : . : . : . : . : . : . : . :	660
40	aa162495.pep	VRFLDFSGNGMGRMWDEGGLYLHFFQGLSGVLKLDLSQNNLHILRPQNLDNLPKSLKLLS	106
	htlr9.pro	LRALDFSGNALGHMWAEGDLYLHFFQGLSGLIWLDSLQNNRLHTLLPQTLRNLPKSLQVLR	659
	mtlr932e.pep	VRFLDFSGNGMGRMWDEGGLYLHFFQGLSGLLKLDLSQNNLHILRPQNLDNLPKSLKLLS	660
		. : . : . : . : . : . : . : . : . :	720
45	aa162495.pep	LRDNYLSFFNWTSLSFLPNLEVLDLAGNQLKALTNGTLPNGTLLQKLDVSSNSIVS	162

1009-2

1009-2

ai451215.pep: SEQ ID NO:16; aa273731.pep: SEQ ID NO:10; ai463056.pep: SEQ ID NO:12; humanpep2: SEQ ID NO:20; and mousepep2: SEQ ID NO:18.

Example 2. Reconstitution of TLR9 signaling in 293 fibroblasts

The cloned mouse TLR9 cDNA (see above) and human TLR9 cDNA (gift from B. Beutler, Howard Hughes Medical Institute, Dallas, TX) in pT-Adv vector (from Clontech) were cloned into the expression vector pcDNA3.1(-) from Invitrogen using the EcoRI site. Utilizing a “gain of function” assay it was possible to reconstitute human TLR9 (hTLR9) and murine TLR9 (mTLR9) signaling in CpG DNA non-responsive human 293 fibroblasts (ATCC, CRL-1573). The expression vectors mentioned above were transfected into 293 fibroblast cells using the calcium phosphate method.

Since NF- κ B activation is central to the IL-1/TLR signal transduction pathway (Medzhitov R et al., *Mol Cell* 2:253-258 (1998); Muzio M et al., *J Exp Med* 187:2097-2101 (1998)), cells were transfected with hTLR9 or co-transfected with hTLR9 and a NF- κ B-driven luciferase reporter construct. Human fibroblast 293 cells were transiently transfected with (**Figure 1A**) hTLR9 and a six-times NF- κ B-luciferase reporter plasmid (NF- κ B-luc, kindly provided by Patrick Baeuerle, Munich, Germany) or (**Figure 1B**) with hTLR9 alone. After stimulus with CpG-ODN (2006, 2 μ M, TCGTTCGTTTTGTCTGTTTTGTCTGTT, SEQ ID NO:112), GpC-ODN (2006-GC, 2 μ M, TGCTGCTTTTGTGCTTTTGTGCTT, SEQ ID NO:118), LPS (100 ng/ml) or media, NF- κ B activation by luciferase readout (8h, **Figure 1A**) or IL-8 production by ELISA (48h, **Figure 1B**) were monitored. Results are representative of three independent experiments. **Figure 1** shows that cells expressing hTLR9 responded to CpG-DNA but not to LPS.

Figure 2 demonstrates the same principle for the transfection of mTLR9. Human fibroblast 293 cells were transiently transfected with mTLR9 and the NF- κ B-luc construct (**Figure 2**). Similar data was obtained for IL-8 production (not shown). Thus expression of TLR9 (human or mouse) in 293 cells results in a gain of function for CpG-DNA stimulation similar to hTLR4 reconstitution of LPS responses.

To generate stable clones expressing human TLR9, murine TLR9, or either TLR9 with the NF- κ B-luc reporter plasmid, 293 cells were transfected in 10 cm plates (2x10⁶ cells/plate) with 16 μ g of DNA and selected with 0.7 mg/ml G418 (PAA Laboratories GmbH,

Cölbe, Germany). Clones were tested for TLR9 expression by RT-PCR, for example as shown in **Figure 3**. The clones were also screened for IL-8 production or NF- κ B-luciferase activity after stimulation with ODN. Four different types of clones were generated.

- | | | |
|---|----------------|--|
| 5 | 293-hTLR9-luc: | expressing human TLR9 and 6-fold NF- κ B-luciferase reporter |
| | 293-mTLR9-luc: | expressing murine TLR9 and 6-fold NF- κ B-luciferase reporter |
| | 293-hTLR9: | expressing human TLR9 |
| | 293-mTLR9: | expressing murine TLR9 |

10 **Figure 4** demonstrates the responsiveness of a stable 293-hTLR9-luc clone after stimulation with CpG-ODN (2006, 2 μ M), GpC-ODN (2006-GC, 2 μ M), Me-CpG-ODN (2006 methylated, 2 μ M; TZGTZGTTTTGTZGTTTTGTZGTT, Z = 5-methylcytidine, SEQ ID NO:128), LPS (100 ng/ml) or media, as measured by monitoring NF- κ B activation. Similar results were obtained utilizing IL-8 production with the stable clone 293-hTLR9.

15 293-mTLR9-luc were also stimulated with CpG-ODN (1668, 2 μ M; TCCATGACGTTTCCTGATGCT, SEQ ID NO:84), GpC-ODN (1668-GC, 2 μ M; TCCATGAGCTTCCTGATGCT, SEQ ID NO:85), Me-CpG-ODN (1668 methylated, 2 μ M; TCCATGAZGTTTCCTGATGCT, Z = 5-methylcytidine, SEQ ID NO:207), LPS (100 ng/ml) or media, as measured by monitoring NF- κ B activation (**Figure 5**). Similar results were

20 obtained utilizing IL-8 production with the stable clone 293-mTLR9. Results are representative of at least two independent experiments. These results demonstrate that CpG-DNA non-responsive cell lines can be stably genetically complemented with TLR9 to become responsive to CpG DNA in a motif-specific manner. These cells can be used for screening of optimal ligands for innate immune responses driven by TLR9 in multiple species.

25 **Example 3. Expression of soluble recombinant human TLR9 in yeast cells (*Pichia pastoris*)**

Human TLR9 cDNA coding for amino acids 1 to 811 was amplified by PCR using the primers 5'-ATAGAATTCAATAATGGGTTTCTGCCGCAGCGCCCT-3' (SEQ ID NO:194)

30 and 5'-ATATCTAGATCCAGGCAGAGGCGCAGGTC-3' (SEQ ID NO:195), digested with EcoRI and XbaI, cloned into the yeast expression vector pPICZB (Invitrogen, Groningen,

Netherlands) and transfected into yeast cells (*Pichia pastoris*). Clones were selected with the antibiotic zeozin and protein production of soluble human TLR9 was induced with methanol (see **Figure 6**: SDS-PAGE, Coomassie stained, arrow marks hTLR9; lane 1: supernatant of culture induced with methanol; lane 2: supernatant of culture not induced). Thus TLR9 protein can be isolated from transfectants and further utilized for protein studies and vaccination purposes.

Example 4. hTLR9 expression correlates with CpG-DNA responsiveness.

Bacterial DNA has been described as a mitogen for both murine and human B cells. Although LPS is also mitogenic for murine B cells, it is generally accepted that LPS is not a mitogen for human B cells. **Figure 7** demonstrates that human B cells proliferate after stimulation with *E. coli* DNA or a CpG-ODN but not Dnase-digested *E. coli* DNA or a control GpC-ODN. Purified human B cells were stimulated with 50µg/ml *E. coli* DNA, a comparable amount of DNase I-digested *E. coli* DNA, 2µM CpG-ODN (2006), 2µM GpC-ODN (2006-GC) or 100 ng/ml LPS. B cell proliferation was monitored at day two by ³H-thymidine uptake. These data demonstrate that it was DNA within the *E. coli* DNA preparation that was mitogenic and that a CpG-motif within the ODN was required.

Human dendritic cells (DC) have been claimed to be responsive to CpG-DNA. While analyzing human dendritic cell responses to CpG-DNA, we noted that plasmacytoid DC (CD123+DC) produced IFN-α, TNF, GM-CSF, and IL-8 upon exposure to CpG-DNA but not to LPS (**Figure 8** and unpublished data). The converse was true for stimulation of monocyte-derived dendritic cells (MDDC) (**Figure 8** and unpublished data). Purified CD123+DC or MDDC were stimulated with 50µg/ml *E. coli* DNA, a comparable amount of DNase I-digested *E. coli* DNA, 2µM CpG-ODN (2006), 2µM GpC-ODN (2006-GC) or 100 ng/ml LPS (**Figure 8**). IL-8 and TNF concentration was determined by enzyme-linked immunosorbent assay (ELISA). The CD123+DC response was DNA- and CpG-motif restricted. Monocyte-derived dendritic cells (MDDC) however demonstrated the converse response pattern, a response to LPS but not CpG-DNA. Due to this segregated response we analyzed TLR expression.

We have shown that CpG-DNA utilizes the Toll/IL-1R (TIR) signal transduction pathway implying the need for a TIR domain in the CpG-DNA signaling receptor. Häcker H

et al., *J Exp Med* 192:595-600 (2000). It was further demonstrated that TLR9-deficient mice are non-responsive to CpG-ODN. Hemmi H et al., *Nature* 408:740-5. By semi-quantitative RT-PCR both B cells and CD123+ DC yielded positive signals for hTLR9 while MDDC, monocytes and T cells were weak to negative (**Figure 9**). The cDNAs were prepared from monocyte-derived dendritic cells (MDDC), lane 1; purified CD14+ monocytes, lane 2; B cells, lane 3; CD123+ DC, lane 4; CD4+ T cells, lane 5; and CD8+ T cells, lane 6. cDNA amounts were normalized based on GAPDH amount determined by TAG-MAN PCR (Perkin-Elmer). RT-PCR was performed for 30 cycles on normalized cDNA diluted 1:5 for human TLR2, 4 and 9, while GAPDH was diluted 1:125. We also tested for hTLR2 and hTLR4 expression. MDDC and monocytes were positive while B cells, T cells and CD123+DC were weak to negative (**Figure 9**). Weak signals delivered by PCR could be explained by contaminating cells, however a strong positive signal implies expression. These data demonstrated a clear correlation between hTLR9 mRNA expression and B cell or CD123+DC responsiveness to CpG-DNA (**Figures 7 and 8**). A correlation could also be shown for hTLR2 and hTLR4 expression and MDDC responsiveness to LPS (**Figure 8**). This data demonstrates that hTLR9 is a relevant receptor for CpG-DNA responses and that its expression determines responsiveness. If TLR9 expression could be modulated, agonism or antagonism of CpG-DNA responses could be achieved.

Example 5. Species specificity of TLR9 signaling

By iterative examination of the flanking sequences surrounding CG dinucleotides, CpG-motifs have been identified. Paradoxically, or by twist of nature, the human optimal CpG-motif, GTCGTT (SEQ ID NO:66), is different from the murine motif, GACGTT (SEQ ID NO:129). Human peripheral blood mononuclear cells (PBMC) (**Figure 10A**) and murine splenocytes (**Figure 10B**) were stimulated with ODN 2006 (filled circle, TCGTCGTTTTGTCGTTTTGTCGTT, SEQ ID NO:112), ODN 2006-GC (open circle), ODN 1668 (filled triangle, TCCATGACGTTCTGATGCT, SEQ ID NO:84) or ODN 1668-GC (open triangle, TCCATGAGCTTCCTGATGCT, SEQ ID NO:85) at indicated concentrations and IL-12 production was monitored after 8 hours. **Figure 10A** shows that titration of the optimal human ODN, 2006, on PBMC induces IL-12 production. The optimal murine sequence, 1668, however was much less effective in eliciting IL-12 from PBMC. The two

control GpC-ODNs were essentially negative. The converse was true for murine splenocytes (Figure 10B), in that the murine sequence induced optimal IL-12 while the human sequence was much less effective. It should also be noted that the K_{ac} (concentration of half-maximal activation) of murine splenocytes for 1668 was greater than human PBMC for 2006 (compare Fig. 10A to Fig. 10B).

Because stable TLR9 transfectants mirrored primary cell responsiveness to CpG-DNA (Figures 4 and 5), it was hypothesized that stable transfectants could potentially discern species-specific CpG-motifs through TLR9 receptors. Therefore 293-hTLR9-luc (expressing human TLR9 and 6-fold NF- κ B-luc reporter), 293-mTLR9-luc (expressing murine TLR9 and 6-fold NF- κ B-luc reporter), 293-hTLR9 (expressing human TLR9) and 293-mTLR9 (expressing murine TLR9) clones were tested for CpG-DNA motif responsiveness. Figure 11 shows titration curves for 2006 or 1668 and their controls versus either hTLR9 or mTLR9 cells. Depicted are both NF- κ B-driven luciferase and IL-8 production as readout. In both 293 hTLR9-luc and 293-mTLR9-luc cells stimulation with CpG-DNA resulted in NF- κ B activation, as determined by measurement of the induced expression of firefly luciferase under the control of a minimal promotor containing six tandem NF- κ B-binding sites. After lysis of the cells luciferase can be detected photometrically based on an enzymatic reaction by luciferase which creates photons. IL-8 production was monitored using enzyme-linked immunosorbent assay (ELISA). Figure 11 depicts clones stimulated with ODN 2006 (filled circle), ODN 2006-GC (open circle), ODN 1668 (filled triangle) or ODN 1668-GC (open triangle) at indicated concentrations and NF- κ B activation or IL-8 production were measured after 10 and 48 hours, respectively. Results shown in Figure 11 are representative of three independent experiments. Strikingly, CpG-motif sequence specificity was conferred in a species-specific manner by TLR9. Additionally, the half-maximal concentration for either 2006 or 1668 appears nearly the same as those determined on primary cells (compare Figure 10 and Figure 11). These data demonstrate that TLR9 is the CpG-DNA receptor and that exquisite specificity to CpG-DNA sequence is conferred by TLR9.

Example 6. Use of stable TLR9 clones to test responsiveness to substances other than phosphorothioate ODN

As described in the foregoing Examples, the stable TLR9 clones were initially

screened for fidelity of phosphorothioate CpG-ODN reactivity. The 293-hTLR9 cells demonstrated reactivity to CpG-DNA and not LPS in a CpG-motif dependent manner (Figures 4 and 5). In the present example the stable TLR9 transfectants were tested for responsiveness to additional DNAs. NF- κ B activation was monitored after stimulation with *E. coli* DNA (black bars) or *E. coli* DNA digested with DNase I (gray bars) in 293-hTLR9-luc cells. Figure 12 demonstrates an *E. coli* DNA dose-dependent induction of NF- κ B-driven luciferase expression to a level comparable to phosphorothioate CpG-ODN (Figure 11). Activity was destroyed by DNase I digestion, indicating specificity of response to DNA and not contaminant bacterial products. The stable TLR9 transfectants can be used to screen the activity of DNAs from various species or vector DNAs intended for immune system stimulation. In particular, TLR9 transfectants can be used to screen and compare the immunostimulatory activity of DNAs from various species of pathogens, DNA constructs, DNAs intended for use as vaccines, gene replacement therapeutics, and nucleic acid vectors.

293-hTLR9-luc cells also were stimulated with the phosphodiester variants of ODN 2006 (filled circle), ODN 2006-GC (open circle), ODN 1668 (filled triangle) or ODN 1668-GC (open triangle) at indicated concentrations, and NF- κ B activation was monitored after 12 hours (Figure 13A). Likewise, 293-mTLR9-luc cells were stimulated with the phosphodiester variants of ODN 2006 (filled circle), ODN 2006-GC (open circle), ODN 1668 (filled triangle) or ODN 1668-GC (open triangle) at indicated concentrations, and NF- κ B activation was monitored after 12 hours (Figure 13B). These assays show that the stable TLR9 transfectants responded to DNAs other than phosphorothioate-modified ODN. These data demonstrate the utility of stable TLR transfectants for screening for agonists of the TLR9 receptor.

Example 7. TLR9 determines CpG-ODN activity

Although 2006 and 1668 are discussed in terms of CpG-motif differences, they are very different in several aspects (see Table 5 for comparison). The lengths are different, 24 versus 20 nucleotides, and 2006 has four CG dinucleotides compared to one in 1668. Additional differences are the CG position relative to the 5' and 3' ends and also 5' sequence differences. In order to determine if motif specificity is a quality of the motif and not the global sequence environment, for this experiment several sequences were produced holding

these variables constant. As a starting point, the 1668 sequence was modified by converting the central C to T and the distal TG to CG, thereby creating a second CG in the resulting sequence 5000 (SEQ ID NO:130, Table 5). Then point nucleotide changes were made, progressing toward a 2006-like sequence, 5007 (SEQ ID NO:98). The ODN 5002 (SEQ ID NO:132) is most like 1668 with the exception that C's at positions 12 and 19 have been converted to T's. The last 16 nucleotides of ODN 5007 are the same as the last 15 nucleotides of 2006 with the exception of an additional T. The ODN concentration of half-maximal activation (K_{ac}) was determined by producing ODN titration curves using either 293-hTLR9-luc or 293-mTLR9-luc cells and NF- κ B-driven luciferase expression as a readout. Example curves are given in **Figure 14**. Stable transfectants 293-hTLR9-luc and 293-mTLR9-luc were stimulated with ODN 5002 (filled circle) or ODN 5007 (open circle) at indicated concentrations and NF- κ B activation was monitored after 12 hours. Results shown in **Figure 14** are representative of three independent experiments. Values for K_{ac} for multiple ODN are given in Table 5. Similar results were obtained for those ODN tested with 293-hTLR9 and 293-mTLR9 cells utilizing IL-8 as readout.

Table 5. CpG-DNA sequence specificity of human and murine TLR9 signaling activity

CpG-DNA	Sequence	293-hTLR9	293-mTLR9	SEQ ID NO:
		K_{ac} (nM)	K_{ac} (nM)	
1668	TCCATGACGTTCTTGATGCT	>10,000	70	84
1668-GC	TCCATGAGCTTCCTGATGCT	>10,000	>10,000	85
2006	<u>TCGTCGTTTTGTCGTTTTGTCGTT</u>	400	>10,000	112
2006-GC	TGCTGCTTTTGTGCTTTTGTGCTT	>10,000	>10,000	118
5000	TCCATGACGTTCTTGACGCT	10,000	82	130
5001	TCCATGACGTTCTTGACGTT	7,000	55	131
5002	TCCATGACGTTCTTGATGTT	7,000	30	132
5003	TCCATGACGTTTTTGATGTT	10,000	30	133
5004	TCCATGTCGTTCTTGATGTT	5,000	400	134
5005	TCCATGTCGTTTTTGATGTT	3,000	2,000	135
5006	TCCATGTCGTTTTTGTTGTT	3,000	650	136
5007	TCCATGTCGTTTTGTCGTT	700	1,000	98
5002	TCCATGACGTTCTTGATGTT	ND	30	132
5008	TCCATGACGTTATTGATGTT	ND	40	137
5009	TCCATGACGTCCTTGATGTT	ND	>10,000	138
5010	TCCATGACGTCATTGATGTT	ND	>10,000	139

In previous unpublished work by the inventors, it had been noted that a CA substitution converting the mouse CpG-motif from GACGTTC to GACGTCA was deleterious. To extend our examination of the motif, three more ODN were created to dissect this effect (5008-5010, SEQ ID NOs:137-139, Table 5).

5 The activity displayed by the 293-hTLR9-luc clone increased with progressive nucleotide substitutions converting the mouse sequence toward the human sequence (Table 5, sequences 5000-5007). The converse was true for the 293-mTLR9-luc clone, which showed highest activity for the mouse sequences. The originally hypothesized CpG-motif was purine-purine-CG-pyrimidine-pyrimidine. Most notable to motif definition as determined by TLR9
10 genetic complementation was the non-conservative pyrimidine for purine change A to T immediately 5' of the CG (Table 5). These changes improved 293-hTLR9-luc responsiveness but diminished 293-mTLR9-luc responsiveness. These results support the notion that the preferred mouse motif contains ACG while the preferred human sequence contains TCG. The conservative pyrimidine for pyrimidine change T to C in the mouse motif, ACGTT versus ACGTC (5002 versus 5009), completely destroyed 293-mTLR9 responsiveness.
15 Although not a complete iterative analysis of the CpG-motif, the data refine our understanding of the motif. More importantly these data strongly support direct CpG-motif engagement by TLR9.

20 **Example 8. Antagonist definition**

It has been demonstrated that DNA uptake and endosomal maturation are required for signal initiation by CpG-DNA. It has been hypothesized that in order for DNA to enter the endosomal/lysosomal compartment a non-CpG dependent uptake receptor may be required. 293 cells were transiently transfected with mTLR9 treated with either medium only or 1.0 μ M
25 CpG-ODN 1668 (**Figure 15**). Additionally the 1668-treated TLR9 transfectants were simultaneously exposed to various doses of a non-CpG ODN (PZ2; 5'-CTCCTAGTGGGGGTGTCCTAT-3', SEQ ID NO:43). IL-8 production was monitored after 48h by ELISA. **Figure 15** shows that PZ2, in a dose-dependent manner, was able to antagonize the activation of TLR9-transfected cells stimulated with a CpG ODN.

30 **Figure 16** demonstrates that the stable TLR9 transfectants, 293-hTLR9-luc cells, are sensitive to non-CpG-ODN blockade. 293-hTLR9-luc cells were incubated with CpG-ODN

(0.5 μ M) (black bars) or TNF- α (10 ng/ml) (gray bars) and increasing concentrations of a blocking ODN (5'-HHHHHHHHHHHHHHHWGGGGG-3', SEQ ID NO:140; H = A, T, C; W = A, T) as indicated. NF- κ B activation was monitored after 12 hours and is presented as percent yields. Thus both mTLR9 and hTLR9 activity can be blocked by non-stimulatory ODN. The blockade is specific to blocking ODN since the TNF-driven NF- κ B signal was not diminished. Antagonism of CpG-DNA responses could thus be defined in stable TLR9 cells and therefore high throughput screening can be done for TLR9 antagonist.

Bafilomycin A poisons the proton pump needed for H⁺ transport into endosomes, which is required for endosomal maturation. **Figure 17** shows that blockade of endosomal maturation in 293-hTLR9-luc cells fully blocks CpG-ODN induction of NF- κ B. 293-hTLR9-luc cells were preincubated with 10 nM Bafilomycin A (gray bars) or dimethylsulfoxide (DMSO) control (black bars) for 30 min and stimulated with CpG-ODN (2006, 0.5 μ M), IL-1 (10 ng/ml) or TNF- α (10 ng/ml) as indicated. NF- κ B activation was monitored after 12 hours and is presented as percent yields. The blockade was specific to CpG-DNA generated signal because both IL-1 and TNF induction of NF- κ B was unaffected. These data demonstrate that 293 cells stably complemented with hTLR9 behave in a manner similar to primary CpG-DNA responsive cells, in that cellular uptake and endosomal maturation are required for induction of signal by CpG-DNA. Thus the stable transfectants can be used as indicator for TLR9 drug antagonist.

CpG-DNA signaling appears to occur via a Toll/IL-1R-like pathway. It was shown in the mouse that CpG-DNA signaling is dependent on MyD88, IRAK and TRAF6. Häcker H et al., *J Exp Med* 192:595-600 (2000). Hemmi et al. demonstrated that mTLR9-deficient mice lack activation of IRAK upon CpG-ODN stimulation. Hemmi H et al., *Nature* 408:740-5 (2000). **Figure 18** shows that CpG-DNA signaling via human TLR9 was MyD88 dependent. hTLR9 (293-hTLR9) was co-transfected with a six-times NF- κ B luciferase reporter plasmid and increasing concentrations of the dominant negative human MyD88 expression vector. Cells were not stimulated (filled circles), stimulated with CpG-ODN (2006, 2 μ M) (open circles) or TNF- α (10 ng/ml) (filled triangles) and NF- κ B activation was monitored after 12 hours. Results are representative of at least two independent experiments.

Figure 18 demonstrates that dominant negative MyD88 blocks NF- κ B induction in 293-hTLR9 cells following CpG-DNA stimulation. The blockade of MyD88 did not affect NF-

κB induction via TNF induced signal transduction. In general these data confirm the central role of MyD88 to TLR signaling and specifically the role of MyD88 in CpG-DNA initiation of signal. Thus human cells transfected with TLR9 can be used as indicators to find molecules to antagonize CpG-DNA via genetic mechanisms.

5

Example 9. Antibody production

Peptides for human and mouse TLR9 were designed for coupling to a carrier protein and injected into rabbits to obtain anti-peptide polyclonal antisera. Mouse peptide 1 (mousepep1, see Table 4) can be found in EST aa197442 and peptide 2 (mousepep2, see
10 Table 4) in EST aa273731 and ai463056. Human peptide 1 (humanpep1, see Table 4) and peptide 2 (humanpep2, see Table 4) were taken from the published human sequence.

Three rabbit antisera were generated by this method: anti-mousepep1, specific for the extracellular domain of murine TLR9; anti-humanpep1, specific for the extracellular domain of hTLR9; and antisera against a combination of mousepep2 and humanpep2, specific for the
15 cytoplasmic domain of both murine and human TLR9. Immunoprecipitates with anti-FLAG antibody were electrophoresed by PAGE and, using standard Western blotting techniques, transferred to membrane and probed with the various antisera. **Figure 19** shows the response to hTLR9-FLAG and mTLR9-FLAG. The TLR9 in these blots are indicated with arrows, while the lower molecular weight bands represent anti-FLAG antibody.

20

Example 10. Mutation adjacent to the CXXC-domain (hTLR9-CXXCm, mTLR9-CXXCh)

The CXXC motif resembles a zinc finger motif and is found in DNA-binding proteins and in certain specific CpG binding proteins, e.g. methyl-CpG binding protein-1 (MBD-1).
25 Fujita N et al., *Mol Cell Biol* 20:5107-5118 (2000). Human and murine TLR9 contain two CXXC motifs. The CXXC domain is highly conserved between human and murine TLR9 but followed by 6 amino acids (aa) which differ quite substantially in polarity and size. By the use of a site-specific mutagenesis kit (Stratagene, La Jolla, CA, USA) these six amino acid residues (human: PRHFPQ 269-274); mouse: GQKSLH 269-274) were interchanged between
30 human and murine TLR9. These mutations were generated by the use of the primers 5'-CTGCATGGAGTGCGGCCAAAAGTCCCTCCACCTACATCCCGATAC-3' (SEQ ID

NO:141) and

5'-GTATCGGGATGTAGGTGGAGGGACTTTTGGCCGCACTCCATGCAG-3' (SEQ ID NO:142) for human TLR9 and the primers

5'-CTGTATAGAATGTCCTCGTCACTTCCCCCAGCTGCACCCTGAGAC-3' (SEQ ID NO:143) and

5'-GTCTCAGGGTGCAGCTGGGGGAAGTGACGAGGACATTCTATACAG-3' (SEQ ID NO:144) for murine TLR9 according to the manufacturer's protocol.

CXXC motif:	CXXCXXXXXXXXCXXC		SEQ ID NO:145
Wildtype hTLR9:	CRRCDHAPNPCMECPRHFPQ	aa 255-274	SEQ ID NO:146
hTLR9-CXXCm:	CRRCDHAPNPCMECGQKSLH	aa 255-274	SEQ ID NO:147
Wildtype mTLR9:	CRRCDHAPNPCMICGQKSLH	aa 255-274	SEQ ID NO:148
mTLR9-CXXCh:	CRRCDHAPNPCMICPRHFPQ	aa 255-274	SEQ ID NO:149

For the stimulation of the hTLR9 variant hTLR9-CXXCm, 293 cells were transiently transfected with hTLR9 or hTLR9-CXXCm and stimulated after 16 hours with ODN 2006 and ODN 1668 at concentrations indicated (**Figure 20**). 48 hours after stimulation supernatant was harvested and IL-8 production was measured by ELISA. The data show that hTLR9 can be improved by converting the human CXXC domain to the murine CXXC domain. For the stimulation of the mTLR9 variant mTLR9-CXXCh, 293 cells were transiently transfected with mTLR9 or mTLR9-CXXCh and stimulated after 16 hours with ODN 2006 and ODN 1668 at concentrations indicated (**Figure 21**). 48 hours after stimulation supernatant was harvested and IL-8 production was measured by ELISA. It appears that the human CXXC domain may diminish mTLR9-CXXCh activity relative to the wild type mTLR9.

Example 11. Mutation in the MBD motif (hTLR9-MBDmut, mTLR9-MBDmut)

The MBD motif is a domain recently described for CpG binding in the protein MBD-1. Fujita N et al., *Mol Cell Biol* 20:5107-5118 (2000); Ohki I et al., *EMBO J* 18:6653-6661 (1999). Human and murine TLR9 contain this motif at position 524-554 and 525-555, respectively.

MBD-1	R-XXXXXXX-R-X-D-X-Y-XXXXXXXXXX-R-S-XXXXXX-Y	SEQ ID NO:125
hTLR9	Q-XXXXXXX-K-X-D-X-Y-XXXXXXXXXX-R-L-XXXXXX-Y	SEQ ID NO:126
mTLR9	Q-XXXXXXX-K-X-D-X-Y-XXXXXXXXXX-Q-L-XXXXXX-Y	SEQ ID NO:127

The core of this domain consists of D-L-Y in human TLR9 (aa 534-536) and mouse TLR9 (aa 535-537). Through site-specific mutagenesis D534 and Y536 in human TLR9, and D535 and Y537 in murine TLR9, were mutated to alanines creating the sequence A-L-A for human (aa 534-536) and murine TLR9 (aa 535-537). These mutations were generated by the use of the primers 5'-CACAATAAGCTGGCCCTCGCCCACGAGCACTC-3' (SEQ ID NO:150) and 5'-GAGTGCTCGTGGGCGAGGGCCAGCTTATTGTG-3' (SEQ ID NO:151) for human TLR9 and the primers 5'-CATAACAACTGGCCTTGGCCCACTGGAAATC-3' (SEQ ID NO:152) and 5'-GATTTCAGTGGGCCAAGGCCAGTTTGTATG-3' (SEQ ID NO:153) for murine TLR9 according to the manufacturer's protocol.

For the stimulation of mTLR9 variant, mTLR9-MBDmut, 293 cells were transiently transfected with mTLR9 or mTLR9-MBD-mut and stimulated after 16 hours with ODN 2006 and ODN 1668 at concentrations indicated (**Figure 22**). 48 hours after stimulation supernatant was harvested and IL-8 production was measured by ELISA. For the stimulation of hTLR9 variant, hTLR9-MBDmut, 293 cells were transiently transfected with hTLR9 or hTLR9-MBD-mut and stimulated after 16 hours with ODN 2006 and ODN 1668 at concentrations indicated (**Figure 23**). 48 hours after stimulation supernatant was harvested and IL-8 production was measured by ELISA. The disruption of the putative CpG binding domain DXY in TLR9 destroyed receptor activity. These data demonstrate that the MBD motif is most likely involved in CpG-DNA binding and can be thus be manipulated to better understand CpG-DNA binding and efficacy.

Example 12. Proline to Histidine mutation in the TIR-domain (hTLR9-PHmut, mTLR9-PHmut)

Toll-like receptors have a cytoplasmic Toll/IL-1 receptor (TIR) homology domain which initiates signaling after binding of the adapter molecule MyD88. Medzhitov R et al., *Mol Cell* 2:253-8 (1998); Kopp EB et al., *Curr Opin Immunol* 11:15-8 (1999). Reports by

others have shown that a single-point mutation in the signaling TIR-domain in murine TLR4 (Pro712 to His) or human TLR2 (Pro681 to His) abolishes host immune response to lipopolysaccharide or gram-positive bacteria, respectively. Poltorak A et al., *Science* 282:2085-8 (1998); Underhill DM et al., *Nature* 401:811-5 (1999). Through site-specific mutagenesis the equivalent Proline at position 915 of human and murine TLR9 were mutated to Histidine (Pro915 to His). These mutations were generated by the use of the primers 5'-GCGACTGGCTGCATGGCAAACCCCTCTTTG-3' (SEQ ID NO:154) and 5'-CAAAGAGGGTTTTGCCATGCAGCCAGTCGC-3' (SEQ ID NO:155) for human TLR9 and the primers 5'-CGAGATTGGCTGCATGGCCAGACGCTCTTC-3' (SEQ ID NO:156) and 5'-GAAGAGCGTCTGGCCATGCAGCCAATCTCG-3' (SEQ ID NO:157) for murine TLR9 according to the manufacturer's protocol.

For the stimulation of mTLR9 variant, mTLR9-PHmut, 293 cells were transiently transfected with mTLR9 or mTLR9-PHmut and stimulated after 16 hours with ODN 2006 and ODN 1668 at concentrations indicated (**Figure 22**). 48 hours after stimulation supernatant was harvested and IL-8 production was measured by ELISA. For the stimulation of hTLR9 variant, hTLR9-PHmut, 293 cells were transiently transfected with hTLR9 or hTLR9-PHmut and stimulated after 16 hours with ODN 2006 and ODN 1668 at concentrations indicated (**Figure 23**). 48 hours after stimulation supernatant was harvested and IL-8 production was measured by ELISA. These data demonstrate that TLR9 activity can be destroyed by the Pro to His mutation. This mutation has the potential to be used as a dominant negative to block TLR9 activity thus a genetic variant could compete for ligand or signaling partners and disrupt signaling.

Example 13. Exchange of the TIR-domain between murine and human TLR9 (hTLR9-TIRm, mTLR9-TIRh)

Toll-like receptors have a cytoplasmic Toll/IL-1 receptor (TIR) homology domain that initiates signaling after binding of the adapter molecule MyD88. Medzhitov R et al., *Mol Cell* 2:253-8 (1998); Kopp EB et al., *Curr Opin Immunol* 11:15-8 (1999). This is also true for TLR9. To generate molecules consisting of human extracellular TLR9 and murine TIR domain (hTLR9-TIRm) or murine extracellular TLR9 and human TIR domain (mTLR9-TIRh), the following approach was chosen. Through site-specific mutagenesis a ClaI

restriction site was introduced in human and murine TLR9. For human TLR9 the DNA sequence 5'-GGCCTCAGCATCTTT-3' (3026-3040, SEQ ID NO:158) was mutated to 5'-GGCCTATCGATT-3' (SEQ ID NO:159), introducing a ClaI site (underlined in the sequence) but leaving the amino acid sequence (GLSIF, aa 798-802) unchanged. For murine TLR9 the DNA sequence 5'-GGCCGTAGCATCTTC-3' (2434-2447, SEQ ID NO:160) was mutated to 5'-GGCCTATCGATT-3' (SEQ ID NO:161), introducing a ClaI site and creating the amino acid sequence (GLSIF, aa 799-803) which differs in one position (aa 800) from the wildtype murine TLR9 sequence (GRSIF, aa 799-803) but is identical to the human sequence.

hTLR9-TIRm. The primers used for human TLR9 were 5'-CAGCTCCAGGGCCTATCGATTTTGCACAGGACC-3' (SEQ ID NO:162) and 5'-GGTCCTGTGCAAAAATCGATAGGCCCTGGAGCTG-3' (SEQ ID NO:163). For creating an expression vector containing the extracellular portion of human TLR9 connected to the murine TIR domain, the human expression vector was cut with ClaI and limiting amounts of EcoRI and the fragment coding for the murine TIR domain generated by a ClaI and EcoRI digestion of murine TLR9 expression vector was ligated in the vector fragment containing the extracellular portion of hTLR9. Transfection into *E.coli* yielded the expression vector hTLR9-TIRm (human extracellular TLR9-murine TIR-domain).

mTLR9-TIRh. The primers used for murine TLR9 were 5'-CAGCTGCAGGGCCTATCGATTTTCGCACAGGACC-3' (SEQ ID NO:164) and 5'-GGTCCTGTGCGAAAATCGATAGGCCCTGCAGCTG-3' (SEQ ID NO:165). For creating an expression vector containing the extracellular portion of murine TLR9 connected to the human TIR domain, the murine expression vector was cut with ClaI and limiting amounts of EcoRI and the fragment coding for the human TIR domain generated by a ClaI and EcoRI digestion of human TLR9 expression vector was ligated in the vector fragment containing the extracellular portion of mTLR9. Transfection into *E.coli* yielded the expression vector mTLR9-TIRh (murine extracellular TLR9-human TIR-domain).

For the stimulation of the mTLR9 variant, mTLR9-TIRh, 293 cells were transiently transfected with mTLR9 or mTLR9-TIRh and stimulated after 16 hours with ODN 2006 and ODN 1668 at concentrations indicated (**Figure 24**). 48 hours after stimulation supernatant was harvested and IL-8 production was measured by ELISA. For the stimulation of the

hTLR9 variant, hTLR9-TIRm, 293 cells were transiently transfected with hTLR9 or hTLR9-TIRm and stimulated after 16 hours with ODN 2006 and ODN 1668 at concentrations indicated (**Figure 25**). 48 hours after stimulation supernatant was harvested and IL-8 production was measured by ELISA. Replacement of the murine TLR9-TIR domain with human does not significantly affect mTLR9 activity. Replacement of the human TLR9-TIR with murine however appears to have a negative effect on hTLR9. These data demonstrate that manipulations could be made to influence TLR9 activities.

Example 14. TLR9-fusion protein with green-fluorescent-protein (hTLR9-GFP, mTLR9-GFP)

Human and murine TLR9 were individually cloned into the vector pEGFP-N1 (Clontech, Palo Alto, CA, USA) to create expression vectors encoding human and murine fusion proteins consisting of an N-terminal TLR9 protein fused to C-terminal green-fluorescent protein (GFP). These constructs can be used to trace TLR9 localization and expression. Such detections can be used for staining in FACS analysis, confocal microscopy and Western blot, or for purification of polypeptides and subsequent antibody production.

Example 15. TLR9-fusion protein with FLAG-peptide (hTLR9-FLAG, mTLR9-FLAG)

Human and murine TLR9 were individually cloned into the vector pFLAG-CMV-1 (Sigma, St. Louis, MO, USA) to create expression vectors encoding human and murine fusion proteins consisting of an N-terminal leader peptide (preprotrypsin, which is cleaved intracellularly during processing of the protein), FLAG-peptide (DYKDDDDK) and TLR9 protein which does not contain its own signal peptide. These constructs can be used to trace TLR9 localization and expression, e.g., using anti-FLAG antibodies. Such detections can be used for staining in FACS analysis, confocal microscopy and Western blot, or for purification of polypeptides and subsequent antibody production.

Example 16. Method of cloning human TLR7

Two accession numbers in the GenBank database, AF245702 and AF240467, describe the DNA sequence for human TLR7. To create an expression vector for human TLR7, human TLR7 cDNA was amplified from a cDNA made from human peripheral mononuclear

blood cells (PBMC) using the primers 5'-CACCTCTCATGCTCTGCTCTCTTC-3' (SEQ ID NO:166) and 5'-GCTAGACCGTTTCCTTGAACACCTG-3' (SEQ ID NO:167). The fragment was cloned into pGEM-T Easy vector (Promega), cut with the restriction enzyme NotI and ligated into a NotI-digested pCDNA3.1 expression vector (Invitrogen). The insert was fully sequenced and translated into protein. The cDNA sequence for hTLR7 is SEQ ID NO:168, is presented in Table 6. The open reading frame starts at base 124, ends at base 3273, and codes for a protein of 1049 amino acids. SEQ ID NO:169 (Table 7), corresponding to bases 124-3273 of SEQ ID NO:168 (Table 6), is the coding region for the polypeptide of SEQ ID NO:170 (Table 8).

The protein sequence of the cloned hTLR7 cDNA matches the sequence described under the GenBank accession number AF240467. The sequence deposited under GenBank accession number AF245702 contains two amino acid changes at position 725 (L to H) and 738 (L to P).

Table 6. cDNA Sequence for Human TLR7 (5' to 3'; SEQ ID NO:168)

agctggctag	cgtttaaacg	ggccctctag	actcgagcgg	ccgcgaattc	actagtgatt	60
cacctctcat	gctctgctct	cttcaaccag	acctctacat	tccatttttg	aagaagacta	120
aaaatggtgt	ttccaatgtg	gacactgaag	agacaaattc	ttatcctttt	taacataatc	180
ctaatttcca	aactccttgg	ggctagatgg	tttccataaa	ctctgccttg	tgatgtcact	240
ctggatgttc	caaagaacca	tgtgatcggt	gactgcacag	acaagcattt	gacagaaatt	300
cctggaggta	ttcccacgaa	caccacgaac	ctcaccctca	ccattaacca	cataccagac	360
atctccccag	cgtcctttca	cagactggac	catctggtag	agatcgattt	cagatgcaac	420
tgtgtaccta	ttccactggg	gtcaaaaaac	aacatgtgca	tcaagaggct	gcagattaaa	480
cccagaagct	ttagtggact	cacttattta	aaatcccttt	acctggatgg	aaaccagcta	540
ctagagatac	cgcagggcct	cccgcctagc	ttacagcttc	tcagccttga	ggccaacaac	600
atcttttcca	tcagaaaaga	gaatctaaca	gaactggcca	acatagaaat	actctacctg	660
ggccaaaact	gttattatcg	aaatccttgt	tatgtttcat	attcaataga	gaaagatgcc	720
ttcctaaact	tgacaaagtt	aaaagtgtct	tccctgaaag	ataacaatgt	cacagccgtc	780
cctactgttt	tgccatctac	tttaacagaa	ctatatctct	acaacaacat	gattgcaaaa	840
atccaagaag	atgattttta	taacctcaac	caattacaaa	ttcttgacct	aagtggaaat	900
tgcctcgtt	gttataatgc	cccatttctt	tgtgcgccgt	gtaaaaataa	ttctccccta	960
cagatccctg	taaatgcttt	tgatgcgctg	acagaattaa	aagttttacg	tctacacagt	1020
aactctcttc	agcatgtgcc	cccaagatgg	tttaagaaca	tcaacaaact	ccaggaactg	1080
gatctgtccc	aaaacttctt	ggccaaagaa	attggggatg	ctaaatttct	gcattttctc	1140
cccagcctca	tccaattgga	tctgtctttc	aattttgaac	ttcaggtcta	tcgtgcatct	1200
atgaatctat	cacaagcatt	ttcttctactg	aaaagcctga	aaattctgcg	gatcagagga	1260
tatgtcttta	aagagttgaa	aagctttaac	ctctcgccat	tacataatct	tcaaaatctt	1320

	gaagttcttg	atcttggcac	taactttata	aaaattgcta	acctcagcat	gtttaaaaca	1380
	tttaaaagac	tgaaagtc	agatctttca	gtgaataaaa	tatcaccttc	aggagattca	1440
	agtgaagttg	gcttctgctc	aaatgccaga	acttctgtag	aaagttatga	accccaggtc	1500
	ctggaacaat	tacattattt	cagatatgat	aagtatgcaa	ggagttgcag	attcaaaaac	1560
5	aaagaggctt	ctttcatgtc	tgtaaatgaa	agctgtctaca	agtatgggca	gaccttggat	1620
	ctaagtaaaa	atagtatatt	ttttgtcaag	tcctctgatt	ttcagcatct	ttctttcctc	1680
	aaatgcctga	atctgtcagg	aaatctcatt	agccaaactc	ttaatggcag	tgaattccaa	1740
	ccttttagcag	agctgagata	tttggacttc	tccaacaacc	ggcttgattt	actccattca	1800
	acagcatttg	aagagcttca	caaactggaa	gttctggata	taagcagtaa	tagccattat	1860
10	tttcaatcag	aaggaattac	tcatatgcta	aactttacca	agaacctaaa	ggttctgcag	1920
	aaactgatga	tgaacgacaa	tgacatctct	tcctccacca	gcaggaccat	ggagagtgag	1980
	tctcttagaa	ctctggaatt	cagaggaaat	cacttagatg	ttttatggag	agaagggtgat	2040
	aacagatact	tacaattatt	caagaatctg	ctaaaattag	aggaattaga	catctctaaa	2100
	aattccctaa	gtttcttgcc	ttctggagtt	tttgatggta	tgcctccaaa	tctaaagaat	2160
15	ctctcttttg	ccaaaaatgg	gctcaaatct	ttcagttgga	agaaactcca	gtgtctaaag	2220
	aacctggaaa	ctttggacct	cagccacaac	caactgacca	ctgtccctga	gagattatcc	2280
	aactgttcca	gaagcctcaa	gaatctgatt	cttaagaata	atcaaatacag	gagtcctgacg	2340
	aagtattttc	tacaagatgc	cttccagttg	cgatatctgg	atctcagctc	aaataaaaatc	2400
	cagatgatcc	aaaagaccag	cttcccagaa	aatgtcctca	acaatctgaa	gatgttgctt	2460
20	ttgcatcata	atcggtttct	gtgcacctgt	gatgctgtgt	ggtttgtctg	gtgggttaac	2520
	catacggagg	tgactattcc	ttacctggcc	acagatgtga	cttgtgtggg	gccaggagca	2580
	cacaagggcc	aaagtgtgat	ctccctggat	ctgtacacct	gtgagttaga	tctgactaac	2640
	ctgattctgt	tctcactttc	catatctgta	tctctctttc	tcattggtgat	gatgacagca	2700
	agtcacctct	atttctggga	tgtgtgggtat	atttaccatt	tctgtaaggc	caagataaag	2760
25	gggtatcagc	gtctaataatc	accagactgt	tgctatgatg	cttttattgt	gtatgacact	2820
	aaagacccag	ctgtgaccga	gtgggttttg	gctgagctgg	tggccaaact	ggaagaccca	2880
	agagagaaac	attttaattt	atgtctcgag	gaaagggact	ggttaccagg	gcagccagtt	2940
	ctggaaaacc	tttcccagag	catacagctt	agcaaaaaga	cagtgtttgt	gatgacagac	3000
	aagtatgcaa	agactgaaaa	ttttaagata	gcattttact	tgtcccatca	gaggctcatg	3060
30	gatgaaaaag	ttgatgtgat	tatcttgata	tttcttgaga	agccttttca	gaagtccaag	3120
	ttcctccagc	tccggaaaag	gctctgtggg	agttctgtcc	ttgagtggcc	aacaaacccg	3180
	caagctcacc	catacttctg	gcagtgtcta	aagaacgccc	tggccacaga	caatcatgtg	3240
	gcctatagtc	aggtgttcaa	ggaaacgggc	tagaatcgaa	ttcccgcggc	cgccactgtg	3300
	ctggatatct	gcagaattcc	accacactgg	actagtggat	ccgagctcgg	taccaagctt	3360
35	aagtttaaac	cgc					3373

Table 7. Coding Region for Human TLR7 (5' to 3'; SEQ ID NO:169)

	atggtgtttc	caatgtggac	actgaagaga	caaattctta	tcctttttta	cataatccta	60
	atttccaaac	tccttggggc	tagatgggtt	cctaaaactc	tgccctgtga	tgctactctg	120
40	gatgttccaa	agaacctagt	gatcgtggac	tgcacagaca	agcatttgac	agaaattcct	180
	ggaggtattc	ccacgaacac	cacgaacctc	accctcacca	ttaaccacat	accagacatc	240

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gagaaacatt ttaatttatg tctcgaggaa agggactggt taccagggca gccagttctg 2820
gaaaaccttt cccagagcat acagcttagc aaaaagacag tgtttgat gatagacaag 2880
tatgcaaaga ctgaaaattt taagatagca ttttactgt cccatcagag gctcatggat 2940
gaaaaagttg atgtgattat cttgatattt cttgagaagc cttttcagaa gtccaagttc 3000
5 ctccagctcc ggaaaaggct ctgtgggagt tctgtccttg agtggccaac aaacccgcaa 3060
gctcaccat acttctggca gtgtctaaag aacgccctgg ccacagacaa tcatgtggcc 3120
tatagtcagg tgttcaagga aacggtc 3147

Table 8. Amino Acid Sequence of Human TLR7

		:	:	:	:	:	:	:	:	:	60
AF240467.pep	MVFPMTLKRQILILFNIILISKLLGARWFPKTLPCDVTLDVDPKNHVIVDCTDKHLTEIP										60
hTLR7.pep	MVFPMTLKRQILILFNIILISKLLGARWFPKTLPCDVTLDVDPKNHVIVDCTDKHLTEIP										60
AF245702.pep	MVFPMTLKRQILILFNIILISKLLGARWFPKTLPCDVTLDVDPKNHVIVDCTDKHLTEIP										60
		:	:	:	:	:	:	:	:	:	120
AF240467.pep	GGIPTNTNTNLTLTINHIPDISPASFHRLDHLVEIDFRCNCVPIPLGSKNNMCIKRLQIKP										120
hTLR7.pep	GGIPTNTNTNLTLTINHIPDISPASFHRLDHLVEIDFRCNCVPIPLGSKNNMCIKRLQIKP										120
AF245702.pep	GGIPTNTNTNLTLTINHIPDISPASFHRLDHLVEIDFRCNCVPIPLGSKNNMCIKRLQIKP										120
		:	:	:	:	:	:	:	:	:	180
AF240467.pep	RSFSGLTLYLKSLYLDGNQLLEIPQGLPPSLQLLSLEANNIFSIRKENLTELANIEILYLG										180
hTLR7.pep	RSFSGLTLYLKSLYLDGNQLLEIPQGLPPSLQLLSLEANNIFSIRKENLTELANIEILYLG										180
AF245702.pep	RSFSGLTLYLKSLYLDGNQLLEIPQGLPPSLQLLSLEANNIFSIRKENLTELANIEILYLG										180
		:	:	:	:	:	:	:	:	:	240
AF240467.pep	QNCYYRNPYVSYVSYIEKDAFLNLTCLKVLSLKDNNTAVPTVLPSTLTLEYLYNNMIAKI										240
hTLR7.pep	QNCYYRNPYVSYVSYIEKDAFLNLTCLKVLSLKDNNTAVPTVLPSTLTLEYLYNNMIAKI										240
AF245702.pep	QNCYYRNPYVSYVSYIEKDAFLNLTCLKVLSLKDNNTAVPTVLPSTLTLEYLYNNMIAKI										240
		:	:	:	:	:	:	:	:	:	300
AF240467.pep	QEDDFNNLNQLQILDLSGNCPRCYNAPFPCAPCKNNSPLQIPVNAFDALTELKVLRLHSN										300
hTLR7.pep	QEDDFNNLNQLQILDLSGNCPRCYNAPFPCAPCKNNSPLQIPVNAFDALTELKVLRLHSN										300
AF245702.pep	QEDDFNNLNQLQILDLSGNCPRCYNAPFPCAPCKNNSPLQIPVNAFDALTELKVLRLHSN										300
		:	:	:	:	:	:	:	:	:	360
AF240467.pep	SLQHVPPrWfKNINKLQELDLSQNFLAKEIGDAKFLHFLPSLIQLDLSFNfELQVYRASM										360
hTLR7.pep	SLQHVPPrWfKNINKLQELDLSQNFLAKEIGDAKFLHFLPSLIQLDLSFNfELQVYRASM										360
AF245702.pep	SLQHVPPrWfKNINKLQELDLSQNFLAKEIGDAKFLHFLPSLIQLDLSFNfELQVYRASM										360
		:	:	:	:	:	:	:	:	:	420
AF240467.pep	NLSQAFSSLSKSLKILRIRGYVfKELKSFNLSPLHNLQNLEVLDLGTNfIKIANLSMfKQF										420
hTLR7.pep	NLSQAFSSLSKSLKILRIRGYVfKELKSFNLSPLHNLQNLEVLDLGTNfIKIANLSMfKQF										420
AF245702.pep	NLSQAFSSLSKSLKILRIRGYVfKELKSFNLSPLHNLQNLEVLDLGTNfIKIANLSMfKQF										420
		:	:	:	:	:	:	:	:	:	480

AF240467.pep KRLKVIDLSV NKISPSGDSSEVGFC SNARTSVESYEPQVLEQLHYFRYDKYARSCRFKNK 480
hTLR7.pep KRLKVIDLSV NKISPSGDSSEVGFC SNARTSVESYEPQVLEQLHYFRYDKYARSCRFKNK 480
AF245702.pep KRLKVIDLSV NKISPSGDSSEVGFC SNARTSVESYEPQVLEQLHYFRYDKYARSCRFKNK 480

5 . : . : . : . : . : . : 540
AF240467.pep EASFMSVNESCYKYGQTLDSLKNSIFFVKSSDFQHLSFLKCLNLSGNLISQTLNGSEFQP 540
hTLR7.pep EASFMSVNESCYKYGQTLDSLKNSIFFVKSSDFQHLSFLKCLNLSGNLISQTLNGSEFQP 540
AF245702.pep EASFMSVNESCYKYGQTLDSLKNSIFFVKSSDFQHLSFLKCLNLSGNLISQTLNGSEFQP 540

10 . : . : . : . : . : . : 600
AF240467.pep LAELRYLDFSNNRLDLLHSTAFEELHKLEVLDISSNSHYFQSEGITHMLNFTKNLKV LQK 600
hTLR7.pep LAELRYLDFSNNRLDLLHSTAFEELHKLEVLDISSNSHYFQSEGITHMLNFTKNLKV LQK 600
AF245702.pep LAELRYLDFSNNRLDLLHSTAFEELHKLEVLDISSNSHYFQSEGITHMLNFTKNLKV LQK 600

15 . : . : . : . : . : . : 660
AF240467.pep LMMNDNDISSSTSRTMESESLRTEFRGNHLDVLWREGDNRYLQLFKNLLKLEELDISKN 660
hTLR7.pep LMMNDNDISSSTSRTMESESLRTEFRGNHLDVLWREGDNRYLQLFKNLLKLEELDISKN 660
AF245702.pep LMMNDNDISSSTSRTMESESLRTEFRGNHLDVLWREGDNRYLQLFKNLLKLEELDISKN 660

20 . : . : . : . : . : . : 720
AF240467.pep SLSFLPSGVFDGMPPNLKNLSLAKNGLKSF SWKKLQCLKNLETDL SHNQLTTVPERLSN 720
hTLR7.pep SLSFLPSGVFDGMPPNLKNLSLAKNGLKSF SWKKLQCLKNLETDL SHNQLTTVPERLSN 720
AF245702.pep SLSFLPSGVFDGMPPNLKNLSLAKNGLKSF SWKKLQCLKNLETDL SHNQLTTVPERLSN 720

25 . : . : . : . : . : . : 780
AF240467.pep CSRS LKNLILKNNQIRSLTKYFLQDAFQLRYLDLSSNKIQMIQKTSFPENVLNNLKMLLL 780
hTLR7.pep CSRS LKNLILKNNQIRSLTKYFLQDAFQLRYLDLSSNKIQMIQKTSFPENVLNNLKMLLL 780
AF245702.pep CSRS HKNLILKNNQIRSPTKYFLQDAFQLRYLDLSSNKIQMIQKTSFPENVLNNLKMLLL 780

30 . : . : . : . : . : . : 840
AF240467.pep HHNRFLCTCDAVWFVWVNHTTEVTIPYLATDVTGPGAHKGQSVISLDLYTCELDLTNL 840
hTLR7.pep HHNRFLCTCDAVWFVWVNHTTEVTIPYLATDVTGPGAHKGQSVISLDLYTCELDLTNL 840
AF245702.pep HHNRFLCTCDAVWFVWVNHTTEVTIPYLATDVTGPGAHKGQSVISLDLYTCELDLTNL 840

35 . : . : . : . : . : . : 900
AF240467.pep ILFSLSISVSLFLMVMMTASHLYFWDVWYIYHFCKAKIKGYQRLISPDCCYDAFIVYDTK 900
hTLR7.pep ILFSLSISVSLFLMVMMTASHLYFWDVWYIYHFCKAKIKGYQRLISPDCCYDAFIVYDTK 900
AF245702.pep ILFSLSISVSLFLMVMMTASHLYFWDVWYIYHFCKAKIKGYQRLISPDCCYDAFIVYDTK 900

40 . : . : . : . : . : . : 960
AF240467.pep DPAVTEWVLAELVAKLEDPREKHFNLCLEERDWLPGQPVLNLSQSIQLSKKTVFVMTDK 960
hTLR7.pep DPAVTEWVLAELVAKLEDPREKHFNLCLEERDWLPGQPVLNLSQSIQLSKKTVFVMTDK 960
AF245702.pep DPAVTEWVLAELVAKLEDPREKHFNLCLEERDWLPGQPVLNLSQSIQLSKKTVFVMTDK 960

45 . : . : . : . : . : . : 1020
AF240467.pep YAKTENFKIAFYLSHQRLMDEKVDVIIIFLEKPFQKSKFLQLRKRLCGSSVLEWPTNPQ 1020

hTLR7 . pep	YAKTENFKIAFYLSHQRLMDEKVDVILIFLEKPFQKSKFLQLRKRLCGSSVLEWPTNPQ	1020
AF245702 . pep	YAKTENFKIAFYLSHQRLMDEKVDVILIFLEKPFQKSKFLQLRKRLCGSSVLEWPTNPQ	1020
	:	1080
5 AF240467 . pep	AHPYFWQCLKNALATDNHVAYSQVFKETV	1049
hTLR7 . pep	AHPYFWQCLKNALATDNHVAYSQVFKETV	1049
AF245702 . pep	AHPYFWQCLKNALATDNHVAYSQVFKETV	1049

In Table 8 the sequences are assigned as follows: hTLR7.pep, SEQ ID NO:170;

10 AF240467.pep, SEQ ID NO:171; AF245702.pep, SEQ ID NO:172.

Example 17. Method of cloning the murine TLR7

Alignment of human TLR7 protein sequence with mouse EST database using tfasta yielded 4 hits with mouse EST sequences bb116163, aa266744, bb210780 and aa276879.

Two primers were designed that bind to aa266744 sequence for use in a RACE-PCR to amplify 5' and 3' ends of the murine TLR7 cDNA. The library used for the RACE PCR was a mouse spleen marathon-ready cDNA commercially available from Clontech. A 5' fragment with a length of 3000 bp obtained by this method was cloned into Promega pGEM-T Easy vector. After sequencing of the 5' end, additional primers were designed for amplification of the complete murine TLR7 cDNA. The primer for the 5' end was obtained from the sequence of the 5' RACE product whereas the primer for the 3' end was selected from the mouse EST sequence aa266744.

Three independent PCR reactions were set up using a murine macrophage RAW264.7 (ATCC TIB-71) cDNA as a template with the primers 5'-

25 CTCCTCCACCAGACCTCTTGATTCC-3' (SEQ ID NO:208) and 5'-

CAAGGCATGTCCTAGGTGGTGACATTC-3' (SEQ ID NO:209). The resulting

amplification products were cloned into pGEM-T Easy vector and fully sequenced (SEQ ID NO:173; Table 9). The open reading frame of mTLR7 (SEQ ID NO:174; Table 10) starts at base 49, ends at base 3201 and codes for a protein of 1050 amino acids (SEQ ID NO:175;

30 Table11). To create an expression vector for murine TLR7 cDNA, pGEM-T Easy vector plus mTLR7 insert was cut with NotI, the fragment isolated and ligated into a NotI digested pCDNA3.1 expression vector (Invitrogen).

Table 9. cDNA Sequence for Murine TLR7 (5' to 3'; SEQ ID NO:173)

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25

	AGTGTCATAT	CCCTTGATCT	GTATACGTGT	GAGTTAGATC	TCACAAACCT	GATTCTGTTC	2580
	TCAGTTTCCA	TATCATCAGT	CCTCTTTCTT	ATGGTAGTTA	TGACAACAAG	TCACCTCTTT	2640
	TTCTGGGATA	TGTGGTACAT	TTATTATTTT	TGGAAAAGCAA	AGATAAAGGG	GTATCAGCAT	2700
	CTGCAATCCA	TGGAGTCTTG	TTATGATGCT	TTTATTGTGT	ATGACACTAA	AAACTCAGCT	2760
5	GTGACAGAAT	GGGTTTTGCA	GGAGCTGGTG	GCAAAAATTGG	AAGATCCAAG	AGAAAAACAC	2820
	TTCAATTTGT	GTCTAGAAGA	AAGAGACTGG	CTACCAGGAC	AGCCAGTTCT	AGAAAACCTT	2880
	TCCCAGAGCA	TACAGCTCAG	CAAAAAGACA	GTGTTTGTGA	TGACACAGAA	ATATGCTAAG	2940
	ACTGAGAGTT	TTAAGATGGC	ATTTTATTTG	TCTCATCAGA	GGCTCCTGGA	TGAAAAAGTG	3000
	GATGTGATTA	TCTTGATATT	CTTGGAAGAG	CCTCTTCAGA	AGTCTAAGTT	TCTTCAGCTC	3060
10	AGGAAGAGAC	TCTGCAGGAG	CTCTGTCCTT	GAGTGGCCTG	CAAATCCACA	GGCTCACCCA	3120
	TACTTCTGGC	AGTGCCTGAA	AAATGCCCTG	ACCACAGACA	ATCATGTGGC	TTATAGTCAA	3180
	ATGTTCAAGG	AAACAGTCTA	GCTCTCTGAA	GAATGTCACC	ACCTAGGACA	TGCCTTGAAT	3240
	CGA						3243

Table 10. Coding Region for Murine TLR7 (5' to 3'; SEQ ID NO:174)

	ATGGTGTTTT	CGATGTGGAC	ACGGAAGAGA	CAAATTTTGA	TCTTTTTTAA	TATGCTCTTA	60
	GTTTCTAGAG	TCTTTGGGTT	TCGATGGTTT	CCTAAAACTC	TACCTTGTGA	AGTTAAAGTA	120
	AATATCCCAG	AGGCCCATGT	GATCGTGGAC	TGCACAGACA	AGCATTTGAC	AGAAATCCCT	180
	GAGGGCATT	CCACTAACAC	CACCAATCTT	ACCCTTACCA	TCAACCACAT	ACCAAGCATC	240
20	TCTCCAGATT	CCTTCCGTAG	GCTGAACCAT	CTGGAAGAAA	TCGATTTAAG	ATGCAATTGT	300
	GTACCTGTTC	TACTGGGGTC	CAAAGCCAAT	GTGTGTACCA	AGAGGCTGCA	GATTAGACCT	360
	GGAAGCTTTA	GTGGACTCTC	TGACTTAAAA	GCCCTTTACC	TGGATGGAAA	CCAACCTCTG	420
	GAGATACCAC	AGGATCTGCC	ATCCAGCTTA	CATCTTCTGA	GCCTTGAGGC	TAACAACATC	480
	TTCTCCATCA	CGAAGGAGAA	TCTAACAGAA	CTGGTCAACA	TTGAAACACT	CTACCTGGGT	540
25	CAAACTGT	ATTATCGAAA	TCCTTGCAAT	GTTTCCTATT	CTATTGAAAA	AGATGCTTTC	600
	CTAGTTATGA	GAAATTTGAA	GGTTCTCTCA	CTAAAAGATA	ACAATGTCAC	AGCTGTCCCC	660
	ACCACTTTGC	CACCTAATTT	ACTAGAGCTC	TATCTTTATA	ACAATATCAT	TAAGAAAATC	720
	CAAGAAAATG	ATTTTAATAA	CCTCAATGAG	TTGCAAGTTC	TTGACCTAAG	TGGAAATTGC	780
	CCTCGATGTT	ATAATGTCCC	ATATCCGTGT	ACACCGTGTG	AAAATAATTC	CCCCTTACAG	840
30	ATCCATGACA	ATGCTTTCAA	TTCATTGACA	GAATTAAAAAG	TTTTACGTTT	ACACAGTAAT	900
	TCTCTTCAGC	ATGTGCCCCC	AACATGGTTT	AAAAACATGA	GAAACCTCCA	GGAACCTAGAC	960
	CTCTCCCAA	ACTACTTGGC	CAGAGAAATT	GAGGAGGCCA	AATTTTTGCA	TTTTCTTCCC	1020
	AACCTTGTTG	AGTTGGATTT	TTCTTTCAAT	TATGAGCTGC	AGGTCTACCA	TGCATCTATA	1080
	ACTTTACCAC	ATTCACCTC	TTCATTGGAA	AACTTGAAAA	TTCTGCGTGT	CAAGGGGTAT	1140
35	GTCTTTAAAG	AGCTGAAAAA	CTCCAGTCTT	TCTGTATTGC	ACAAGCTTCC	CAGGCTGGAA	1200
	GTTCTTGACC	TTGGCACTAA	CTTCATAAAA	ATTGCTGACC	TCAACATATT	CAACATTTT	1260
	GAAAACCTCA	AACTCATAGA	CCTTTTCAGTG	AATAAGATAT	CTCCTTCAGA	AGAGTCAAGA	1320
	GAAGTTGGCT	TTTGTCTTAA	TGCTCAAAC	TCTGTAGACC	GTCATGGGCC	CCAGGTCCTT	1380
	GAGGCCTTAC	ACTATTTCCG	ATACGATGAA	TATGCACGGA	GCTGCAGGTT	CAAAAACAAA	1440
40	GAGCCACCTT	CTTCTCTGCC	TTTGAATGCA	GACTGCCACA	TATATGGGCA	GACCTTAGAC	1500
	TTAAGTAGAA	ATAACATATT	TTTTATTAAA	CCTTCTGATT	TTCAGCATCT	TTCATTCTCT	1560

	mTLR7 . pep	QNCYYRNPNCVSYSEKDAFLVMRNLKVLSLKDNNTAVPTTLPNLELYLYNNIIKKI	240
		. : . : . : . : . : . : . : . : . :	300
	hTLR7 . pep	QEDDFNNLNQLQILDLSGNCPRCYNAPFPCAPCKNNSPLQIPVNAFDALTELKVLRLHSN	300
5	mTLR7 . pep	QENDFNNLNELQVLDLSGNCPRCYNVPYPCTPCENNSPLQIHDNAFNSLTELVRLHSN	300
		. : . : . : . : . : . : . : . : . :	360
	hTLR7 . pep	SLQHVPPRWFKNINKLQELDLSQNFLAKEIGDAKFLHFLPSLIQLDLSFNFELQVYRASM	360
10	mTLR7 . pep	SLQHVPPTWFKNMNRNLQELDLSQNYLAREIEEAKFLHFLPNLVELDFSFNVELQVYHASI	360
		. : . : . : . : . : . : . : . : . :	420
	hTLR7 . pep	NLSQAFSSLKSLKILRIRGYVFKELKSFNLSPLHNLQNLEVLDLGTNFIKIANLSMFKQF	420
	mTLR7 . pep	TLPHSLSSLENLKILRVKGYVFKELKNSSSVLHKLPRLEVLDLGTNFIKIADLNIFKHF	420
		. : . : . : . : . : . : . : . : . :	480
15	hTLR7 . pep	KRLKVIDLSVNKISPSPGDSSEVGFCNARTSVESYEPQVLEQLHYFRYDKYARSCRFKNK	480
	mTLR7 . pep	ENLKLIDLSVNKISPSEESREVGFCNQAQTSVDRHGPVLEALHYFRYDEYARSCRFKNK	480
		. : . : . : . : . : . : . : . : . :	540
20	hTLR7 . pep	EA-SFMSVNESCYKYGTLDLSKNSIFFVKSSDFQHLSFLKCLNLSGNLISQTLNGSEFQ	539
	mTLR7 . pep	EPPSFLPLNADCHIYGQTLDLRSNNIFFIKPSDFQHLSFLKCLNLSGNTIGQTLNGSELW	540
		. : . : . : . : . : . : . : . : . :	600
	hTLR7 . pep	PLAELRYLDFSNNRLDLLHSTAFEELHKLEVLDISSNSHYFQSEGITHMLNFTKNLKVLO	599
25	mTLR7 . pep	PLRELRYLDFSNNRLDLLYSTAFEELQSLEVLDLSSNSHYFQAEGITHMLNFTKKLRLLD	600
		. : . : . : . : . : . : . : . : . :	660
	hTLR7 . pep	KLMMNDNDISSSTSRMESLSRLTLEFRGNHLDVLWREGDNRYLQLFKNLLKLEELDISK	659
30	mTLR7 . pep	KLMMNDNDISTSASRTMESDSLRILEFRGNHLDVLWRAGDNRYLDFFKNLFNLEVLDISR	660
		. : . : . : . : . : . : . : . : . :	720
	hTLR7 . pep	NSLSFLPSGVFDGMPPNLKNLSLAKNGLKSFSWKKLQCLKNLETDLDSHNQLTTVPERLS	719
	mTLR7 . pep	NSLNSLPPEVFEGMPPNLKNLSLAKNGLKSFFWDRLQLLKHLEILDLSHNQLTKVPERLA	720
		. : . : . : . : . : . : . : . : . :	780
35	hTLR7 . pep	NCSRSLKNLILKNNQIRSLTKYFLQDAFQLRYLDLSSNKIQMIQKTSFPENVLNNLKMLL	779
	mTLR7 . pep	NCSKSLTTLILKHNQIRQLTKYFLEDALQLRYLDISSNKIQVIQKTSFPENVLNNLEMLV	780
		. : . : . : . : . : . : . : . : . :	840
40	hTLR7 . pep	LHHNRFLCTDAVWFVWVWNHTEVTIPYLATDVTCVGPAGAHKGQSVISLDLYTCELDLTN	839
	mTLR7 . pep	LHHNRFLCNCDAVWFVWVWNHTEVTIPYLATDVTCVGPAGAHKGQSVISLDLYTCELDLTN	840
		. : . : . : . : . : . : . : . : . :	900
	hTLR7 . pep	LILFSLSISVSLFLMVMMTASHLYFWDVWYIYHFCKAKIKGYQRLISPDCCYDAFIVYDT	899
45	mTLR7 . pep	LILFVSISSVLFLMVMMTSHLFFWDMWYIYFWKAKIKGYQHLQSMESCYDAFIVYDT	900

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The protein sequence of the cloned hTLR8 cDNA matches the sequence described

under the GenBank accession number AF245703. The sequence deposited under GenBank accession number AF246971 contains an insertion at the N-terminus of 15 amino acids (MKESSLQNSSCSLGGKETKK; SEQ ID NO:185) and three single amino acid changes at positions 217 (P to S), 266 (L to P) and 867 (V to I).

5

Table 12. cDNA Sequence for Human TLR8 (5' to 3'; SEQ ID NO:182)

	gctccccggcc gccatggcgg ccgcgggaat tcgattctgc gctgctgcaa gttacggaat 60
	gaaaaattag aacaacagaa acatggaaaa catgttcctt cagtcgtcaa tgctgacctg 120
	cattttcctg ctaatatctg gttcctgtga gttatgcgcc gaagaaaatt tttctagaag 180
10	ctatccttgt gatgagaaaa agcaaaatga ctcagttatt gcagagtgca gcaatcgctg 240
	actacaggaa gttccccaaa cgggtgggcaa atatgtgaca gaactagacc tgtctgataa 300
	tttcatcaca cacataacga atgaatcatt tcaagggctg caaaatctca ctaaaataaa 360
	tctaaaccac aaccccaatg tacagcacca gaacggaaat cccggtatac aatcaaatgg 420
	cttgaatatc acagacgggg cattcctcaa cctaaaaaac ctaaggaggt tactgcttga 480
15	agacaaccag ttaccccaaa taccctctgg tttgccagag tctttgacag aacttagtct 540
	aattcaaaac aatatataca acataactaa agagggcatt tcaagactta taaacttgaa 600
	aaatctctat ttggcctgga actgctatatt taacaaagtt tgcgagaaaa ctaacataga 660
	agatggagta tttgaaacgc tgacaaatth ggagttgcta tcactatctt tcaattctct 720
	ttcacacgtg ccacccaaac tgccaagctc cctacgcaaa ctttttctga gcaacaccca 780
20	gatcaaatac attagtgaag aagatttcaa gggattgata aatttaacat tactagattt 840
	aagcgggaac tgtccgaggt gcttcaatgc cccatttcca tgcgtgcctt gtgatggtgg 900
	tgcttcaatt aatatagatc gttttgcttt tcaaaacttg acccaacttc gatacctaaa 960
	cctctctagc acttccctca ggaagattaa tgctgcctgg tttaaaaata tgctctatct 1020
	gaagggtgctg gatcttgaat tcaactatth agtgggagaa atagcctctg gggcattttt 1080
25	aacgatgctg ccccgcttag aaatacttga cttgtctttt aactatataa aggggagtta 1140
	tccacagcat attaatatth ccagaaactt ctctaaactt ttgtctctac gggcattgca 1200
	tttaagaggt tatgtgttcc aggaactcag agaagatgat ttccagcccc tgatgcagct 1260
	tccaaactta tcgactatca acttggtgat taattttatt aagcaaatcg atttcaaact 1320
	tttccaaaat ttctccaatc tggaaattat ttacttgctc gaaaacagaa tatcacctgt 1380
30	ggtaaaagat acccggcaga gttatgcaaa tagttcctct tttcaacgtc atatccggaa 1440
	acgacgtcga acagatthttg agtttgaccc acattcgaac ttttatcatt tcacccgtcc 1500
	tttaataaaag ccacaatgtg ctgcttatgg aaaagcctta gatttaagcc tcaacagtat 1560
	tttcttcatt gggccaaacc aatttgaaaa tcttctcgac attgcctgtt taaatctgtc 1620
	tgcaaatagc aatgctcaag tgttaagtgg aactgaatth tcagccattc ctcatgtcaa 1680
35	atatttggtat ttgacaaaca atagactaga ctttgataat gctagtgtct ttactgaatt 1740
	gtccgacttg gaagttctag atctcagcta taattcacac tatttcagaa tagcaggcgt 1800
	aacacatcat ctagaattta ttcaaaatth cacaaatcta aaagtthttaa acttgagcca 1860
	caacaacatt tatactthtaa cagataagta taacctggaa agcaagtccc tggtagaatt 1920
	agttttcagt ggcaatcgcc ttgacattht gtggaatgat gatgacaaca ggtatatctc 1980
40	cattttcaaa ggtctcaaga atctgacacg tctggattta tcccttaata ggctgaagca 2040

	catcccaaat	gaagcattcc	ttaatttgcc	agcgagtctc	actgaactac	atataaatga	2100
	taatattgta	aagtttttta	actggacatt	actccagcag	tttcctcgtc	tcgagttgct	2160
	tgacttacgt	ggaaacaaac	tactcttttt	aactgatagc	ctatctgact	ttacatcttc	2220
	ccttcggaca	ctgctgctga	gtcataacag	gatttcccac	ctaccctctg	gctttctttc	2280
5	tgaagtcagt	agtctgaagc	acctcgattt	aagttccaat	ctgctaaaaa	caatcaacaa	2340
	atccgcactt	gaaactaaga	ccaccaccaa	attatctatg	ttggaactac	acggaaaccc	2400
	ctttgaatgc	acctgtgaca	ttggagattt	ccgaagatgg	atggatgaac	atctgaatgt	2460
	caaaattccc	agactggtag	atgtcatttg	tgccagtcct	ggggatcaaa	gaggggaagag	2520
	tattgtgagt	ctggagctaa	caacttggtg	ttcagatgtc	actgcagtga	tattatTTTT	2580
10	cttcacgttc	tttatcacca	ccatggttat	gttggctgcc	ctggctcacc	atttgTTTTa	2640
	ctgggatgtt	tggttttatat	ataatgtgtg	tttagctaag	gtaaaaggct	acaggctctc	2700
	ttccacatcc	caaactttct	atgatgctta	catttcttat	gacaccaaag	acgcctctgt	2760
	tactgactgg	gtgataaatg	agctgcgcta	ccaccttgaa	gagagccgag	acaaaaacgt	2820
	tctcctttgt	ctagaggaga	gggattggga	cccgggattg	gccatcatcg	acaacctcat	2880
15	gcagagcatc	aaccaaagca	agaaaacagt	atttgTTTTa	acaaaaaat	atgcaaaaag	2940
	ctggaacttt	aaaacagctt	tttacttggc	tttgacagag	ctaattggatg	agaacatgga	3000
	tgtgattata	tttatcctgc	tggagccagt	gttacagcat	tctcagtatt	tgaggctacg	3060
	gcagcggatc	tgtaaagagct	ccatcctcca	gtggcctgac	aaccggaagg	cagaaggctt	3120
	gttttgga	actctgagaa	atgtggtctt	gactgaaaat	gattcacggt	ataacaatat	3180
20	gtatgtcgat	tccattaagc	aataactaact	gacgttaagt	catgatttcg	cgcaatcact	3240
	agtgaattcg	cggccgctg	caggctgacc	atatgggaga	gctcccaacg	cgttggatgc	3300
	atagcttgag						3310

Table 13. Coding Region for Human TLR8 (5' to 3'; SEQ ID NO:183)

25	atggaaaaca	tggttccttca	gtcgtcaatg	ctgacctgca	tttcctcgct	aatatctggt	60
	tcctgtgagt	tatgcgccga	agaaaatttt	tctagaagct	atccttgtga	tgagaaaaag	120
	caaaatgact	cagttattgc	agagtgcagc	aatcgctgac	tacaggaagt	ccccaaaacg	180
	gtgggcaaat	atgtgacaga	actagacctg	tctgataatt	tcatcacaca	cataacgaat	240
	gaatcatttc	aagggtgca	aaatctcact	aaaataaatc	taaaccacaa	ccccaatgta	300
30	cagcaccaga	acggaaatcc	cggatatacaa	tcaaattggct	tgaatatcac	agacggggca	360
	ttcctcaacc	taaaaaacct	aagggtggtta	ctgcttgaag	acaaccagtt	accccaaata	420
	ccctctgggt	tgccagagtc	tttgacagaa	cttagtctaa	ttcaaaacaa	tatatataac	480
	ataactaaag	agggcatttc	aagacttata	aacttgaaaa	atctctattt	ggcctggaac	540
	tgctatttta	acaaagtgtg	cgagaaaact	aacatagaag	atggagtatt	tgaaacgctg	600
35	acaaatttgg	agttgtatc	actatctttc	aattctcttt	cacacgtgcc	acccaaactg	660
	ccaagctccc	tacgcaaact	ttttctgagc	aacaccgaga	tcaaatacat	tagtgaagaa	720
	gatttcaagg	gattgataaa	tttaacatta	ctagatttaa	gcgggaactg	tccgaggtgc	780
	ttcaatgccc	catttccatg	cgtgccttgt	gatggtggtg	cttcaattaa	tatagatcgt	840
	tttgcttttc	aaaacttgac	ccaacttcga	tacctaaacc	tctctagcac	ttccctcagg	900
40	aagattaatg	ctgcctgggt	taaaaatatg	cctcatctga	aggtgctgga	tcttgaattc	960
	aactatttag	tgggagaaat	agcctctggg	gcatttttaa	cgatgctgcc	ccgcttagaa	1020

[illegible]

15 20 25

Example 19. Method of cloning the murine TLR8

Alignment of human TLR8 protein sequence with mouse EST database using tfasta yielded 1 hit with mouse EST sequence bf135656. Two primers were designed that bind to bf135656 sequence for use in a RACE-PCR to amplify 5' and 3' ends of the murine TLR8 cDNA. The library used for the RACE PCR was a mouse spleen marathon-ready cDNA commercially available from Clontech. A 5' fragment with a length of 2900 bp and a 3' fragment with a length of 2900 bp obtained by this method were cloned into Promega pGEM-T Easy vector. After sequencing of the 5' end and 3' end of each fragment, partial sequences of mTLR8 were obtained and allowed the design of primers for amplification of the complete murine TLR8 cDNA.

Three independent PCR reactions were set up using a spleen murine cDNA from Clontech as a template with the primers 5'-GAGAGAAACAAACGTTTTACCTTC-3' (SEQ ID NO:188) and 5'-GATGGCAGAGTCGTGACTTCCC-3' (SEQ ID NO:189). The resulting amplification products were cloned into pGEM-T Easy vector, fully sequenced, translated into protein, and aligned to the human TLR8 protein sequence (GenBank accession number AF245703). The cDNA sequence for mTLR8 is SEQ ID NO:190, presented in Table 15. The open reading frame of mTLR8 starts at base 59, ends at base 3157, and codes for a protein of 1032 amino acids. SEQ ID NO:191 (Table 16), corresponding to bases 59-3154 of SEQ ID NO:190 (Table 15), is the coding region for the polypeptide of SEQ ID NO:192 (Table 17). To create an expression vector for murine TLR8, cDNA pGEM-T Easy vector with the mTLR8 insert was cut with NotI, the fragment isolated, and ligated into a NotI-digested pCDNA3.1 expression vector (Invitrogen).

Table 15. cDNA Sequence for Murine TLR8 (5' to 3'; SEQ ID NO:190)

attcagagtt	ggatgttaag	agagaaacaa	acgttttacc	ttcctttgtc	tatagaacat	60
ggaaaacatg	ccccctcagt	catggattct	gacgtgcttt	tgtctgctgt	cctctggaac	120
cagtgccatc	ttccataaag	cgaactattc	cagaagctat	ccttgtgacg	agataaggca	180
caactccctt	gtgattgcag	aatgcaacca	tcgtcaactg	catgaagtgc	cccaaactat	240
aggcaagtat	gtgacaaaca	tagacttgct	agacaatgcc	attacacata	taacgaaaga	300
gtcctttcaa	aagctgcaaa	acctcactaa	aatcgatctg	aaccacaatg	ccaaacaaca	360
gcacccaaat	gaaaataaaa	atggtatgaa	tattacagaa	ggggcacttc	tcagcctaag	420
aaatctaaca	gttttactgc	tggaagacaa	ccagttatat	actataacctg	ctgggttgcc	480
tgagtctttg	aaagaactta	gcctaattca	aaacaatata	tttcaggtaa	ctaaaaacaa	540
cacttttggg	cttaggaact	tggaaagact	ctatttgggc	tggaaactgct	attttaaatg	600
taatcaaacc	tttaaggtag	aagatggggc	atttaaaaat	cttatacact	tgaaggtact	660

40

atcataaaaa cacacagctt ctccttacia tgaaccgaat

3220

Table 16. Coding Region for Murine TLR8 (5' to 3'; SEQ ID NO:191)

	atggaaaaca	tgccccctca	gtcatggatt	ctgacgtgct	tttgtctgct	gtcctctgga	60
5	accagtgcc	tcttccataa	agcgaactat	tccagaagct	atccttgtga	cgagataagg	120
	cacaactccc	ttgtgattgc	agaatgcaac	catcgtcaac	tgcatgaagt	tccccaaact	180
	ataggcaagt	atgtgacaaa	catagacttg	tcagacaatg	ccattacaca	tataacgaaa	240
	gagtcctttc	aaaagctgca	aaacctcact	aaaatcgatc	tgaaccacaa	tgccaaacaa	300
	cagcacccaa	atgaaaataa	aaatgggtatg	aatattacag	aaggggcact	tctcagccta	360
10	agaaatctaa	cagttttact	gctggaagac	aaccagttat	atactatacc	tgctggggtg	420
	cctgagtctt	tgaagaact	tagcctaatt	caaaacaata	tatttcaggt	aactaaaaac	480
	aacacttttg	ggcttaggaa	cttggaagaa	ctctatttgg	gctggaactg	ctattttaaa	540
	tgtaatcaaa	cctttaaggt	agaagatggg	gcatttaaaa	atcttatata	cttgaaggta	600
	ctctcattat	ctttcaataa	ccttttctat	gtgcccccca	aactaccaag	ttctctaagg	660
15	aaactttttc	tgagtaatgc	caaaatcatg	aacatcactc	aggaagactt	caaaggactg	720
	gaaaatttaa	cattactaga	tctgagtggg	aactgtccaa	gggtgttaca	tgctccattt	780
	ccttgacac	cttgcaagga	aaactcatcc	atccacatac	atcctctggc	ttttcaaagt	840
	ctcaccaca	ttctctatct	aaacctttcc	agcacttccc	tcaggacgat	tccttctacc	900
	tggtttgaaa	atctgtcaaa	tctgaaggaa	ctccatcttg	aattcaacta	tttagttcaa	960
20	gaaattgcct	cgggggcatt	tttaacaaaa	ctaccagtt	tacaaatcct	tgatttgtcc	1020
	ttcaactttc	aatataagga	atatttacia	tttattaata	tttctcaaa	tttctctaag	1080
	cttcgttctc	tcaagaagtt	gcacttaaga	ggctatgtgt	tccgagaact	taaaaagaag	1140
	catttcgagc	atctccagag	tcttccaaac	ttggcaacca	tcaacttggg	cattaacttt	1200
	attgagaaaa	ttgatttcaa	agctttccag	aatttttcca	aactcgacgt	tatctattta	1260
25	tcaggaaatc	gcatagcatc	tgtattagat	ggtacagatt	attcctcttg	gcgaaatcgt	1320
	cttcggaaac	ctctctcaac	agacgatgat	gagtttgatc	cacacgtgaa	tttttaccat	1380
	agcaccaaac	ctttaataaa	gccacagtgt	actgcttatg	gcaaggcctt	ggattttaagt	1440
	ttgaacaata	ttttcattat	tgggaaaagc	caatttgaag	gttttcagga	tatcgcttgc	1500
	ttaaatctgt	ccttcaatgc	caatactcaa	gtgtttaatg	gcacagaatt	ctcctccatg	1560
30	ccccacatta	aatattttgga	tttaaccaac	aacagactag	actttgatga	taacaatgct	1620
	ttcagtgatc	ttcacgatct	agaagtgtctg	gacctgagcc	acaatgcaca	ctatttcagt	1680
	atagcagggg	taacgcaccg	tctaggattt	atccagaact	taataaacct	caggggtgta	1740
	aacctgagcc	acaatggcat	ttacaccctc	acagaggaaa	gtgagctgaa	aagcatctca	1800
	ctgaaagaat	tggtttttcag	tggaaatcgt	cttgaccatt	tgtggaatgc	aaatgatggc	1860
35	aaatactggg	ccatttttaa	aagtctccag	aatttgatac	gcctggactt	atcatacaat	1920
	aaccttcaac	aaatcccaaa	tggagcattc	ctcaatttgc	ctcagagcct	ccaagagtta	1980
	cttatcagtg	gtaacaaatt	acgtttcttt	aattggacat	tactccagta	ttttcctcac	2040
	cttcacttgc	tggattttatc	gagaaatgag	ctgtattttc	tacccaattg	cctatctaag	2100
	tttgacacatt	ccctggagac	actgctactg	agccataatc	atttctctca	cctaccctct	2160
40	ggcttctctc	ccgaagccag	gaatctgggtg	cacctggatc	taagtttcaa	cacaataaag	2220
	atgatcaata	aatcctccct	gcaaaccaag	atgaaaacga	acttgtctat	tctggagcta	2280

5

10

	. : . : . : . : . :	60
mTLR8.pep	MENMPPQSWILTCFCLLSSGTSAl FHKANYSRSYPCDEIRHNSLVIAECNHRQLHEVPQT	60
hTLR8.pep	MENMFLQSSMLTCl FLLISGSCELCAEENFSRSYPCDEKKQNDSVlAECSNRRLQEVPQT	60
	. : . : . : . : . : . :	120
mTLR8.pep	IGKYVTNIIDLSDNAITHITKESfQKLQNLTKIDLNHNAKQQH----PNENKNGMNITEGA	116
hTLR8.pep	VgKYVTELDLSDNFiTHITNESfQGLQNLTKINLNHNPNVQHqONGNPgiQSnglnITDga	120
	. : . : . : . : . : . :	180
mTLR8.pep	LLSLRNLTvLLLEDnQLyTiPaGlPEslKElsLIQNNIfQvTKnnTFglRNlerLYLGwN	176
hTLR8.pep	fLnLknlREllLEdNQlPQiPSglPEslTElsLIQNNIyniTkeGISrLINlkNlyLaWN	180
	. : . : . : . : . : . :	240
mTLR8.pep	CYFK--CNQTFKVEdGaFKNlIHlKVLSlSFnnLFyVppKLPssLRklfLSnaKiMniTQ	234
hTLR8.pep	CyFNkVCEKT-NIEdgVFETlTNlellSLSfNsLSHVppKLPssLRklfLSntQIKYISE	239
	. : . : . : . : . : . :	300
mTLR8.pep	EDfKGlenLTllDLsgncPRcYNAPFPCTPCKENSSIHIHPLAFQSLtQLLYLNLSSTSL	294
hTLR8.pep	EDfKGlinLTllDLsgncPRCFNAPFCVPCDGgASinIdRFafQNLtQLRYLNLSSTSL	299
	. : . : . : . : . : . :	360
mTLR8.pep	RtiPstWfEnLSnlKeLHLefNyLvqeIASGaFlTKLPsLQILDLSfNFQYKEYLQFINI	354
hTLR8.pep	rKinAAwfKnMPHLkvLDLeFNyLvGEIASGaFlTMlPrLeILDLSfNYIKGSYPQHINI	359
	. : . : . : . : . : . :	420
mTLR8.pep	SsnfSKLRslKkLHLRGyVFRELkkKHFEHLQSLPNLATInLGInfiEKIDfkAfQnfSk	414
hTLR8.pep	sRnfsKLLsLRALHLRGyVFQELRedDFQPlMQLPNLStInLGInfiKQIDfkLfQnfSN	419

	mTLR8 . pep	LDVIYLSGNRIASVLDGT--DY---SSWRNRLRKPLSTDDDEFDPHVNFYHSTKPLIKPQ	469
	hTLR8 . pep	LEIIYLSENRIISPLVKDTRQSYANSSSFQRHIRKRRTDF-EFDPHSNFYHFTRPLIKPQ	478
5		. : . : . : . : . : . :	540
	mTLR8 . pep	CTAYGKALDLSLNNIFIIGKSQFEGFQDIACLNLSFNANTQVFNGTEFSSMPHIKYLDLT	529
	hTLR8 . pep	CAAYGKALDLSLNSIFFIGPNQFENLPDIACLNLSANSNAQVLSGTEFSAPHVKYLDLT	538
		. : . : . : . : . : . :	600
10	mTLR8 . pep	NNRLDFDDNNAFSDLHDLEVLDLSHNAHYFSIAGVTHRLGFIQNLIINLRVLNLSHNGIYT	589
	hTLR8 . pep	NNRLDFDNASALTELSDELVDLSYNSHYFRIAGVTHHLEFIQNFTNLKVLNLSHNNIYT	598
		. : . : . : . : . : . :	660
	mTLR8 . pep	LTEESELKSISLKELVFGSNGRLDHLWNANDGKYWSIFKSLQNLIIRDLDSYNNLQQIPNGA	649
15	hTLR8 . pep	LTDKYNLESKSLVELVFGSNGRLDILWNDNDRYISIFKGLKNLTRDLDSLNRLLKHIPNEA	658
		. : . : . : . : . : . :	720
	mTLR8 . pep	FLNLPQSLQELLISGNKLRFNWTTLLQYFPHLHLLDLSRNELYFLPNCLSKFAHSLETLL	709
20	hTLR8 . pep	FLNLPASLTEHLINDNMLKFFNWTTLLQQFPRELLDLRGNKLLFLTDSLSDFTSSLRTLL	718
		. : . : . : . : . : . :	780
	bf135656 . pep	NHFSHLPSGFLSEARNLVHLDLSFNTIKMINKSSLQTKMKTNLSILELHGNYFDCTC	57
	mTLR8 . pep	LSHNHFSHLPSGFLSEARNLVHLDLSFNTIKMINKSSLQTKMKTNLSILELHGNYFDCTC	769
	hTLR8 . pep	LSHNRIHSLPSGFLSEVSLLKHLDDLSSNLLKTINKSALETKT'TT'KLSMLELHGNNPFECTC	778
25		. : . : . : . : . : . :	840
	bf135656 . pep	DISDFRSWLDENLNITIPKLVNVICSNPGDQKSXSIMSLDLTTCVSDTTAAVLFFLTFLT	117
	mTLR8 . pep	DISDFRSWLDENLNITIPKLVNVICSNPGDQKSXSIMSLDLTTCVSDTTAAVLFFLTFLT	829
	hTLR8 . pep	DIGDFRRWMDEHLNVKI'PRLVDVICASPGRQKGSI'VSELELTTCVSDVTAVILFFF'TFF'I	838
30		. : . : . : . : . : . :	900
	bf135656 . pep	TSMVMLAALVHHLFYWDVWFIIYHMCSAKLKGYSSTSQTIFYDAYISYDTKDASVTDWVI	177
	mTLR8 . pep	TSMVMLAALVHHLFYWDVWFIIYHMCSAKLKGYSSTSQTIFYDAYISYDTKDASVTDWVI	889
	hTLR8 . pep	TTMVMLAALAHHLFYWDVWFIIYNVCLAKVGYSRLSTSQTIFYDAYISYDTKDASVTDWVI	898
35		. : . : . : . : . : . :	960
	bf135656 . pep	NELRYHLE	185
	mTLR8 . pep	NELRYHLEESDKSVLLCLEERDWDPLPIIDNLMQSINQSKKTI'FVLTKKYAKSWNFKT	949
	hTLR8 . pep	NELRYHLEESRDKNVLLCLEERDWDPLAIIDNLMQSINQSKKTV'FVLTKKYAKSWNFKT	958
40		. : . : . : . : . : . :	1020
	mTLR8 . pep	AFYLALQRLMDENMDVIIIFILLEPVLYSQYLRLRQRICKSSILQWPNNPKAENLFWQSL	1009
	hTLR8 . pep	AFYLALQRLMDENMDVIIIFILLEPVLYHQSYLRLRQRICKSSILQWPDNPKEGLFWQTL	1018
45		. : . : . : . : . : . :	1080
	mTLR8 . pep	KNVVLTENDSRYYDDLYIDSIRQY	1032

hTLR8 . pep

RNVVLTENDSRYNMYVDSIKQY

1041

In Table 17 the sequences are assigned as follows: mTLR8.pep, SEQ ID NO:192;
hTLR8.pep, SEQ ID NO:184; and bfl35656.pep, SEQ ID NO:193.

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Example 20. Transient transfectants expressing TLR8 and TLR7

The cloned human TLR7 and human TLR8 cDNA (our result) were cloned into the expression vector pCDNA3.1(-) from Invitrogen using the NotI site. Utilizing a “gain of function” assay, hTLR7 and hTLR8 expression vectors were transiently expressed in human
293 fibroblasts (ATCC, CRL-1573) using the calcium phosphate method. Activation was monitored by IL-8 production after stimulus with CpG-ODN (2006 or 1668, 2μM) or LPS (100 ng/ml). None of the stimuli used activated 293 cells transfected with either hTLR7 or hTLR8.

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Example 21. Screening for TLR9, 8 and 7 modulators

Human TLR receptors 9, 8 and 7 are expressed differentially among tissues which may be due to differences in promoter structure. Du X et al., *Eur Cytokine Netw* 11:362-71 (2000); Chuang TH et al., *Eur Cytokine Netw* 11:372-8 (2000). For the human Toll-like receptors 9, 8 and 7 the genomic locus has been defined and sequenced. TLR9 is located on chromosome 3 (GenBank accession numbers NT_005985, AC006252), TLR7 on chromosome X (GenBank accession numbers NT_011774, AC005859, AC003046) and TLR8 close to TLR7 also on chromosome X (GenBank accession numbers NT_011774, AC005859). To verify differences in the promoter regions the putative promoter region of each gene are cloned in reporter vectors like pGL2-Basic (Promega, Madison, WI, USA) which contains the luciferase gene (luc) adjacent to a multiple cloning site. After transient transfection of these constructs in various cell lines, different stimuli can be tested for the activation of the inserted promoter region which is detected by luciferase activity. The promoter regions defined by the cloning of mTLR9, mTLR8 and mTLR7 can be utilized in the same manner. Definition of compounds that agonize or antagonize TLR9, 8, or 7 expression can be used to enhance or dampen responses to nucleic acid ligands or to any TLR9, 8 or 7 ligand defined by screening. These constructs can be adapted to high throughput screening after stable transfection similar to the use of TLR9 stable transfectants.

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Each of the foregoing patents, patent applications and references is hereby incorporated by reference. While the invention has been described with respect to certain embodiments, it should be appreciated that many modifications and changes may be made by those of ordinary skill in the art without departing from the spirit of the invention. It is intended that such modification, changes and equivalents fall within the scope of the following claims.

Example 22. Method cloning the murine and human extracellular TLR9 domain fused to human IgG1 Fc

Human IgG1 Fc was amplified from human B cell cDNA using the sense and antisense primers 5' TATGGATCCTCTTGTGACAAACTCACACATGC (SEQ ID NO:216) and 5' ATA AAGCTTTCATTTACCCGGAGACAGGGAGAG (SEQ ID NO:217) and ligated into pCDNA3.1(-) (Invitrogen) after digestion with the restriction endonucleases BamHI and HindIII creating the vector pcDNA-IgGFc. The extracellular domain of human TLR9 (amino acids 1 to 815) was amplified with the sense and antisense primers 5' TATGAATTCCCACCATGGGTTTCTGCCGCAG (SEQ ID NO:218) and 5' ATAGGATCCCCGGGGCACCAGGCCGCCGCCGCCGCCGCCGGAGAGGGCCTCAT CCAGGC (SEQ ID NO:219). The primers amplify the extracellular domain of human TLR9 and create adjacent to amino acid 815 an additional NotI restriction site, a glycine linker and thrombin protease recognition site. The translated sequence of this region starting at amino acid 812 is DEALSGGRGGGLVPRGS (SEQ ID NO:220). The fragment was cut with EcoRI and BamHI and cloned into pcDNA-IgGFc, creating the vector coding for the fusion protein of the extracellular domain of human TLR9 fused to the Fc part of human IgG1 (pcDNAhTLR9IgGFc). Expressed extracellular TLR9 protein can be separated from the IgG1 Fc fragment by digestion with Thrombin (see figure).

The extracellular part of murine TLR9 (amino acids 1 to 816) was amplified with the sense and antisense primers 5' TATATGCGGCCGCCACCATGGTTCTCCGTCGAAG (SEQ ID NO:221) and 5' TATATGCGGCCGCCAGAGAGGACCTCATCCAGGC (SEQ ID NO:222) and cloned into pcDNAhTLR9IgGFc after NotI digestion of PCR fragment and vector. This procedure exchanged the human extracellular part of TLR9 with the murine counterpart.

Example 23. Method of expression and purification of the extracellular domain of TLR9 fused to human IgG1 Fc

Vector DNA coding for the human or murine TLR9 human IgGFc fusion protein was transfected by Ca_2PO_4 method into 293 fibroblast cells. Transfected cells were selected with 0.7 mg/ml G418 and cloned. Expression of fusion protein was monitored by enzyme-linked immunosorbent assay (ELISA). Cells were lysed in lysis buffer (PBS, 1% Triton X-100) and supernatant was applied to ELISA plates coated with polyclonal antibody against human IgG-Fc. Bound fusion protein was detected by incubation with biotinylated polyclonal antibodies against human IgG-Fc and streptavidin-horseradish peroxidase conjugate.

For purification of the fusion protein cell lysates from 10^9 cells were produced and incubated with Protein A sepharose which binds tightly to human IgG-Fc. Incubation with the protease thrombin releases the soluble extracellular domain of human TLR9. **Figure 27** shows an example of the TLR9 fusion protein visualized by a silver stained SDS-gel. **Figure 27** demonstrates that lysates of transfected cells included a strong band travelling between 100 and 150 kD which was not present either in lysates of mock-transfected cells or in supernatants transfected or mock-transfected cells. The apparent molecular weight of the band decreased following thrombin treatment, consistent with cleavage at the thrombin protease recognition site interposed between the extracellular TLR9 domain and the Fc fragment.

Example 24. Method of cloning the murine and human extracellular TLR7 and TLR8 domain fused to human IgG1 Fc and its expression in 293 cells

The extracellular domains of murine TLR7 (amino acids 1 to 837), human TLR7 (amino acids 1 to 836), murine TLR8 (amino acids 1 to 816) and human TLR8 (amino acids 1 to 825) were amplified with the primer pairs

5' TATATGCGGCCGCCCACCATGGTGTTCGATGTGGACACG (SEQ ID NO:223) and 5' TATATGCGGCCGCCATCTAACTCACACGTATACAGATC (SEQ ID NO:224);
5' TATATGCGGCCGCCCACCATGGTGTTCCTCAATGTGGACACTG (SEQ ID NO:225) and 5' TATATGCGGCCGCCATCTAACTCACAGGTGTACAGATC (SEQ ID NO:226);
5' TATATGCGGCCGCCCACCATGGAAAACATGCCCCCTCAG (SEQ ID NO:227) and

5' TATATGCGGCCCGCCATCCGATACACAAGTCGTGAGATC (SEQ ID NO:228); and
5' TATATGCGGCCCGCCCACCATGGAAAACATGTTTCCTTCAGTC (SEQ ID NO:229)
and 5' TATATGCGGCCCGCCATCTGAAACACAAGTTGTTAGCTC (SEQ ID NO:230),
respectively. Fragments were cloned into pcDNA-IgGFc after NotI digestion.

5 Vector DNA coding for the extracellular domain of human or murine TLR7 or TLR8
fused to human IgGFc fusion protein was transfected by Ca_2PO_4 method into 293 fibroblast
cells. Transfected cells were selected with 0.7 mg/ml G418 and cloned. Expression of fusion
protein was monitored by ELISA. Cells were lysed in lysis buffer (PBS, 1% Triton X-100)
and supernatant was applied to ELISA plates coated with polyclonal antibody against human
10 IgG-Fc. Bound fusion protein was detected by incubation with biotinylated polyclonal
antibodies against human IgG-Fc and Streptavidin-horseradish peroxidase conjugate.

**Example 25. Method of antibody production against murine and human TLR9 and
characterization of activity**

15 C57/B6 mice were immunized three times by intraperitoneal administration of 20 μg
of the extracellular domain of human TLR9 mixed with 10 nmol of the CpG-ODN 1668.
B cells taken from immunized mice were fused with a non antibody producing B-cell
hybridoma P3XAG8 using standard protocols. Hybridoma supernatants were screened for
reactivity in ELISA using murine and human TLR9 fusion proteins. For identification of
20 positive hybridomas ELISA plates were coated with polyclonal antibody against human IgG-
Fc and incubated with lysate containing murine or human TLR9 IgG-Fc fusion protein.
Plates were then incubated with individual hybridoma supernatants, and bound TLR9-specific
antibodies were detected by incubation with biotinylated polyclonal antibodies against murine
IgG and Streptavidin-horseradish peroxidase conjugate.

25 Ten antibodies have been isolated which are of IgG1, IgG2a and IgG2b isotype. They
have been tested for reactivity against human and murine TLR9 and their performance in
western blotting or intracellular staining. Table 18 shows the names (ID), isotypes, reactivity
and performance in western blotting and intracellular staining.

All isolated antibodies were readily purified using standard protein A affinity
30 chromatography.

Table 18. Monoclonal Antibodies Raised Against Murine and Human TLR9

#	ID	Isotype	Reactivity in ELISA		Western Blotting	Intracellular Staining
			mTLR9	hTLR9		
1	1-3A11	G1	YES	YES	YES	NO
2	1-1B1	G1	YES	YES	YES	NO
3	1-2A9	G2a	NO	YES	YES	YES
4	1-3F2	G1	YES	YES	YES	NO
5	2-1E2	G2a	NO	YES	YES	YES
6	1-5G5	G2a	YES	YES	YES	YES
7	1-2F1	G1	YES	YES	YES	NO
8	1-5F12	G2b	NO	YES	NO	NO
9	1-3C9	G2a	NO	YES	YES	YES
10	1-3F5	G2b	NO	YES	NO	NO

Example 26. Method for Intracellular Staining

Mock transfected 293 cells and human TLR9 transfected 293 cells were seeded on cover slips and cultured overnight. The following day cells were washed in PBS and fixed with 2% formalin for 10 minutes at room temperature. Cells were permeabilized with 0.2% saponin in PBS and incubated with 2 μ g/ml anti human TLR9-specific antibody 2-1E2 for 1h. After two wash steps cells were incubated with Alexis488-conjugated goat anti-mouse IgG antibody and TLR9 was visualized utilizing confocal microscopy on a Zeiss LSM510 microscope. Results indicated that cytoplasm of human TLR9 transfected 293 cells, but not mock transfected 293 cells, stained positive for human TLR9.

Example 27. Method for Western Blotting

Lysates of 293 cells transfected with murine TLR9, human TLR9 or murine TLR2 IgG1-Fc fusion protein were separated by SDS-PAGE. Proteins were transferred to a nylon membrane utilizing a BioRad semi dry blotter according to the manufacturer's protocol. The membrane was incubated with 2 μ g/ml of the human TLR9-specific antibody 2-1E2, and human TLR9 was detected by polyclonal goat anti-mouse peroxidase conjugate. Peroxidase activity was monitored with ECL reagent (Amersham) and incubation of the membrane on film (see Figure 29).

What is claimed is: